

Positionally cloned asthma susceptibility gene polymorphisms and disease risk in the British 1958 Birth Cohort

J D Blakey,¹ I Sayers,¹ S M Ring,² D P Strachan,³ I P Hall¹

► Additional figures are published online only at <http://thorax.bmj.com/content/vol64/issue5>

¹ Division of Therapeutics and Molecular Medicine, Nottingham Respiratory Biomedical Research Unit, University Hospital Nottingham, Nottingham, UK;

² Department of Community Based Medicine, University of Bristol, UK; ³ Division of Community Health Sciences, St George's Hospital, University of London, UK

Correspondence to: Professor I P Hall, Division of Therapeutics and Molecular Medicine, Nottingham Respiratory Biomedical Research Unit, University Hospital Nottingham, Nottingham NG7 2UH, UK; Ian.Hall@nottingham.ac.uk

Received 20 May 2008
Accepted 24 January 2009
Published Online First
2 February 2009

ABSTRACT

Objective: The aim of this study was to estimate the contribution of polymorphisms in the positionally cloned asthma candidate genes *ADAM33*, *PHF11*, *DPP10*, *GPRA* and *PTGDR* to the risk of asthma, total and specific immunoglobulin E level, lung function and wheezing in a large, nationally representative, population.

Methods: An association analysis was undertaken using genotype data for tagging and previously associated single nucleotide polymorphisms (SNPs) in regions of these genes and longitudinal phenotype data from singletons of white ethnicity in the British 1958 Birth Cohort DNA archive (n = 7703). Population-attributable risk fractions for SNPs showing association were calculated.

Results: Polymorphisms producing small but statistically significant increases in asthma risk (OR 1.1 per allele) were identified in *DPP10* and *ADAM33*, with the strongest evidence being for SNPs tagging the *DPP10* gene. No individual SNP in any gene under study markedly increased risk for any of the phenotypes in the population studied.

Conclusions: These data suggest that *DPP10* and *ADAM33* influence asthma risk in the UK population. However, the effects driven by any given locus are small, and genotyping of multiple polymorphisms in many genes will be needed to define a full genetic profile for disease risk.

Over the past 10 years, major efforts have been made using positional cloning approaches to identify genes predicting the risk of developing asthma and related phenotypes. At least 14 genome-wide linkage studies have been undertaken in a range of populations: this has resulted in the identification of a number of potential disease risk genes including *ADAM33*, *PHF11*, *DPP10* and *GPRA*.^{1–4} A fifth gene, for the prostanoid D receptor (*PTGDR*), was identified using a combination of linkage and candidate gene approaches.⁵ The initial studies on these genes suggested potentially major contributions to disease risk. However, subsequent attempts at replication in small populations have given conflicting results (see Supplementary material). We have therefore utilised the longitudinal phenotypic data from the British 1958 Birth Cohort in order to assess the contribution of these genes to the risk of asthma and related phenotypes in the UK population. We previously used this approach in a similar candidate gene study to investigate the potential role of *ADRB2* polymorphism in asthma incidence and prognosis.⁶ The specific aim of this study was to define population-attributable risk for single nucleotide

polymorphisms (SNPs) in *ADAM33*, *PHF11*, *DPP10*, *GPRA* and *PTGDR*.

METHODS

Study population and procedures

The DNA archive established from the British 1958 Birth Cohort (also known as the National Child Development Study) was used. Details of this study population and phenotyping procedures have been reported previously.^{6–11} In brief, 18 558 individuals born during 1 week in March 1958 were followed up at ages 7, 11 and 16 years by parental interviews and examinations by school medical officers, and at ages 23, 33 and 42 years by means of interviews. The presence of diagnosed asthma or a history of wheezy bronchitis were ascertained by parental interviews in childhood. In adulthood, questionnaires were used to define asthma ever, wheezing ever and wheezing episodes in the previous 12 months. During 2002–2004 all cohort members still in contact with the study team (12 069) and who had not needed a proxy interview were invited to be examined by a trained research nurse at home: 9377 were visited and adequate blood samples for DNA extraction obtained from 8018. Of these, 7703 were singletons of white ethnicity and were included in the present study. The lifetime histories of asthma and wheezing illness for these individuals who were included in the analyses in this study are similar to those who are not included: at age 42 years, the lifetime prevalence of asthma or wheeze in the full cohort was 49.9% and in the cohort with DNA was 48.8%. Total circulating immunoglobulin E (IgE) and specific IgE to house dust mite, mixed grass pollen and cat fur were measured with a Hytec enzyme immunoassay (Hycor Biomedical, Irvine, CA, USA¹²) at age 45.

Spirometry at age 44–45 years was done in the standing position without nose clips, using a Vitalograph hand-held spirometer as described in full elsewhere.¹³ In the analysis, all readings with a best-test variation >10% were excluded. Measures of lung function were adjusted for height and sex, and values with standardised residuals >3 SD units were excluded from analyses.

Protocols for the 2002–2004 biomedical examination were approved by the South East MultiCentre Research Ethics Committee. All participants gave informed written consent to participate in genetic association studies and the present study was approved by the oversight committee for the biomedical examination of the British 1958 Birth Cohort.

Table 1 Risk of self-reported asthma or wheezing illness by age 42 and SNPs in asthma candidate genes

Gene; SNP	Allele			Homozygote allele 1 (n)	Heterozygote (n)	Homozygote allele 2 (n)	2df p Value	1df p Value	OR (per allele)	95% CI for OR
	1	2								
Ptgdrs2_441	G	A	Case	2017	1481	242	0.400	0.201	1.05	0.98 to 1.13
			Control	2036	1587	263				
Ptgdrs3_197	A	G	Case	2873	807	60	0.547	0.515	0.97	0.88 to 1.07
			Control	3012	838	51				
Ptgdrs rs17125273	G	A	Case	58	746	2925	0.364	0.710	1.02	0.92 to 1.13
			Control	46	790	3044				
DPP10 rs13011555	C	T	Case	479	1760	1495	0.044	0.014	1.09	1.02 to 1.16
			Control	456	1770	1661				
DPP10 rs13392783	G	A	Case	1403	1756	562	0.024	0.007	1.09	1.03 to 1.17
			Control	1351	1880	642				
DPP10 rs1430091	G	T	Case	2213	1291	200	0.388	0.171	0.95	0.88 to 1.02
			Control	2361	1298	195				
DPP10 rs17048359	C	T	Case	392	1615	1677	0.177	0.084	0.94	0.88 to 1.01
			Control	429	1744	1666				
DPP10 rs4241129	T	A	Case	1485	1541	382	0.607	0.471	1.03	0.96 to 1.10
			Control	1536	1595	426				
DPP10 rs48493333	C	T	Case	234	1428	2043	0.372	0.206	0.95	0.89 to 1.03
			Control	272	1500	2086				
PHF11 rs2274282	G	T	Case	2704	895	75	0.539	0.503	0.97	0.88 to 1.06
			Control	2834	930	65				
PHF11 rs6561533	G	A	Case	1162	1722	717	0.702	0.95	1.01	0.95 to 1.08
			Control	1214	1767	775				
PHF11 rs6561539	G	C	Case	89	975	2660	0.677	0.387	1.04	0.95 to 1.14
			Control	89	986	2811				
GPRA rs323917	C	G	Case	3280	414	12	0.233	0.216	1.09	0.95 to 1.24
			Control	3372	475	9				
GPRA rs323922	G	C	Case	560	1792	1367	0.117	0.040	0.93	0.88 to 1.00
			Control	632	1904	1348				
GPRA rs324377	C	A	Case	665	1859	1198	0.218	0.084	0.95	0.89 to 1.01
			Control	740	1954	1188				
GPRA rs324396	C	T	Case	1653	1685	384	0.274	0.121	0.95	0.86 to 1.01
			Control	1795	1711	379				
GPRA rs740347	G	C	Case	78	897	2758	0.110	0.830	1.01	0.92 to 1.11
			Control	58	966	2845				
ADAM33 rs511898	C	T	Case	1667	1657	392	0.179	0.181	1.05	0.98 to 1.12
			Control	1704	1696	459				
ADAM33 rs528557	C	G	Case	2088	1463	221	0.042	0.032	0.92	0.86 to 0.99
			Control	2110	1538	283				
ADAM33 rs574174	C	T	Case	2513	1100	118	0.537	0.461	1.03	0.95 to 1.12
			Control	2571	1188	118				
ADAM33 rs612709	G	A	Case	61	860	2821	0.227	0.094	0.92	0.84 to 1.01
			Control	69	957	2871				

The 2df test relates to whether prevalence varies across the three genotypes (without an assumed genetic model). The 1df test assumes a per-allele (additive) model. SNP, single nucleotide polymorphism.

SNP selection

For each of the genes examined, the SNPs for genotyping were chosen either based on previously published studies identifying putative risk alleles or, where this information was not available in the literature, by choosing tagging SNPs which covered the major haplotype blocks across each gene of interest. A total of four SNPs in *ADAM33* (based on our previous meta-analysis,¹⁴), five SNPs in *GPRA*, six SNPs in *DPP10*, three SNPs in *PHF11* and three SNPs in *PTGDR* were genotyped (table 1) following SNP selection and quality control in assays using test samples from an in-house DNA archive (see Supplementary information for full details on SNP selection and quality control).

Genotyping

Genotyping was performed using Taqman technology (Applied Biosystems, Foster City, California, USA) by Geneservice

(Cambridge, UK). Genotyping call rates for each SNP included in this study were $\geq 98\%$. A random selection of 384-well plates were re-run and concordance on genotype calling was $>99\%$. We have previously reported high levels of genotyping concordance with a different technology using separately aliquoted DNA from the central archive for rs1042713 and rs1042714 in a subset of 856 individuals.⁶ For one SNP, rs528557, a double heterozygote cluster was seen; these were combined for the present analysis. Only rs6561533 deviated from Hardy–Weinberg equilibrium (Pearson χ^2 $p = 0.02$), though examination of genotype clustering revealed no apparent problems with allele calling.

Statistical analysis

Stata version 8.0 was used for cross-tabulations and regression modelling as described in a previous publication.⁶

Table 2 Risk of atopy (positive specific immunoglobulin E) at age 45 and SNPs in asthma candidate genes

Gene; SNP	Alleles			Homozygote	Heterozygote	Homozygote	2df p Value	1df p Value	OR (per allele)	95% CI for OR
	1	2		allele 1	n	allele 2				
Ptgdrs2_441	G	A	Case	135	824	1068	0.938	0.841	1.01	0.93 to 1.10
			Control	343	2050	2710				
Ptgdrs3_197	A	G	Case	1567	426	32	0.769	0.747	1.02	0.91 to 1.14
			Control	3935	1111	74				
Ptgdrs rs17125273	G	A	Case	25	396	1604	0.610	0.332	0.94	0.84 to 1.06
			Control	73	1033	3982				
DPP10 rs13011555	C	T	Case	249	941	829	0.913	0.834	1.01	0.93 to 1.09
			Control	634	2350	2119				
DPP10 rs13392783	G	A	Case	736	977	304	0.478	0.390	1.03	0.96 to 1.11
			Control	1838	2428	827				
DPP10 rs1430091	G	T	Case	1218	697	94	0.421	0.478	1.03	0.95 to 1.13
			Control	3055	1736	276				
DPP10 rs17048359	C	T	Case	206	880	911	0.378	0.165	0.95	0.88 to 1.02
			Control	560	2270	2219				
DPP10 rs4241129	T	A	Case	803	839	203	0.574	0.552	1.03	0.95 to 1.11
			Control	2026	2092	557				
DPP10 rs48493333	C	T	Case	128	777	1098	0.842	0.629	0.98	0.90 to 1.07
			Control	343	1971	2760				
PHF11 rs2274282	G	T	Case	1457	495	40	0.675	0.474	0.96	0.87 to 1.07
			Control	3717	1233	87				
PHF11 rs6561533	G	A	Case	651	914	392	0.512	0.435	1.03	0.96 to 1.11
			Control	1571	2366	991				
PHF11 rs6561539	G	C	Case	53	508	1449	0.305	0.909	1.01	0.91 to 1.11
			Control	107	1337	3659				
GPRA rs323917	C	G	Case	1762	240	6	0.912	0.710	0.97	0.84 to 1.13
			Control	4461	593	13				
GPRA rs323922	G	C	Case	322	963	730	0.458	0.616	0.98	0.91 to 1.06
			Control	796	2518	1781				
GPRA rs324377	C	A	Case	389	986	640	0.229	0.987	1.00	0.93 to 1.08
			Control	926	2605	1562				
GPRA rs324396	C	T	Case	929	892	195	0.798	0.524	1.03	0.95 to 1.11
			Control	2304	2287	505				
GPRA rs740347	G	C	Case	43	492	1481	0.431	0.722	1.02	0.92 to 1.13
			Control	86	1265	3743				
ADAM33 rs511898	C	T	Case	871	898	237	0.290	0.139	0.94	0.87 to 1.02
			Control	2279	2261	541				
ADAM33 rs528557	C	G	Case	1100	801	141	0.396	0.242	1.05	0.97 to 1.14
			Control	2831	2008	314				
ADAM33 rs574174	C	T	Case	1357	605	59	0.850	0.608	1.03	0.93 to 1.13
			Control	3398	1542	160				
ADAM33 rs612709	G	A	Case	37	483	1512	0.841	0.781	1.01	0.91 to 1.13
			Control	83	1218	3814				

The 2df test relates to whether incidence varies across the three genotypes (without an assumed genetic model). The 1df test assumes a per-allele (additive) model. SNP, single nucleotide polymorphism.

Asthma prevalence was studied as described previously.⁶ In brief, the major outcomes were: risk of asthma or wheeze by age 42; atopy (defined as the presence of specific IgE to the allergens tested) and total IgE. Secondary analyses were done to study clinical asthma, maturity onset (age >17) asthma or wheeze, and wheezing in the absence of a diagnosis of asthma.

Haplotype analyses were performed by deriving haplotypes by the EM algorithm implemented in SNPHAP (<http://www-gene.cimr.cam.ac.uk/clayton/software/snphap.txt>). For all genes except *DPP10*, all genotyped SNPs were included and haplotypes with frequency >10% used for analyses. For *DPP10* there were only two haplotypes with a frequency >10%; therefore, we derived simpler haplotypes using the loci showing association in the initial SNP analyses. Haplotype analyses were only performed where main effects were seen with at least one SNP in the relevant gene.

The population-attributable risk fraction associated with haplotype variation within each candidate gene was estimated as $(\Sigma f(R-1)/(\Sigma f(R-1)+1))$, where f is the frequency of each haplotype and R is the haplotype per copy odds ratio (OR).

RESULTS

Primary end points

Tables 1 and 2 show the allelic frequencies of the SNPs studied for the major outcome of this study (asthma/wheeze ever; atopy). Significant associations with asthma or wheeze were seen for rs13011555 and rs13392783 (table 1), in both *DPP10* and rs528557 (*ADAM33*, also known as *S2*). A borderline significant association was also seen with rs323922 in *GPRA* but only under an additive model. However, the highest OR seen for any individual SNP was only 1.1. No significant associations were seen with atopy as measured by specific IgE (table 2).

Table 3 Risk of developing asthma or wheeze after age 17 years and risk SNPs in *DPP10*, *ADAM33* and *GPR4*

Gene; SNP	Alleles		Asthma/wheeze after age 17 years	Homozygote allele 1	Heterozygote	Homozygote allele 2	2df p Value	1df p Value	OR (per allele)	95% CI for OR
	1	2		n	n	n				
<i>DPP10</i> rs13011555	C	T	Case	173	611	509	0.180	0.066	1.09	0.99 to 1.19
			Control	832	3143	2820				
<i>DPP10</i> rs13392783	G	A	Case	508	608	174	0.007	0.002	1.15	1.05 to 1.25
			Control	2246	3028	1030				
<i>GPR4</i> rs323922	G	C	Case	198	610	480	0.439	0.290	0.95	0.88 to 1.04
			Control	994	3086	2235				
<i>ADAM33</i> rs528557	C	G	Case	728	499	79	0.532	0.262	0.95	0.84 to 1.04
			Control	4370	2502	425				

The 2df test relates to whether prevalence varies across the three genotypes (without an assumed genetic model). The 1df test assumes a per-allele (additive) model. SNP, single nucleotide polymorphism.

None of the SNPs studied showed significant associations with log total IgE despite the known higher heritability of this trait (data not shown).

Secondary end points

None of the SNPs studied predicted the incidence of asthma or wheezing from birth to age 16 (data not shown); however, rs13392783 (*DPP10*) predicted risk of the development of asthma in later life; a weaker association was also seen with this end point for rs574174 (*ADAM33* ST+7) (OR 1.09, $p = 0.03$). rs1430091 in *DPP10* predicted risk of clinical asthma (as opposed to wheezing disorders) in the cohort. In contrast, three SNPs in *DPP10*, rs13011555, rs13392783 and rs17048359, all showed association with lifetime risk of adult wheezing not labelled as asthma. rs13011555 also showed borderline association with lung function, in keeping with the risk allele for lower lung function being the same allele showing association with adult wheezing. Tables 3–6 show data for the main risk SNPs for these secondary end points.

Haplotype analyses

Given the association seen with the range of SNPs in *DPP10*, *ADAM33* and wheezing/asthma outcomes, we undertook a haplotype-based analysis to identify the major risk haplotype(s) in the Caucasian population. Statistically significant heterogeneity in incidence was evident only for *DPP10*; this was mainly attributable to variations in adult-onset disease ($p = 0.0002$) and wheezing not labelled as asthma ($p = 0.005$). A simple haplotype analysis using rs13011555 and rs13392783 (see the Methods section for details) showed that the combination T.G had the highest risk for developing phenotypes related to wheezing illness (see table 7): the major risk

appears to be for wheezing rather than diagnosed asthma. The proportion of asthma or wheezing illness statistically attributable to common variation in *DPP10* was 7%. Asthma risk was not associated with any haplotype in *ADAM33* based upon rs511898, rs528557, rs574174 and rs612709.

In the light of previous published data, we looked specifically at an extended haplotype across *GPR4* for SNPs rs323917, rs323922, rs324377, rs324396 and rs740347 which had previously been suggested to be associated with asthma risk, C.C.A.C.C, but saw no significant increase in risk of asthma or wheezing phenotypes (OR for asthma/wheeze ever 1.06, 95% CI 0.95 to 1.19).

DISCUSSION

Extensive efforts have been invested in attempting to identify genes predicting risk of asthma and related traits using positional cloning approaches. In this study we have assessed the contribution of polymorphisms in the genes identified to date to estimate population-attributable risks. We found evidence supporting a role for *DPP10* in the development of wheezing phenotypes, particularly in later life, and also saw weaker associations of SNPs in *ADAM33* with progression of wheezing illness. Borderline association with *DPP10* and lung function was also seen. Perhaps surprisingly, given the known heritability of the relevant traits, we did not see association of any individual SNP with either atopy or log total IgE, despite reasonable data from linkage studies suggesting a contribution of both *DPP10* and *PHF11* to these phenotypes.^{2,3} The maximum OR seen with risk alleles for wheezing phenotypes was around 1.1. However, as the major allele of these SNPs is often the risk allele for these common complex traits this is perhaps not surprising. Because the major alleles were usually

Table 4 Risk of developing clinical asthma at any age and risk SNPs in *DPP10*, *ADAM33* and *GPR4*

Gene; SNP	Alleles		Asthma at any age	Homozygote allele 1	Heterozygote	Homozygote allele 2	2df p Value	1df p Value	OR (per allele)	95% CI for OR
	1	2		n	n	n				
<i>DPP10</i> rs13011555	C	T	Case	103	387	336	0.901	0.674	1.02	0.92 to 1.14
			Control	832	3143	2820				
<i>DPP10</i> rs13392783	G	A	Case	290	406	124	0.598	0.942	1.00	0.90 to 1.11
			Control	2464	3230	1080				
<i>GPR4</i> rs323922	G	C	Case	131	392	300	0.836	0.825	0.99	0.89 to 1.10
			Control	1061	3304	2415				
<i>ADAM33</i> rs528557	C	G	Case	472	319	46	0.307	0.142	0.92	0.81 to 1.03
			Control	3726	2682	458				

The 2df test relates to whether prevalence varies across the three genotypes (without an assumed genetic model). The 1df test assumes a per-allele (additive) model. SNP, single nucleotide polymorphism.

Table 5 Risk of developing wheezing illness in the absence of a clinical diagnosis of asthma and risk SNPs in *DPP10*, *ADAM33* and *GPRA*

Gene; SNP	Alleles		Wheeze not asthma any age	Homozygote allele 1	Heterozygote	Heterozygote	2df p Value	1df p Value	OR (per allele)	95% CI for OR
	1	2		n	n	n				
<i>DPP10</i> rs13011555	C	T	Case	376	1373	1159	0.073	0.025	1.08	1.01 to 1.16
			Control	559	2157	1997				
<i>DPP10</i> rs13392783	G	A	Case	1113	1350	438	0.010	0.005	1.10	1.03 to 1.18
			Control	1641	2286	766				
<i>GPRA</i> rs323922	G	C	Case	429	1400	1067	0.136	0.048	0.93	0.87 to 1.00
			Control	763	2296	1648				
<i>ADAM33</i> rs528557	C	G	Case	1616	1144	175	0.257	0.202	0.95	0.88 to 1.03
			Control	2582	1857	329				

The 2df test relates to whether prevalence varies across the three genotypes (without an assumed genetic model). The 1df test assumes a per-allele (additive) model. SNP, single nucleotide polymorphism.

the risk alleles, calculations of the population-attributable risk suggest that as much as 6–7% of disease risk in the population is potentially attributable to the SNPs studied in *DPP10* and *ADAM33*. These values are in keeping with estimates derived from a meta-analysis of *ADAM33*¹⁴ and are likely to be typical for risk alleles for asthma. In all we are potentially able to explain a maximum of <13% of the total population risk of developing asthma by the polymorphisms studied in *DPP10* and *ADAM33*, implying that other genes must contribute overall to the risk of disease development. These effects were not due to geographical stratification, which for the risk SNPs identified was not statistically significant when analysed by UK regions (data available at www.b58cgenegene.sgu.ac.uk).

There are several important implications of these findings. First it is clear that studies attempting accurately to define population-attributable risks for specific polymorphisms in risk genes for common polygenic disorders such as asthma are going to require very large study populations. There have been a number of studies in smaller populations, typically of a few hundred individuals, looking at the genes we investigated; most of these studies were thus underpowered to determine risk accurately. During the analysis of this study, the first genome-wide association study in childhood asthma was reported¹⁵; interestingly, whilst one new region was identified with strong evidence for an asthma-related gene, a number of suggestive regions were identified which would be in keeping with our conclusions at least for this phenotype, suggesting that multiple genes of small effect are involved. It is clear that studies will require many thousands of cases and controls to determine adequately the contribution of specific risk alleles to disease status overall.

Secondly, despite the weak effects of individual SNPs, our data provide support for a role for *DPP10* in the development of

wheezing disorders, particularly later in life. *DPP10* codes for a homologue of dipeptidyl peptidase which cleaves terminal dipeptides from cytokines and chemokines, hence one might think polymorphisms in this gene would be more likely to show association with atopy-related phenotypes. However, in the original study which identified this gene as a potential candidate, two loci were identified: one showing association with IgE levels and one (in a separate LD block) showing association with asthma.² One SNP in each of these regions showed significant association with wheezing phenotypes in this study, but we did not find evidence supporting a contribution of this gene to total IgE. Because of the LD structure of *DPP10*, the large size of the gene and the lack of knowledge regarding functional polymorphisms, it is conceivable we may have missed an association with atopy by our choice of tagging SNPs. *DPP10* was initially identified in families with children with asthma,² and our primary observation of an association with asthma or wheeze by age 42 is consistent with this. It is interesting, therefore, that a secondary analysis suggests that the effect in our study appears to be driven by a relationship with asthma or wheeze of onset after 17. It is unclear whether the signal for asthma in children in this study is attenuated by noise from factors contributing to non-asthmatic childhood wheezing as we use a composite end point. Additionally, it is uncertain whether the parents in the initial linkage study had a significant degree of asthma of later onset.

The association seen between rs528557 in *ADAM33* and wheezing phenotypes is consistent with those previously reported in the literature^{14 16–18} in terms of the risk allele. Although a previous meta-analysis¹⁴ identified this polymorphism as a risk factor for asthma, it is worth emphasising that the total numbers of cases and controls combined in the meta-analysis (2964) is

Table 6 Lung function (FEV₁ in litres) and risk SNPs in *DPP10*, *ADAM33* and *GPRA*

Gene; SNP	Alleles			Homozygote allele 1	Heterozygote	Homozygote allele 2	2df p Value	Beta per allele	95% CI for beta
	1	2		n	n	n			
<i>DPP10</i> rs13011555	C	T	n	866	3236	2868	0.047	0.03	0.00 to 0.03
			Mean FEV ₁	3.27 (0.47)	3.31 (0.47)	3.31 (0.46)			
<i>DPP10</i> rs13392783	G	A	n	2515	3323	1110	0.099	−0.01	−0.03 to 0.00
			Mean FEV ₁	3.3 (0.45)	3.30 (0.47)	3.33 (0.48)			
<i>GPRA</i> rs323922	G	C	n	1091	3369	2488	0.500	−0.01	−0.02 to 0.01
			Mean FEV ₁	3.32 (0.47)	3.30 (0.47)	3.30 (0.46)			
<i>ADAM33</i> rs528557	C	G	n	3837	1820	455	0.174	0.01	−0.01 to 0.03
			Mean FEV ₁	3.30 (0.47)	3.30 ± 0.47	3.34 (0.43)			

Regression models adjusted for height, sex, month of year, recent chest infection, and nurse undertaking test (and thus spirometer and area).

2df p Values indicate each genotype at a locus was assigned a separate parameter estimate.

FEV₁, forced expiratory volume in 1 s; SNP, single nucleotide polymorphism.

Table 7 Estimated frequency of simplified common haplotypes for *DPP10* and *ADAM33*, and per-copy ORs and 95% CIs for their association with wheezing outcomes (relative to the haplotype with the lowest risk of asthma and/or wheezing through the lifecourse), heterogeneity tests and estimates of population-attributable risk fraction (PARF) for those haplotypes with significant association

Gene	rs13011555	rs13392783	Haplotype frequency (%)	Asthma/WB/wheeze ever		Asthma/WB onset 0 to 16 years		Asthma/wheeze onset after 16 years		Asthma ever by 42 years		Wheeze not asthma by 42 years		Prognosis 0–16 to age 42 years		Atopy at 45		
				OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR
<i>DPP10</i>	C	A	27.4	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	
	C	G	37.1	1.07, 0.99 to 1.15	1.05, 0.96 to 1.15	1.07, 0.97 to 1.18	0.98, 0.88 to 1.10	1.08, 1.00 to 1.17	1.08, 0.90 to 1.29	1.05, 0.97 to 1.14	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.08, 0.90 to 1.29	1.05, 0.97 to 1.14	1.08, 0.90 to 1.29	1.05, 0.97 to 1.14	1.04, 0.94 to 1.15
	T	A	12.4	1.05, 0.96 to 1.14	1.03, 0.93 to 1.15	0.96, 0.85 to 1.08	0.99, 0.87 to 1.15	1.05, 0.96 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.04, 0.94 to 1.15	1.04, 0.94 to 1.15
	T	G	23.1	1.17, 1.08 to 1.28	1.05, 0.94 to 1.17	1.22, 1.10 to 1.38	1.02, 0.89 to 1.17	1.18, 1.07 to 1.28	1.15, 0.93 to 1.44	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.08, 0.87 to 1.33	1.04, 0.94 to 1.15	1.04, 0.94 to 1.15	1.04, 0.94 to 1.15
p Value (3df)			0.004		0.72		0.0002		0.93		0.005		0.65		0.65		0.65	
PARF (%)			6.7		6.7		6.7		7.2		7.2		7.2		7.2		7.2	

Gene	rs511898	rs528557	rs574174	rs612709	Asthma/WB/wheeze ever		Asthma/WB onset 0 to 16 years		Asthma/wheeze onset after 16 years		Asthma ever by 42 years		Wheeze not asthma by 42 years		Prognosis 0–16 to age 42 years		Atopy at 45	
					OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI	OR	95% CI
<i>ADAM33</i>	C	G	T	G	11.6	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference	1.00, reference
	T	C	C	A	65.1	1.09, 0.99 to 1.21	1.07, 0.94 to 1.21	1.13, 0.98 to 1.30	1.13, 0.98 to 1.30	1.13, 0.96 to 1.33	1.05, 0.94 to 1.16	1.05, 0.81 to 1.36	1.00, 0.89 to 1.12	1.05, 0.81 to 1.36	1.00, 0.89 to 1.12	1.05, 0.81 to 1.36	1.00, 0.89 to 1.12	1.00, 0.89 to 1.12
	T	G	C	G	11.9	1.02, 0.90 to 1.16	1.04, 0.89 to 1.22	1.15, 0.96 to 1.37	1.02, 0.83 to 1.26	1.02, 0.83 to 1.26	1.02, 0.89 to 1.16	0.96, 0.69 to 1.33	1.10, 0.95 to 1.27	0.96, 0.69 to 1.33	1.10, 0.95 to 1.27	0.96, 0.69 to 1.33	1.10, 0.95 to 1.27	1.10, 0.95 to 1.27
	All other combinations					11.3	1.08, 0.95 to 1.24	1.14, 0.97 to 1.34	1.06, 0.89 to 1.27	1.15, 0.93 to 1.42	1.03, 0.90 to 1.18	1.26, 0.91 to 1.74	1.04, 0.89 to 1.20	1.26, 0.91 to 1.74	1.04, 0.89 to 1.20	1.26, 0.91 to 1.74	1.04, 0.89 to 1.20	1.04, 0.89 to 1.20
p Value (3df)					0.23		0.41		0.29		0.30		0.82		0.33		0.43	
PARF (%)					6.7		6.7		6.7		6.7		6.7		6.7		6.7	

Data on *GPRA* are summarised in the text.

substantially smaller than the population used for the current study. Furthermore, this polymorphism has also been associated with decline in lung function¹⁶ which may explain its association with wheezing but not a diagnosis of asthma. However, the size of effect we report is smaller than that previously seen, and no significant effect was seen on lung function.

In contrast to *ADAM33* and *DPP10*, we were unable to identify robust associations between any relevant end point and the SNPs chosen in *PHF11*, *PGDR* or *GPRA*. We extended our analysis of *GPRA* to look at the possible association between the major risk haplotype previously identified in the PARSIFAL population at this locus but did not identify a significant association. This suggests that previously defined risk alleles/haplotypes within these three genes do not appear to predict risk of asthma or allergic disease in the UK population. These data contrast with a recent study where association was seen with SNPs upstream of *GPRA*.¹⁹ That study differs from the data presented here in several key aspects: it included data from both American non-Hispanic white and Costa Rican populations, all cases were children, measurements were taken at a single time point and family-based analysis techniques were used. The study also genotyped approximately four times as many SNPs but each population was <500 individuals, raising the possibility of false-positive association explaining the findings.

The data we present have not been corrected for multiple testing. It is difficult to know the best way to do this, given that there were a priori reasons for suspecting the genes studied would be linked to asthma risk, and that effects would probably be additive between different genes. The SNPs and phenotypes are not truly independent variables, and hence comprehensive correction for multiple testing would be overly conservative. Hence we have chosen to present the data uncorrected. If one were to apply a Bonferroni correction in order to control the family-wise error rate at the level of the number of genes examined, then the associations with *DPP10* would remain significant. Had the extremely conservative approach of correcting at the level of the effective number of independent SNPs (following the method of Nyholt²⁰) been used, rs13392783 would still have shown significant association with adult-onset asthma or wheeze.

In conclusion, this study suggests that a genetic risk profile for the development of asthma and/or allergic disease in general is likely to require information on multiple SNPs in a large number of genes, and raises the question of how, even if such a profile could be derived, it would be feasible to evaluate prospectively the predictive value of a complex profile without access to extremely large populations probably involving tens of thousands of cases and controls.

Acknowledgements: This study was funded in part by Asthma UK grant no: 05/55. The 2002–4 biomedical survey and DNA collection were funded by Medical Research Council grant no, G0000934. We are especially grateful to all members of the British 1958 Birth Cohort who participated in the field studies and who gave consent for the use of their DNA for genetic epidemiological analyses, and to the research nurses who contributed to the completion of field studies. We are additionally grateful for helpful discussions with WOCM Cookson on study design.

Competing interests: None.

Ethics approval: Protocols for the 2002–2004 biomedical examination were approved by the South East MultiCentre Research Ethics Committee. The present study was approved by the oversight committee for the biomedical examination of the British 1958 Birth Cohort.

Patient consent: All participants gave informed written consent to participate in genetic association studies.

Contributors: JB, IPH and DPS designed this study. DPS and SMR were responsible for the management of the 1958 birth cohort genetic resource. SNP selection was performed by JB and IS. JB was responsible for genetic data handling. Statistical

analysis was performed by DPS with input from IPH. The manuscript was drafted by IPH and JB; all authors reviewed and revised the manuscript and approved the final version.

REFERENCES

1. **Van Eerdewegh P**, Little RD, Dupuis J, *et al*. Association of the ADAM33 gene with asthma and bronchial hyperresponsiveness. *Nature* 2002;**418**:426–30.
2. **Allen M**, Heinzmann A, Noguchi E, *et al*. Positional cloning of a novel gene influencing asthma from chromosome 2q14. *Nat Genet* 2003;**35**:258–63.
3. **Laitinen T**, Polvi A, Rydman P, *et al*. Characterization of a common susceptibility locus for asthma-related traits. *Science* 2004;**304**:300–4.
4. **Zhang Y**, Leaves NI, Anderson GG, *et al*. Positional cloning of a quantitative trait locus on chromosome 13q14 that influences immunoglobulin E levels and asthma. *Nat Genet* 2003;**34**:181–6.
5. **Oguma T**, Palmer LJ, Birben E, *et al*. Role of prostanoid DP receptor variants in susceptibility to asthma. *N Engl J Med* 2004;**351**:1752–63.
6. **Hall IP**, Blakey JD, Al Balushi KA, *et al*. Beta2-adrenoceptor polymorphisms and asthma from childhood to middle age in the British 1958 birth cohort: a genetic association study. *Lancet* 2006;**368**:771–9.
7. **Strachan DP**, Butland BK, Anderson HR. The incidence and prognosis of asthma and wheezing illness from early childhood to age 33 in a national British cohort. *BMJ* 1996;**312**:1195–9.
8. **Strachan DP**, Griffiths JM, Anderson HR, *et al*. Ventilatory function in British adults after asthma and wheezing illness at ages 0–35. *Am J Respir Crit Care Med* 1996;**154**:1629–35.
9. **Power C**, Elliott J. Cohort profile: 1958 British birth cohort (National Child Development Study). *Int J Epidemiol* 2006;**35**:34–41.
10. **Anderson HR**, Pottier AC, Strachan DP. Asthma from birth to age 23: incidence and relationship to prior and concurrent atopic disease. *Thorax* 1992;**47**:537–42.
11. **Johnston IDA**, Strachan DP, Anderson HR. Longitudinal study of the effect of pneumonia and whooping cough in childhood on adult lung function. *N Engl J Med* 1998;**338**:581–7.
12. **Nolte H**, DuBuske LM. Performance characteristics of a new automated enzyme immunoassay for the measurement of allergen-specific IgE. Summary of the probability outcomes comparing results of allergen skin testing to results obtained with the HYTEC system and CAP system. *Ann Allergy Asthma Immunol* 1997;**79**:27–34.
13. **Fuller E**, Power C, Shepherd P, *et al*. Technical Report on the National Child Development Study Biomedical Survey 2002–2004. www.cls.ioe.ac.uk (accessed March 2009).
14. **Blakey J**, Halapi E, Bjornsdottir US, *et al*. Contribution of ADAM33 polymorphisms to the population risk of asthma. *Thorax* 2005;**60**:274–6.
15. **Moffatt MF**, Kabesch M, Liang L, *et al*. Genetic variants regulating ORMDL3 expression contribute to the risk of childhood asthma. *Nature* 2007;**448**:470–3.
16. **van Diemen CC**, Postma DS, Vonk JM, *et al*. A disintegrin and metalloprotease 33 polymorphisms and lung function decline in the general population. *Am J Respir Crit Care Med* 2005;**172**:329–33.
17. **Howard TD**, Postma D, Jongepier H, *et al*. Association of a disintegrin and metalloprotease 33 (ADAM33) gene with asthma in ethnically diverse populations. *J Allergy Clin Immunol* 2003;**112**:717–22.
18. **Werner M**, Herbon N, Gohlke H, *et al*. Asthma is associated with single-nucleotide polymorphisms in ADAM33. *Clin Exp Allergy* 2004;**34**:26–31.
19. **Hersh CP**, Raby BA, Soto-Quirós ME, *et al*. Comprehensive testing of positionally cloned asthma genes in two populations. *Am J Respir Crit Care Med* 2007;**176**:849–57.
20. **Nyholt DR**. A simple correction for multiple testing for single-nucleotide polymorphisms in linkage disequilibrium with each other. *Am J Hum Genet* 2004;**74**:765–9.

Drug and Therapeutics Bulletin (DTB)

Your key source of unbiased, independent advice

For over 45 years DTB has been an independent, indispensable part of evidence-based clinical practice. DTB offers healthcare professionals detailed assessment of, and practical advice on, individual medicines and other treatments, groups of treatment and the overall management of disease.

DTB is now also available online at <http://dtb.bmj.com>:

- ▶ browse or search all DTB content from the latest issue back to 1994
- ▶ email alerting, sophisticated searching, RSS feeds and full text links from cited references
- ▶ interactive services such as My Folders for quick access to articles that you have viewed previously and My Searches to save and re-use useful searches
- ▶ comment online on any DTB article

To subscribe, or for further information, please visit <http://dtb.bmj.com>

