

Relative contribution of *Prevotella intermedia* and *Pseudomonas aeruginosa* to lung pathology in airways of patients with cystic fibrosis

Martina Ulrich,¹ Isabelle Beer,¹ Peter Braitmaier,¹ Michaela Dierkes,¹ Florian Kummer,¹ Bernhard Krismer,¹ Ulrike Schumacher,¹ Ute Gräpler-Mainka,² Joachim Riethmüller,² Peter Ø Jensen,³ Thomas Bjarnsholt,³ Niels Høiby,³ Gabriel Bellon,⁴ Gerd Döring¹

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¹Institute of Medical Microbiology and Hygiene, Universitätsklinikum Tübingen, Tübingen, Germany

²Kinderklinik, Universitätsklinikum Tübingen, Tübingen, Germany

³Department of Clinical Microbiology, Rigshospitalet, Copenhagen, Denmark

⁴Groupement Hospitalier Est, Hôpital Femme Mère Enfant, Bron, France

Correspondence to

Gerd Döring, Institute of Medical Microbiology and Hygiene, Universitätsklinikum Tübingen, Wilhelmstrasse 31, D-72074 Tübingen, Germany; gerd.doering@med.uni-tuebingen.de

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ABSTRACT

Background Patients with cystic fibrosis (CF) with *Pseudomonas aeruginosa* lung infections produce endobronchial mucus plugs allowing growth of obligate anaerobes including *Prevotella* spp. Whether obligate anaerobes contribute to the pathophysiology of CF lung disease is unknown.

Methods The virulence of *Prevotella intermedia* and *Ps aeruginosa* was investigated in vitro and in mice, antibodies against *P intermedia* in CF sera were assessed and a culture-independent detection method for *P intermedia/P nigrescens* in CF sputum was tested.

Results *P intermedia* reached cell numbers of $>10^5$ – $>10^7$ colony-forming units (CFU)/ml sputum. The majority of patients with CF (16/17; 94.1%) produced antibodies against two immunoreactive antigens of *P intermedia*. Culture supernatant fluids, collected from 10^9 *P intermedia* cells, were more cytotoxic to respiratory epithelial cells in vitro and inflammatory in mouse lungs than respective fluids from anaerobically grown *Ps aeruginosa*, while fluids from aerobically grown *Ps aeruginosa* had the highest cytotoxicity and inflammation. Both pathological effects were largely reduced when culture supernatant fluids from 10^7 cells of either species were used. *P intermedia* cells ($\sim 10^6$ CFU/lung) did not induce mortality in the agar beads lung infection mouse model, while *Ps aeruginosa* cells caused death in 30% of mice due to rapid multiplication. A *P intermedia/P nigrescens*-specific PNA probe was significantly more sensitive than culture-dependent diagnostic assays to detect these strict anaerobes.

Conclusion *Ps aeruginosa* and *P intermedia* become significantly virulent in vitro and in vivo when cell numbers exceed 10^8 CFU/lung.

INTRODUCTION

In patients with cystic fibrosis (CF), chloride retention as a consequence of defective CF transmembrane conductance regulator and increased sodium and water absorption causes an abnormally high viscosity of the mucus layer on the respiratory epithelium.^{1,2} The resulting impaired mucociliary clearance initiates a vicious circle of chronic infection and inflammation in the airways of patients with CF.³ Epidemiological data from a large patient registry have shown that the facultative anaerobic bacteria *Staphylococcus aureus* and *Pseudomonas*

aeruginosa are the dominating pathogens in CF lung disease.⁴ Both pathogens form biofilms under hypoxic conditions,^{5–8} thereby resisting host defence and antibiotic therapy.^{9,10} The persistence of facultative anaerobic bacteria results in a continuous influx of neutrophils into the airway lumen³ and, particularly in the absence of effective antibiotic therapy, large hypoxic mucus plugs are formed which lead to considerable airway obstruction.

Rapid consumption of oxygen by facultative anaerobic pathogens⁸ and by the large number of luminal neutrophils¹¹ may favour substantial growth of obligate anaerobes. With the exception of two early reports,^{12,13} obligate anaerobes in CF have only recently gained increased interest. Many obligate anaerobe species have been detected in CF sputum samples.^{14–20} High cell numbers of these anaerobes have ruled out the notion that the presence of these microorganisms in sputum is derived from upper airway contamination.^{16,17} The oral cavity has been regarded as a ‘stepping stone’ for lung colonisation and infection for certain bacterial anaerobic species.¹⁶

However, the important question—whether obligate anaerobes contribute to the pathophysiology of lung disease in CF—is still unresolved. In addition, it remains to be investigated whether widely used culture-dependent identification methods for obligate anaerobes correctly reflect the presence of obligate anaerobes in CF sputum.

We have addressed these issues by focusing on *Prevotella intermedia*. This Gram-negative black-pigmented anaerobic rod has been repeatedly detected in CF airway specimens.^{17–20} We comparatively investigated the virulence of *P intermedia* and *Ps aeruginosa* in vitro and in mice, assessed antibodies against *P intermedia* in CF serum and tested a culture-independent detection method for *P intermedia*, *P nigrescens* and *P melaninigenica* in CF sputum.

METHODS

Patients

Serum and sputum samples and lung function data were collected from 17 patients with CF attending the CF centre of the University Children’s Clinic, Universitätsklinikum Tübingen, Tübingen, Germany. Patients were eligible if they were infected with *Ps aeruginosa* and were sputum producers. To obtain a representative sputum

sample, at least 2 ml of purulent sputum had to be produced by the patients during routine visits to the CF centre in Tübingen. Serum samples from 30 healthy individuals were also collected. Healthy individuals were recruited from members of staff at the University Hospital of Tübingen. In addition, four patients with CF who attended the CF centre of the Service de Pédiatrie, Centre Hospitalier Lyon-Sud, Pierre-Benite, France were studied.²¹

Identification of *Prevotella* species

Bronchopulmonary samples for bacteriological investigations including sputum, bronchial aspiration, protected catheter brushing, mucosal biopsies and bronchial lavages were processed by routine methods as described in the online supplement. *Prevotella* species were identified using API 32A, a fluorescein-labelled 15-mer PNA probe by in situ hybridisation, MALDI-TOF-MS and sequencing (for details see online supplement).^{22–23}

Antibody response to *P. intermedia* antigens in CF serum

Serum samples from patients with CF were subjected to western blotting and ELISA using isolated membrane proteins of the *P. intermedia* strain ATCC 25611 (for details see online supplement).²⁴

Cytotoxicity assays

Culture supernatant fluids from *Ps. aeruginosa* strain PAO1 grown under aerobic and anaerobic conditions and *P. intermedia* strain ATCC 25611 grown under anaerobic conditions were incubated with the human alveolar type II cell line A549 for 1 h at 37°C. Cytotoxicity was assessed using the Syto13/propidium iodide viability test.²⁵ Additionally, the culture supernatant fluids were incubated for various time periods with purified human polymorphonuclear leucocytes.²⁶ Finally, the ability of the *P. intermedia* strain ATCC 25611 to degrade casein, elastin or gelatin was assessed on agar plates with the respective substrates (for details see online supplement).

Animal studies

For lung infection of C57Bl/6 mice with *Ps. aeruginosa* or *P. intermedia* the agar bead model was used.²⁷ Twenty-four hours after challenge, lungs were excised and bacterial colony-forming units (CFUs) were determined using routine methods. Mice were also challenged with sterile culture supernatant fluids from the microorganisms. In both experiments, neutrophils and macrophages were quantified in lung tissue sections. All animal experiments were approved in advance by the Regierungspräsidium Tübingen, Germany (for details see online supplement).

Statistical analysis

For statistical evaluations the Spearman correlation coefficient ρ , Wilcoxon/Kruskal–Wallis test, χ^2 test and Fisher exact test were used (for details see online supplement).

RESULTS

Prevotella intermedia was selected as the prototype organism for these investigations because it has been identified as one of the most frequent obligate anaerobe in CF sputum cultures by us and others. In our study, 38% of the patients harboured *P. intermedia*.

Culture supernatant fluids of *P. intermedia* are cytotoxic to airway epithelial cells and neutrophils

Microscopic evidence showed that undiluted culture supernatant fluids of *P. intermedia* corresponding to 7.8×10^8 CFU caused

cell damage in the A549 monolayer including cell rounding after 1 h of incubation followed by complete cell detachment (figure 1A, B). Cell death was visible in 47.5% of A549 cells after incubation for 1 h at 37°C when examined using a live/dead fluorescent staining method. When the culture supernatant fluids of *P. intermedia* had been diluted 1:10 and 1:100, the cell death rate decreased to 34.8% and 10.3% (figure 1A, B). Because *P. intermedia* strains produce various proteases,^{28–29} we tested the culture supernatant fluid of the *P. intermedia* strain ATCC 25611 for degradation of gelatin, elastin and casein. In contrast to *Ps. aeruginosa* strain PAO1 grown anaerobically or aerobically, undiluted culture supernatant fluids of *P. intermedia* at comparable cell numbers did not have activity against these substrates (data not shown).

Culture supernatant fluids from *Ps. aeruginosa* grown anaerobically corresponding to 7.8×10^8 CFU had a low cytotoxic effect (figure 1A, B). Culture supernatant fluids from aerobically grown *Ps. aeruginosa* were significantly more cytotoxic ($p=1.53 \times 10^{-9}$ – 2.21×10^{-12} , χ^2 test/Fisher exact test; figure 1A, B). A 1:100 dilution produced cytotoxicity of 16.3% towards A549 cells. A similar result was obtained when the culture supernatant fluids were collected when the culture was grown aerobically to a cell number of 7.2×10^8 CFU/ml (cytotoxicity 13.5%). This cytotoxicity is significantly lower than undiluted culture supernatant fluids of *P. intermedia* ($p=8.08 \times 10^{-40}$ – 1.35×10^{-42}). Undiluted culture supernatant fluids from anaerobically grown *Ps. aeruginosa* were significantly less cytotoxic than those from *P. intermedia* ($p=9.08 \times 10^{-40}$ – 1.44×10^{-52}).

When purified human neutrophils were incubated with undiluted culture supernatant fluids from *P. intermedia*, flow cytometry showed an increasing percentage of necrotic neutrophils with time, reaching ~50% dead cells after 60 min of incubation, corroborating the results obtained with A549 cells (figure 1C). Undiluted *Ps. aeruginosa* culture supernatant fluids from strain PAO1 grown anaerobically were significantly less cytotoxic than culture supernatant fluids from *P. intermedia* ($p=0.003$) and killed only 22% of the cells (figure 1C). In contrast, culture supernatant fluids from aerobically grown *Ps. aeruginosa* were highly cytotoxic and killed 99% of neutrophils within 1 min of incubation (figure 1C). In further experiments the inflammatory potential of culture supernatant fluids of *P. intermedia* and *Ps. aeruginosa* were compared in mouse lungs (figure 1D, E). *P. intermedia* and *Ps. aeruginosa* caused a significantly higher influx of neutrophils ($p<0.025$) (figure 1D) and macrophages ($p<0.025$) (figure 1E) than anaerobically grown *Ps. aeruginosa*.

Collectively, these in vitro and animal data show that the cytotoxicity of *P. intermedia* and anaerobically grown *Ps. aeruginosa* is significantly different. They further reveal that 1:100 diluted culture supernatant fluids from both pathogens, equivalent to cell numbers of $\sim 10^7$ CFU and irrespective of growth conditions, are markedly reduced.

Virulence of *P. intermedia* in mouse lungs is dependent on high growth rates

When groups of mice were challenged with 3×10^6 CFU *P. intermedia* embedded in agar beads, bacterial CFUs remained stable after 24 h (figure 2A). None of the mice died 48 h after infection. In contrast, agar bead-embedded *Ps. aeruginosa* grew 1.1×10^6 – 1.5×10^{10} CFU within 24 h after challenge (figure 2A) and killed 4/12 (30%) of the mice after 48 h. Similar to their differential capacity to induce mortality in mice, *P. intermedia* and *Ps. aeruginosa* cells differed in their inflammatory potential in mouse lungs (figure 2B, C). Washed *P. intermedia* cells caused less

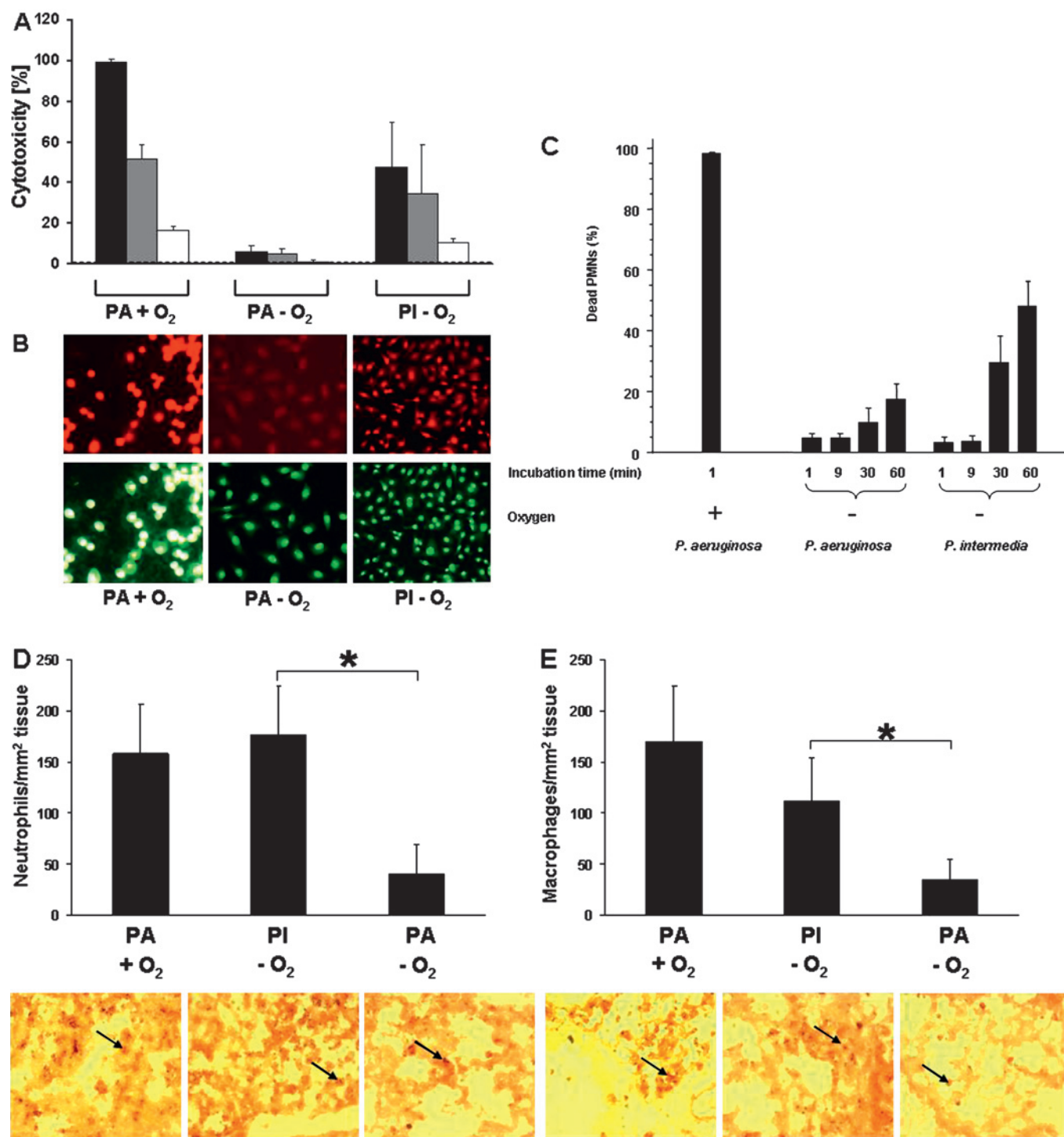


Figure 1 *Prevotella intermedia* produces more extracellular toxins than *Pseudomonas aeruginosa* under anaerobic conditions. (A, B) Cytotoxic effects of culture supernatant fluids (black bars) and diluted culture supernatant fluids (1:10, grey bars and 1:100, white bars) of *P. intermedia* (PI) grown anaerobically (-O₂) and *Ps. aeruginosa* (PA) grown anaerobically (-O₂) or aerobically (+O₂) on the respiratory epithelial cell line A549 determined using propidium iodide (dead cells: red) and syto13 (all cells: green). The 1:100 diluted culture supernatant fluid of *Ps. aeruginosa* grown aerobically corresponds to a cell number of 8×10^8 CFU/ml and is comparable to the undiluted culture supernatant fluids of anaerobically grown *Ps. aeruginosa* and *P. intermedia* (7.8×10^8 CFU/ml each). Data were obtained from 4 or 5 independent experiments which were performed in triplicate. (C) Cytotoxic effects of culture supernatant fluids of PI or PA on neutrophils (PMN) from five healthy human subjects expressed as the percentage of live neutrophils. Data were obtained from three independent experiments performed in triplicate. (D, E) Neutrophil and macrophage numbers in lung tissue of six C57Bl/6 mice and the representative pictures of lung tissue sections from C57Bl/6 mice intratracheally challenged with culture supernatant fluids of PI or PA. Lung tissue sections were stained with specific antibodies against mouse neutrophils (D) or macrophages (E). Arrows depict positive staining. Original magnification $\times 100$. * $p < 0.025$.

influx of neutrophils (figure 2B) and macrophages (figure 2C) than *Ps. aeruginosa* cells within 24 h of the challenge. Our in vitro data showed that *P. intermedia* can grow outside agar beads when

anaerobic growth conditions prevail and reach cell numbers of $2.1 \times 10^8 \pm 9.5 \times 10^7$ CFU/ml while, under aerobic growth conditions, cell numbers remained at $3.7 \times 10^6 \pm 4.9 \times 10^5$ CFU/ml.

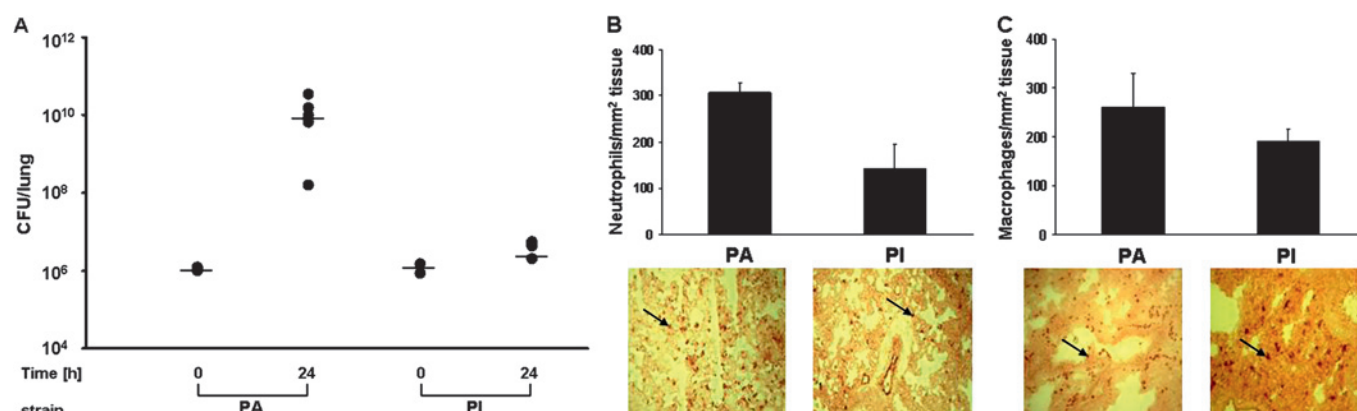


Figure 2 *Prevotella intermedia* cells are less virulent than *Pseudomonas aeruginosa* cells grown under anaerobic conditions in vivo. (A) Mice were challenged with *Ps aeruginosa* (PA) or *P intermedia* (PI) embedded in anaerobic agar beads and bacterial cell numbers (CFU) were determined in lung tissues at time 0 and 24 h. (B) Neutrophil and (C) macrophage numbers in lungs of mice challenged with PA or PI for 24 h determined with immunohistochemistry. Arrows depict positive staining. Original magnification $\times 100$.

These data suggest that *P intermedia* contributes to lung pathology if appropriate growth conditions are present which allow multiplication to high cell numbers ($>10^8$ CFU/lung). This suggestion needs to be validated in further studies.

P intermedia is recognised by specific serum antibodies in patients with CF

To investigate whether the *P intermedia* in CF sputum samples is recognised by the adaptive immune system of patients, as has been investigated for *Ps aeruginosa*,^{30,31} we determined antibody titres against *P intermedia* in serum samples from patients with CF and healthy individuals using a newly developed ELISA (figure 3A). Cohorts comprised 17 patients with CF (median age 15 years) and 30 healthy individuals (median age 25 years). A cut-off at a reciprocal titre of 510 in the ELISA was determined as three times the unspecific binding value of a group of healthy human individuals. Patients with CF harboured positive antibody titres against *P intermedia* antigens in 94.1% of serum samples while none of the healthy control serum samples was positive. Thus, IgG antibody titres were significantly increased in patients with CF compared with healthy individuals ($p<0.001$).

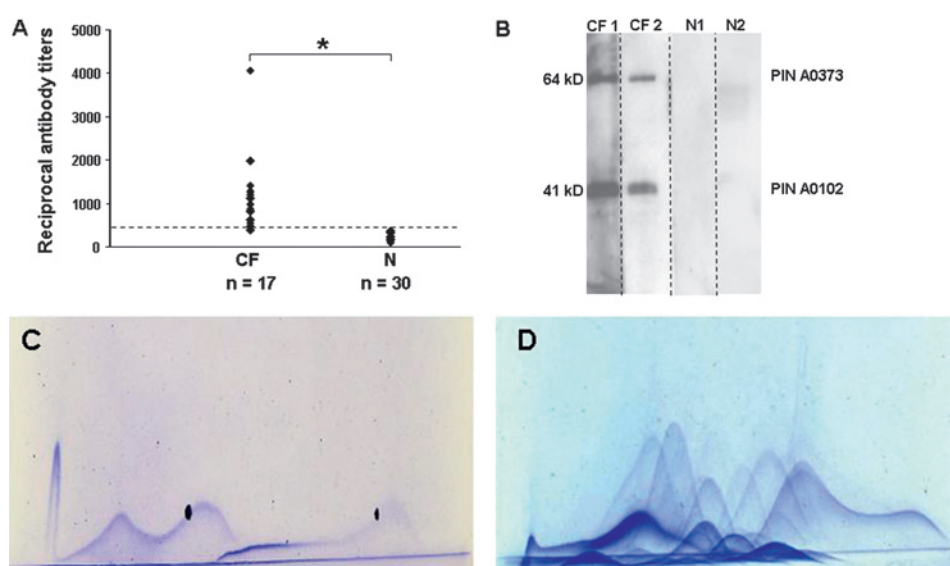
Western blotting and MALDI-TOF showed that serum antibodies from patients with CF but not antibodies from healthy

individuals bind to a 64 kD protein (PIN A0373) with hypothetical function and to a 42 kD immunoreactive protein (PIN A0102) (figure 3B). In order to prevent unspecific Fc-mediated binding of the antibodies to PIN A0373 and PIN A0102, we blocked the membrane with 17 μ g/ml of the Fc fragment of human IgG. The western blot results were corroborated when crossed immunoelectrophoresis was used to determine the number of different antibodies in serum samples from our CF population. In two high-titre serum samples only two antibody arcs against single *P intermedia* cell antigens were visible (figure 3C); in contrast, *Ps aeruginosa* provoked the production of up to 64 different antibodies during chronic infection in patients with CF (figure 3D). A weak non-correlation ($\rho=-0.253$, $p>0.001$) was obtained between antibody titres against *P intermedia* cell proteins and lung function in patients with CF. The data suggest that *P intermedia* is recognised by the humoral immune response in patients with CF.

Numbers of strict anaerobic bacteria in CF bronchopulmonary samples

Tunney and colleagues have determined the numbers of anaerobes in sputum specimens from patients with CF.¹⁷ They reported total viable counts of *Prevotella* species of $>10^4$ – 10^7 CFU/g

Figure 3 Immune recognition of *Prevotella intermedia* in patients with cystic fibrosis (CF). (A) ELISA: IgG antibody titres against *P intermedia* cell proteins in 17 CF and 30 control (N) serum samples. Reciprocal cut-off titre 510 ($*p<0.001$). (B) Western blot: IgG antibodies in serum from two patients with CF (CF1, CF2) but not from two healthy individuals (N1, N2) reacted with two membrane proteins of *P intermedia* identified as PIN A0373 and PIN A0102. (C, D) Crossed immunoelectrophoresis: serum from (C) a patient with CF with a high antibody titre against *P intermedia* cell wall proteins (1:4682) and (D) a patient with CF with chronic *Pseudomonas aeruginosa* infection were subjected to crossed immunoelectrophoresis using the respective *P intermedia* or *Ps aeruginosa* cell proteins as antigens.



sputum. To exclude the possibility that sputum specimens are contaminated by *Prevotella* species from oral flora, we quantified these micro-organisms in bronchopulmonary samples collected by fiberoptic bronchoscopy. When bronchopulmonary samples from four patients with CF (median age 11.5 years) were cultured anaerobically on media we found different *Prevotella* species in numbers of $>10^5$ – $>10^7$ CFU, corroborating the results by Tunney *et al*¹⁷ and suggesting that bacterial growth had occurred within the lower airways rather than contamination of airway samples with the oral anaerobic flora during recovery (table 2).

Culture-independent diagnosis yielded a higher prevalence of *Prevotella* species in sputum specimens from patients with CF. Not all patients with antibodies against *P. intermedia* had *P. intermedia* recovered from sputum taken at the time the serum was obtained. While this may be due to the persistence of specific antibodies even after eradication of the pathogen, it may also result from a lack of growth of the anaerobes on culture plates (ie, a false negative diagnosis). We therefore tested a culture-independent novel PNA probe for the identification of *Prevotella* spp. According to the sequence information of the distributor, *P. intermedia* cannot be distinguished from *P. nigrescens*

by this technique. The specificity of the probe for *P. intermedia* and *P. nigrescens* was confirmed by applying the probe to a large number of other *Prevotella* strains (table 1). Four other bacterial strains were also used (table 1). Only negative staining results were obtained (data not shown). The probe was applied to 17 sputum samples from patients with CF and identified *Prevotella* spp. in 47% of the samples (figure 4A, B).

In contrast, growth of *Prevotella* spp. from sputum specimens on culture plates was only achieved in four cases (23%). To specify to which species grown colonies belong, we applied MALDI-TOF-MS. As gold standards for detection of *Prevotella* species by MALDI-TOF-MS, 27 different sequenced *Prevotella* strains stored in our large strain collection at the Institute of Medical Microbiology and Hygiene were used (table 1). As examples, distinct MALDI-TOF-MS patterns for *P. intermedia*, *P. nigrescens* and *P. melaninogenica* are depicted in figure 4C–E. Using these references, two of the four *Prevotella* sputum isolates were identified as *P. intermedia* and the other two as *P. nigrescens* and *P. melaninogenica*, respectively. These results were corroborated by the sequence analysis of the four *Prevotella* CF isolates (accession numbers: HM998853, HM998854, HM998855, HM998856).

Collectively, these data show that *Prevotella* species and possibly other obligate anaerobic species present in sputum specimens from patients with CF may be missed by culture-dependent methods.

DISCUSSION

Contribution of *P. intermedia* to the pathogenicity of lung disease in CF

In this study we show that *P. intermedia* may contribute to the complex pathophysiology in the chronically infected CF lung. Several lines of evidence support this notion. First, *Prevotella* species were detected in numbers $>10^5$ CFU, showing that bacterial growth had occurred within the lower airways rather than contamination of airway samples with the oral anaerobic flora during recovery (table 2). This result corroborates data from other investigators.¹⁷

Second, in contrast to healthy individuals, *P. intermedia* antigens are recognised by the adaptive immune system of the vast majority of patients with CF resulting in specific antibody titres, which again argues against contamination of airway samples with the oral anaerobic flora during recovery (figure 3A). The fact that, in some patients, positive serum antibody titres against *P. intermedia* antigens were found in the absence of the microorganism may be explained by either a false negative detection of *P. intermedia* (see below) or the possibility that patients who cleared the microorganism still had circulating antibodies. Similar phenomena occur with *Ps. aeruginosa* serum antibody titres and detection of the microorganism in CF airway specimens.^{30, 31}

Third, extracellular toxins of *P. intermedia* are cytotoxic for human alveolar type II cells and neutrophils (figure 1A, B, C), and they induce the influx of macrophages and neutrophils in the airway lumen thereby increasing lung inflammation (figure 1D, E). Using assays for gelatin, elastin and casein cleavage, we were unable to identify proteases with these activities in culture supernatant fluids of *P. intermedia* strain ATCC 25611 which had pronounced cytotoxicity to A549 cells.

Last, *P. intermedia* possesses a surface protein³² which, in analogy to *S. aureus*,³³ may avoid opsonophagocytosis by binding the Fc portion of human IgG, rendering specific antibodies inefficiently protective (figure 3B).

Importantly, the cytotoxic and inflammatory potential of culture supernatant fluids of *Ps. aeruginosa* grown anaerobically

Table 1 Strains used in the study

Microorganism	Strain	Source
<i>Prevotella intermedia</i>	ATCC 25611	A
	ATCC 20706	A
<i>Prevotella oralis</i>	ATCC 20702	
<i>Prevotella paludivivens</i>	ATCC 17968	
<i>Prevotella nigrescens</i>	ATCC 13386	
<i>Bacteroides fragilis</i>	ATCC 25285	
<i>Prevotella nigrescens</i>	138	B
	582	B
	621	B
	634	B
	655	B
	658	B
	672	B
	1006	B
	1024	B
	1032	B
	1044	B
	1048	B
<i>Prevotella oralis</i>	1057	B
	1243	B
	1037	B
<i>Prevotella oris</i>	651	B
	1033	B
<i>Prevotella baroniae</i>	1050	B
	1071	B
	578	B
<i>Prevotella buccae</i>	674	B
	583	B
<i>Prevotella denticola</i>	608	B
	1059	B
<i>Prevotella disiens</i>	1074	B
	1029	B
<i>Prevotella melaninogenica</i>	1049	B
	507	B
<i>Prevotella multiformis</i>	524	B
	1045	B
<i>Prevotella intermedia</i>	R2	B
<i>Actinomyces odontolyticus</i>	PAO1	B
<i>Pseudomonas aeruginosa</i>	ATCC 35556	A
<i>Staphylococcus aureus</i>		

A, German Collection of Microorganisms and Cell Cultures (DSMZ); B, Institute for Medical Microbiology and Hygiene, Tübingen, Germany; identified by API.

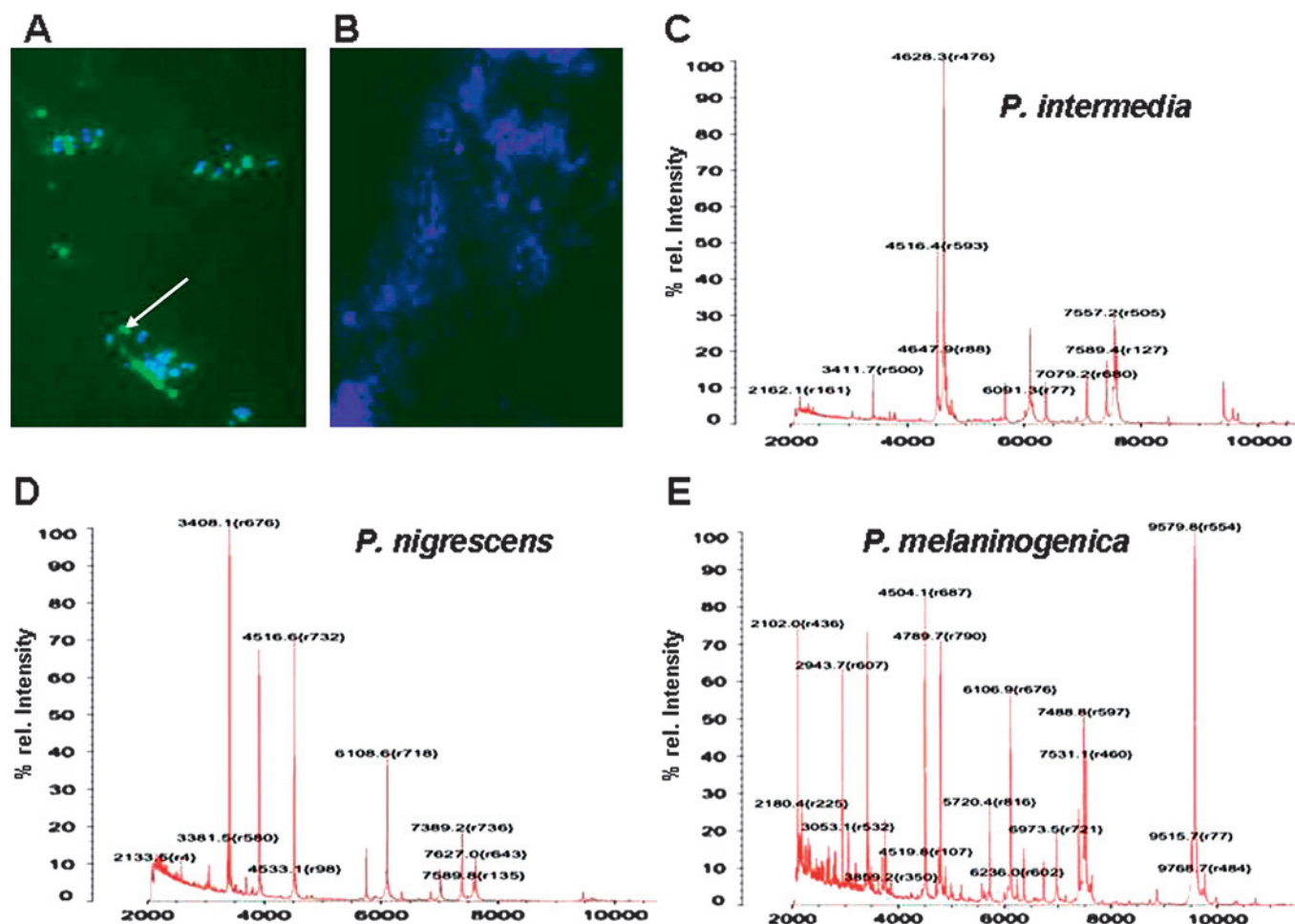


Figure 4 Detection of *Prevotella intermedia*/*Prevotella nigrescens* by a novel FITC-labelled PNA probe. *Prevotella* spp. were stained by the PNA probe in a representative sample of patients with cystic fibrosis. (A,B) DAPI staining. (C–E) MALDI-TOF-MS spectra of (C) *P. intermedia*, (D) *P. nigrescens* and (E) *P. melaninogenica*.

to cell numbers comparable to *P. intermedia* was much lower (figure 1A,D,E). Provided that comparable *P. intermedia* cell numbers are reached in CF airways to those used in our in vitro assays ($>10^5$ CFU), *P. intermedia* would be more pathogenic than anaerobically grown *Ps. aeruginosa*. However, although growth under anaerobic environmental conditions is the dominant life style for *Ps. aeruginosa* in the CF lung³⁴ and the its virulence decreases during chronic CF lung infection due to adaptive radiation,³⁵ *Ps. aeruginosa* can also multiply in aerobic compartments of the CF lung, in contrast to *P. intermedia*, and contribute to the pathophysiology of CF lung disease. Our animal experiments support this notion.

Detection of *Prevotella* species in bronchopulmonary samples of patients with CF

In general, reports on anaerobic bacterial species in CF sputum samples are rare,^{14–20} although growth of obligate anaerobic species in CF sputum specimens may be facilitated by the high viscosity of the sputum² and the presence of facultative anaerobic bacteria which rapidly consume oxygen, thus lowering the oxygen tension to degrees which allow obligate anaerobic bacteria to thrive.

In this study we show that the fluoreszenz in situ hybridisierung (FISH) method for detection of *P. intermedia* and *P. nigrescens* yields better results in this context than the classical API system. By using the novel PNA probe, 47% of the sputum

samples were found positive for *P. intermedia* and/or *P. nigrescens* while only in 23% of the sputum samples were *Prevotella* spp. identified by culture and API. One explanation is that we may have missed *P. intermedia* during sputum solubilisation. Our finding may also explain in part why we detected *P. intermedia*-specific serum antibody titres in several *P. intermedia* culture-negative CF patients. Compared with the API system—which is known to have various disadvantages such as interpretation of biochemical reactions, long incubation time periods and costs—the novel PNA probe is much more rapid. Furthermore, FISH is more sensitive than API due to different sample processing. While the probe cannot be applied to detect these *Prevotella* spp. in unprocessed CF sputum, solubilisation with 1,4-Dithiothreitol (DTT) followed by centrifugation and fluorescence microscopy is sufficient to obtain a result within 4 h. The PNA probe may allow rapid screening of *P. intermedia* in the sputum from patients with CF which, in positive cases, may be followed

Table 2 *Prevotella* species in bronchopulmonary samples of patients with cystic fibrosis

Patient	Strains	CFU/ml
1	<i>Prevotella denticola</i>	$>10^7$
2	<i>Prevotella intermedia</i>	$>10^5$
3	<i>Prevotella melaninogenica</i>	5×10^6
4	<i>Prevotella melaninogenica</i>	5×10^6

by a more elaborate and time-consuming quantitative assessment of *P. intermedia*. These data can be used for the control of treatment directed against strict anaerobes.

Taken together, the results of this study suggest that *P. intermedia* plays a critical role in the complex pathophysiology of lung disease in patients with CF when cell numbers of $>10^9$ CFU are reached in the anaerobic sputum plugs.

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Competing interests None.

Patient consent Obtained.

Ethics approval This study was conducted with the approval of the ethic committees of the University of Tübingen, the Hospices Civils de Lyon and the Université Claude Bernard, Lyon, France.

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