Neutrophils in cystic fibrosis

D G Downey, 1,2 S C Bell, 2,3 J S Elborn⁴

¹ Adult Cystic Fibrosis Centre, Bristol Royal Infirmary, Bristol, UK; ² Adult Cystic Fibrosis Centre, The Prince Charles Hospital, Brisbane, Australia; ³ Department of Medicine, University of Queensland, Australia; ⁴ Adult Cystic Fibrosis Centre, Belfast City Hospital, Belfast. Northern Ireland. UK

Correspondence to: Dr D Downey, Adult Cystic Fibrosis Centre, Bristol Royal Infirmary, Bristol BS2 8HW, UK; damian.downey@ubht.nhs.uk

Received 5 April 2007 Accepted 13 May 2007

ABSTRACT

Lung injury in cystic fibrosis is caused by recurrent airway infection and inflammation. Neutrophils are important in combating these infections but are also the predominate cells involved in the inflammatory process. This review of neutrophils in cystic fibrosis describes the cellular mechanisms involved in their migration into the airways and their role in bacterial phagocytosis. We discuss the inflammatory process and its resolution and ultimately how neutrophil function can be modulated.

Infection and inflammation damage the lungs in patients with cystic fibrosis (CF), with changes beginning early in life. Recent CF data suggest infection initiates and sustains airway inflammation. The resultant lung injury is the main cause of morbidity and mortality in CF. Neutrophils play a vital role in lung defence against bacteria and are a fundamental component of the innate immune response. The airways in CF are characterised by a neutrophil dominated inflammatory process predominately on the respiratory epithelial surface. Indeed, neutrophils are considered responsible for the onset and promotion of the inflammatory response within the CF lung.

This review describes aspects of neutrophil biology, including migration, activation, phagocytosis, apoptosis and modulation. There are few data on whether there is a fundamental difference in the initial mechanisms of neutrophil migration into the airways of patients with CF. Here, inference is derived mainly from non-CF in vitro and in vivo studies but specific CF differences are discussed where available. Subsequent sections primarily reference available CF data.

NEUTROPHIL MIGRATION INTO THE LUNG

Recruitment of neutrophils into the lung occurs via the alveolar capillary bed and postcapillary venules. It involves complex interactions between cytokines, cell adhesion molecules and chemoattractants, leading to migration across the endothelium, the extracellular matrix and the alveolar epithelium due to a chemoattractant gradient. These processes are summarised in fig 1.

Transendothelial migration

Transendothelial migration principally involves three stages: rolling adhesion, strong adhesion and migration. Rolling adhesion is mediated by carbohydrate binding molecules called selectins, which promote leucocyte rolling.⁵ In the resting state, the endothelium expresses few adhesion molecules and is relatively non-adhesive. Adhesion begins by the rolling of neutrophils along the endothelium. This transient tethering of neutrophils is mediated by selectins and is initiated

by expression of E- and P-selectins on the endothelial surface. It is recognised that patients with CF have raised levels of circulating E-selectin⁶ and P-selectin⁷ compared with healthy controls and may reflect a persistent inflammatory process. Leucocytes constitutively express L-selectin, which can be found on the tips of leucocyte microvilli that make contact with the endothelium first.8 The binding capacity of L-selectin is rapidly and transiently increased following neutrophil activation, possibly via receptor oligomerisation.9 However, the adhesion is only strong enough to induce rolling and not to stop the neutrophil completely.⁵ L-selectin is rapidly shed from the surface of the leucocytes after their activation. $^{\tiny 10\ 11}$ This can be mediated by interleukin 8 (IL8) and other chemoattractants, such as formyl-methionylleucyl-phenylalanine and platelet activating factor (PAF). 11 L-selectin shedding is required for the regulation of leucocyte rolling. Russell et al described a decrease in the shedding of L-selectin in stimulated neutrophils from patients with CF with an acute infective exacerbation. 12 Inhibition of L-selectin shedding decreases the leucocyte rolling velocity and increases the transit time of rolling leucoytes.13 The transit time has been shown to be an important determinant of leucocyte recruitment in vivo.14 The shedding of L-selectin may also increase neutrophil recruitment from the bone marrow.¹⁵ During their time in the bone marrow, neutrophils increase their mobility, deformability and chemotactic responsiveness. 16 17 Normally only fully differentiated neutrophils enter the circulation, but stimulation of the bone marrow during an inflammatory reaction results in the release of more immature neutrophils into the circulation.18 As immature neutrophils are larger and less deformable than mature ones, they preferentially sequester in lung microvessels and may mediate inappropriate lung injury.19 20

Strong adhesion of neutrophils involves \(\beta 2 \) integrins. They are glycoproteins that have a common β -chain (CD18) and different α -chains, including CD11a, CD11b, CD11c and CD11d. Integrins are present on the neutrophil surface but in a low avidity state and unable to bind with ligands on the endothelial surface until the neutrophils are activated. It has been shown that neutrophil expression of CD11b is reduced following intravenous antibiotics for pulmonary exacerbations of CF.21 Inflamed endothelium produces chemoattractants such as PAF, leukotriene B4 and various chemokines, including the most potent neutrophil chemokine in CF, IL8. IL8 binds to the luminal surface of activated endothelium where it is able to activate neutrophils.22 Bacterial cell products such as formylated peptides and lipopolysaccharide also activate neutrophils. These cell

products and other cytokines such as tumour necrosis factor α (TNFα) also stimulate endothelial cells to synthesise IL8 and Eselectin.²³ Also, CD18 is activated during E-selectin mediated neutrophil adherence to endothelium.24 The activation of CD11/CD18 on the neutrophil surface causes a change in its conformational state to a form that recognises the endothelial ligand. The important ligands for CD11/CD18 on the endothelial surface are intercellular adhesion molecule 1 (ICAM-1) and ICAM-2, which are members of the immunoglobulin superfamily. ICAM-2 is constitutively expressed whereas ICAM-1 expression is increased on inflamed endothelium by proinflammatory cytokines (eg. TNFα).²⁵ Serum levels of soluble ICAM-1 are raised in patients with CF, even at times of clinical stability, compared with healthy controls,6 suggesting an ongoing inflammatory process. This interaction between integrins and their ligands promotes strong adhesion and stops the neutrophil rolling.

Neutrophil migration occurs predominately at the borders of endothelial cells where modifications of cell junctions allow this. The cell adhesion molecules, platelet endothelial cell adhesion molecule 1 (PECAM-1 or CD31) and junctional adhesion molecule²⁶⁻²⁸ are involved in neutrophil transmigration. Migration occurs via PECAM-1/PECAM-1 interaction while maintaining the permeability barrier of the endothelial cell monolayer.²⁷ The endothelial surface density of ICAM-1 is important in regulating this migration.²⁹

Neutrophil adhesion to pulmonary endothelial cells and migration into the lung may occur by CD11/CD18 dependent or CD11/CD18 independent mechanisms.⁵⁰ Different stimuli

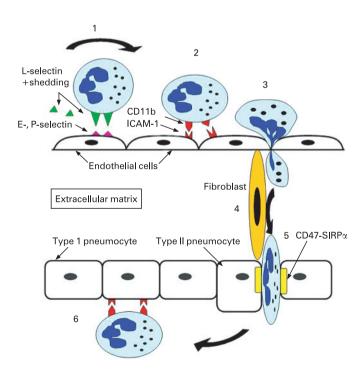


Figure 1 (1) Transient tethering of the neutrophil to the endothelial surface is mediated by selectins (green and pink) and results in rolling adhesion. (2) Strong adhesion to the vessel wall is mediated by activated $\beta 2$ integrins (red). (3) Migration between the endothelial cells is facilitated by chemokines and chemoattractants. (4) Migration through the extracellular matrix occurs along fibroblasts. (5) Migration through the alveolar epithelial cells uses CD47-signal regulatory protein α (SIRP α) (yellow). (6) Tethering to the apical surface of the alveolar epithelial cells via $\beta 2$ integrins and ICAM-1 (red). ICAM-1, intercellular adhesion molecule 1.

within the lung can determine whether CD18 is required for neutrophil migration into the lung. It is felt that stimuli from gram negative bacteria require CD18, as 75–80% of neutrophil migration is inhibited by CD18 antibodies. Animal studies indicate that neutrophils migrate to the lung via the CD18 dependent pathway in acute *Pseudomonas aeruginosa* infection, but the migration pathway shifts to the CD18 independent route after chronic exposure. Anackarel *et al* have demonstrated the preferential use of the CD18 independent migratory mechanism by both control and CF neutrophils, suggesting that blockade of the CD18 independent pathway may be a method of decreasing neutrophil influx into the CF airways.

Migration through the extracellular matrix

Following transendothelial migration, neutrophils must pass through the interstitium before entry into the lung. The interstitium consists of two types of fibroblast, one that is arranged parallel to the epithelium and the other perpendicular. Burns et al demonstrated that neutrophils had increased adherence (which is partially CD18 dependent) and motility (which is totally CD18 dependent) on canine lung fibroblasts when stimulated with PAF and IL8, respectively.35 Neutrophil adhesion to cultured lung fibroblasts is also partially dependent on fibroblast ICAM-1.35 It has also been noted that neutrophilic expression of β1 integrin is significantly increased after transendothelial migration, ³⁶ and ligation of β2 integrin provides a signal for \(\beta 1 \) integrin upregulation. 37 Several extracellular matrix proteins, including fibronectin, vitronectin, collagen and laminin have been shown to function as ligands for integrins.³⁸ It has recently been demonstrated that mindin, a extracellular matrix protein, functions as a novel ligand for integrins and plays a critical role in neutrophil recruitment.³⁹

Migration across the alveolar epithelium

Neutrophils enter the alveolar lumen at epithelial tricellular corners where the border of two type I pneumocytes meet the border of a type II pneumocyte. 40 41 It is possible that the positioning of fibroblasts in the interstitium direct them to this site. 42 The processes regulating neutrophil migration across the alveolar epithelium are not well understood. Transepithelial migration is in a basal to apical direction and, unlike transendothelial migration, the selectins and PECAM do not appear to be involved.43 Much of the understanding of the processes of transepithelial migration are based on studies of intestinal epithelial monolayers. CD11b/CD18 plays an important role in neutrophil migration but its ligand on the basal wall of alveolar cells to facilitate this has not been totally elucidated. ICAM-1 is not expressed in this region⁴⁴ but is expressed on the apical region of cell boundaries⁴⁵ and is strongly induced by viral infection of airway epithelial cells. 46 Thus expression of ICAM-1 on the apical epithelial surface provides adhesive sites for neutrophils at the site of infection that assist in antibacterial activity. 47 This has been investigated by blocking ICAM-1 receptors on CF bronchial epithelial cells; this inhibited the adherence of neutrophils by 64%.48 Furthermore, lung tissue collected at transplantation showed that neutrophils preferentially accumulated in the CF surface epithelium which overexpressed ICAM-1.7 Therefore, following neutrophil migration, ICAM-1 can provide a mechanism for retention of neutrophils at sites where they are required.

CD47, an immunoglobulin superfamily transmembrane glycoprotein, is expressed on epithelial cells and neutrophils (where it is stored in secondary specific granules). This

glycoprotein is involved in neutrophil transmigration of intestinal epithelia following $\beta 2$ -integrin dependent adhesion. Its role in neutrophil migration of the alveolar epithelium has not been described. However, Rosseau *et al* demonstrated that monocyte migration across cultured alveolar epithelial cells depended on both CD11b/CD18 and CD47. Signal regulatory protein (SIRPa) is a transmembrane glycoprotein and is a cellular ligand for CD47. Interactions between CD47 and SIRPa have been shown to regulate neutrophil transmigration. The interactions are complex and may involve SIRP on neutrophils and tissue expressed CD47 (trans interactions) or cis interactions between SIRPa and CD47 within the neutrophil membrane.

Further studies are required to explore these interactions and enhance our understanding of neutrophil migration across the CF alveolar epithelium to the site of infection.

GENES, CFTR AND THE NEUTROPHIL

Gene expression has been compared in blood neutrophils from patients with CF and healthy controls. A macroarray of 1050 genes revealed upregulation of 62 genes (including those coding for some chemokines and IL8) and downregulation of 27 genes in CF neutrophils. None of the genes coding for adhesion molecules were modulated (eg, ICAM-1 and ICAM-2). CF sputum and blood neutrophils were also compared; this demonstrated upregulation of two genes in sputum neutrophils. This included amphiregulin which is an epidermal growth factor receptor ligand that contributes to TNF induced IL8 release from airway epithelial cells, thus suggesting that amphiregulin is a new marker of lung inflammation in CF.

The most common genetic defect in CF results in defective transmembrane regulator protein (CFTR) processing so that the CFTR protein does not reach the apical surface of the epithelial cell. There is no direct link between the CF genetic defect and the process of neutrophilic migration across the airway epithelium. FETR mutations do not lead to aberrant synthesis of IL8. However, when CF neutrophils are cocultured with CFTR deficient bronchial epithelial cells, there is increased adherence and a threefold increase in IL8 levels. This interaction may contribute to the sustained inflammatory response seen in CF. It has also been demonstrated that CFTR is expressed in neutrophils at the mRNA and protein levels but it is unclear whether this specifically alters CF neutrophil function

NEUTROPHIL FUNCTIONS

When neutrophils arrive in the CF airway, they are primed, activated and engage in bactericidal phagocytosis releasing oxidants and proteases. These functions are described below.

Neutrophil priming and activation

Circulating neutrophils need to be primed to express their full bactericidal capacity. Neutrophils can also cause extensive endothelial cell injury after priming. There is evidence that TNF α and IL8 in bronchoalveolar lavage from patients with CF play a significant role in the priming and activation of CF neutrophils. When TNF α and IL8 are used as activating stimuli, CF neutrophils release significantly greater amounts of neutrophil elastase compared with neutrophils from control subjects and bronchiectatic patients. Recent data have shown that airway neutrophils from patients with CF are primed and resistant to anti-inflammatory signals delivered by IL10.

Neutrophil phagocytosis

Following transport to the site of infection, bacterial phagocytosis by neutrophils can take place. This involves two different receptor classes found on the neutrophil surface: Fc γ receptors, which include Fc γ RIIA (CD32) and Fc γ RIIB (CD16), and complement receptors which include CR1 (CD35) and CR3 (CD11b/CD18 integrin). Fc γ RIIIB and CD11b/CD18 are the functional phagocytic receptors. Fc γ receptor ligation initiates the vigorous extension of pseudopods that surround and ultimately entrap the bacteria. Changes in the level of cytosolic calcium are required for granule secretion and for granular fusion with neutrophilic phagosomes.

Both intracellular and extracellular environments are important in the regulation of neutrophil function. It is recognised that pH can modulate neutrophil function. Lack of a CFTR dependent apical epithelial bicarbonate conductance has been suggested to cause increased acidification of the airway surface liquid. In this normally acidic milieu, bacterial ingestion may induce neutrophil necrosis, rather than apoptosis, and thus promote lung parenchymal degradation.⁶³

Furthermore, with the overwhelming bacterial load and mucus characteristics of CF, the efficiency of neutrophil phagocytosis is reduced. Chronic P aeruginosa infection results in the secretion of quorum sensing compounds which play an important role in the formation of bacterial biofilms.64 Neutrophils that settle onto biofilms appear to be unable to migrate away from the point of contact even though they are still capable of phagocytosis. 65 Neutrophil accumulation within biofilms may result in self-injury of the neutrophil by released oxidants which in turn compromises host defense mechanisms66 67 and necrotic neutrophils can also serve as a biological matrix to facilitate *P aeruginosa* biofilm formation. ⁶⁸ Morris *et al* demonstrated that neutrophils from patients with CF had a lower phagocytic capacity than circulating neutrophils from the same patients or from normal control subjects.⁶⁹ The authors postulated that a failure of neutrophil phagocytic priming during migration into the lung was the cause.

Brinkmann *et al* have demonstrated that neutrophils generate extracellular fibres, or neutrophil extracellular traps. They are composed of granule (eg, neutrophil elastase and myeloperoxidase (MPO)) and nuclear constituents that disarm and kill bacteria extracellularly. Interestingly, when neutrophil extracellular traps were dismantled with recombinant human deoxyribonuclease, an enzyme which selectively cleaves DNA, the killing of bacteria was negligible.

Oxidative burst

During phagocytosis of bacteria, neutrophils increase their oxygen consumption through the activity of NADPH-oxidase and superoxide is produced (O_2^-) . The O_2^- then rapidly dismutates to form hydrogen peroxide (H2O2) catalysed by superoxide dismutase, and MPO is released from neutrophil primary granules.71 This oxidative burst is crucial for bacterial killing but it has also been implicated in inflammatory damage to the CF airways. MPO is capable of enhancing oxidative induced injury to epithelial cells, most likely because of the formation of the cytotoxic oxidant HOCl. 72-74 It has been shown that stimulated neutrophils from individuals with CF release significantly more oxidants. 75 However, a recent study demonstrated that CF neutrophils exhibit normal extracellular production of HOCl but have a defect in their ability to chlorinate bacterial proteins from *P aeruginosa*, unveiling defective intraphagolysosomal HOCl production.⁵⁵ This potentially results in reduced neutrophil phagocytic efficacy against this important CF organism. Van Der Vliet *et al* have demonstrated the formation of MPO derived oxidising and possibly nitrating species within the respiratory tract of patients with CF, which collectively may contribute to lung damage. MPO levels have been found to correlate with decreases in pulmonary function and disease severity with the MPO polymorphism -463G associated with more aggressive pulmonary disease in CF. P

Proteases

Neutrophils also release proteases which are an important component of the phagocytic process. They can also degrade extracellular matrix and are therefore implicated in CF lung damage. The most important are elastase and the matrix metalloproteinases (MMPs).

A central role in the pathophysiology of CF has been attributed to neutrophil elastase. Neutrophil elastase is stored within the primary (azurophilic) granules and released following surface activation, phagocytosis and cell death. It has been demonstrated that isolated peripheral blood neutrophils from patients with CF spontaneously release more elastase than control neutrophils.80 Importantly, this elastase production was not significantly altered following treatment with intravenous antibiotics, suggesting continuing elastase activity despite clinical improvement. Although its physiological role is to degrade phagocytosed proteins, it causes significant damage to the CF airway by degrading nearly all the structural proteins of the lung, including elastin, collagen type I-IV, fibronectin and proteoglycans.81 Urinary excretion of desmosine, a cross linking amino acid specific to elastin, is a reflection of elastin degradation. We have recently demonstrated that this and neutrophil elastase are raised in patients that ultimately have a poor outcome.82 Elastase can also cause prolongation of the inflammatory process by degrading complement and releasing C5a, a potent chemoattractant for neutrophils.83 Neutrophil recruitment may be further augmented by the effect of elastase on the epithelium to synthesise and secrete IL8.84 It can cause a reduced ciliary beat frequency of the respiratory epithelium and directly damage the epithelial cells.85 As it is a potent stimulator of airway gland serous cells,86 bacterial colonisation can be facilitated by excessive mucus production. Elastase may inactivate several components of the immune system (eg, immunoglobulins, immune complexes, complement components⁸⁷ and neutrophil cell surface receptors), ⁸⁸ thus interfering with the ability of neutrophils to opsonise and eliminate bacterial pathogens. Recently, Hartl et al have demonstrated that IL8 promotes bacterial killing by neutrophils through its chemokine receptor CXCR1 (IL8RA) and that elastase activity in bronchoalveolar lavage fluid from patients with CF cleaves CXCR1 on neutrophils and disables their bactericidal capacity.⁸⁹

Twenty-three different MMPs have been cloned to date, with additional members continuing to be identified. Matrilysin (MMP-7) is induced in response to airway injury and is markedly upregulated in CF and its catalytic activity is essential for the repair of epithelial wounds. MMP-7 is much more efficient than other metalloproteinases in the proteolytic inactivation of α_1 -antitrypsin (AAT). Macrophage metalloelastase (MMP-12) is the most elastolytic enzyme of the MMP family. Liu et al demonstrated that neutrophil derived MMP-9 provides a shield for neutrophil elastase activity. A recent study of children with CF showed that induced sputum MMP-9 had a significant correlation with neutrophils, IL8 and neutrophil elastase and an inverse relationship with forced expiratory volume in 1 s (FEV1).

In the normal lung, the airways are protected from the damaging effects of proteases mainly by AAT and secretory leucoprotease inhibitor. Secretory leucoprotease inhibitor is produced by the respiratory epithelium mainly in the larger airways. Only 33% is functionally active in the epithelial lung fluid and it is therefore unlikely that it plays a significant role in lung protection.⁹⁶

AAT is the main elastase inhibitor in CF sputum.⁹⁷ In the absence of inflammation in the normal lung the antiprotease activity of these molecules outweighs the protease burden, preventing elastase damage to the airways and local host defences. However, elevated levels of bronchoalveolar lavage neutrophil elastase have been found in patients with cystic fibrosis younger than 6 months.⁹⁸ Therefore, AAT is overwhelmed due to a relative imbalance of protease and antiprotease. It can also be inactivated by the proteases themselves.⁹⁹ Thus neutrophil elastase induced lung injury may potentially occur early in life.

RESOLUTION OF INFLAMMATION

Apoptosis plays a critical role in the host immune response and contributes to the regulation of inflammation. It is the major mechanism for removal of neutrophils from the sites of lung inflammation. ¹⁰⁰ It involves a coordinated series of morphological and biochemical steps in the cell causing its removal by scavenger phagocytes. ¹⁰¹ Early changes on the neutrophil cell surface are particularly important as they signal macrophages to phagocytose rapidly moribund cells before toxic breakdown products or contents can injure surrounding tissue. ¹⁰² It also prevents macrophages from releasing proinflammatory mediators such as chemokines, granule enzymes and thromboxane ¹⁰³ thus limiting the potential damage to the lung.

Apoptosis is a mechanism essential to the regulation of neutrophil haemostasis and inflammation. Therefore, alteration of neutrophil apoptosis in CF would have significant effects on the inflammatory response and resolution of infection.

The Fas (CD95)/Fas ligand (FasL) system is an important cellular pathway regulating the induction of apoptosis. Fas is a type 1 integral membrane protein and is a member of the TNF receptor (TNFr) family that mediates apoptosis following interaction with FasL. FasL is a type II protein member of the TNF family that includes TNFα. Fas is constitutively expressed on neutrophils, monocytes and eosinophils, whereas FasL expression is restricted to neutrophils. Therefore, co-expression of Fas and FasL on neutrophils could provide a mechanism for the spontaneous apoptosis seen in neutrophils, 104 105 or adjacent cells could initiate apoptosis if one expressed Fas and the other FasL. 106 However, a recent study has investigated Fas expression on neutrophils following intravenous antibiotics for pulmonary exacerbations of CF. Its expression on sputum neutrophils did not alter with treatment but its expression on blood neutrophils decreased following antibiotics.²¹ Soluble FasL, an inducer of apoptosis, also decreased following treatment. This at first seems counter intuitive as it would be expected that neutrophil apoptosis should increase to aid resolution of infection and inflammation. Therefore, other apoptotic pathways may be involved to assist in this.

Other important factors to consider are TNF receptors 55 and 75 (p55 TNFr-I and p75 TNFr-II). TNF α acting through these receptors has a unique ability, unlike other neutrophil priming and activating agents, to induce apoptosis. Murray $\it et~al~$ have shown this action to be bimodal (ie, prolonged incubation of human neutrophils with TNF α can reduce apoptosis). 107 Receptor p75 has a relatively short cytoplasmic domain with

no death domain sequence, unlike p55. 108 Therefore, p75 TNFr appears to function as a facilitator of the death signal primarily initiated via p55 TNFr. It is suggested that occupancy of p55 by TNF α is a prerequisite for TNF α induced neutrophil apoptosis and that TNF α binding to p75 TNFr is not critical for this process. 109 A recent study has shown a reduction in soluble p55 TNFr in sputum following treatment of pulmonary exacerbations of CF. 21

Pyocyanin, the major phenazine exotoxin, produced by P aeruginosa, has also been shown to induce apoptosis. The authors suggested that inappropriate induction of apoptosis could deplete neutrophil numbers and function and in turn impair host defence. However, it has been shown that the percentage of apoptotic neutrophils in CF sputum did not vary with different types of bacterial infection. Defective airway clearance of apoptotic cells in CF may be due to elastase mediated cleavage of phosphatidylserine receptors on phagocytes and therefore may contribute to ongoing airway inflammation. 112

It is clear that further work is required to define the role of neutrophil apoptosis in CF and its place in the resolution of the inflammatory process.

NEUTROPHIL MODULATION

Most of the major clinical manifestations, morbidity and mortality of CF are related to the progressive damage to the airways. Therefore, modulation of neutrophil function may attempt to redress this. The effects of pharmacological treatments on neutrophil function are discussed below.

Non-steroidal anti-inflammatory drugs

As CF involves infection and chronic inflammation, studies have examined the role of non-steroidal anti-inflammatory agents such as ibuprofen.113 A study by Konstan et al revealed that ibuprofen led to a slower decline in pulmonary function and improved body weight. However, there were concerns over side effects, and plasma concentrations must be tightly controlled. Paradoxically, neutrophil activation is increased by treatment with ibuprofen at doses lower than the therapeutic range. More recently, Konstan et al demonstrated a 31% reduction of neutrophils in CF oral mucosa if peak plasma ibuprofen concentration was >50 µg/ml. 114 However, Fennell et al have investigated the use of high dose ibuprofen in a paediatric CF centre and discovered that nearly half of the patients discontinued therapy due to adverse events.115 The authors commented that neither the use of ibuprofen nor its cessation resulted in a significant change in the rate of decline in pulmonary function or influenced hospitalisation rates.

Corticosteroids

An in vitro study has demonstrated that prednisolone can reduce neutrophil migration across cultured human endothelial and bronchial epithelial cells. 116 Oral corticosteroids have also reduced the rate of decline of CF lung disease but side effects have limited the use of this as therapy. 117 Despite frequent usage, the role of inhaled corticosteroids is unclear at present but a double blind placebo controlled trial of inhaled corticosteroid therapy showed no benefit from inhaled beclomethasone. 118 A recent multicentre randomised controlled trial in CF demonstrated no change in lung function or usage of rescue bronchodilators when inhaled corticosteroids were withdrawn. 119

Macrolide antibiotics

Several recent studies have reported important clinical benefits of azithromycin in patients with CF, 120-122 including improved lung function and quality of life, reduced hospitalisation and reduced systemic markers of inflammation. Macrolides may act through antimicrobial action or through immunomodulatory properties. Recent years has seen the use of macrolides, especially azithromycin, as an anti-inflammatory agent. The mode of action is not entirely clear but is thought to act by suppressing proinflammatory cytokines and altering neutrophil function. 123 124 Azithromycin in a CF mouse model attenuated neutrophil recruitment and inhibited cytokine (TNFa and macrophage inflammatory protein 2) release in lipopolysaccharide induced inflammation. 125 Macrolides can inhibit superoxide generation by activated neutrophils in vitro 126 and accelerate apoptosis. 127 A recent study of healthy human subjects administered azithromycin observed initial neutrophil degranulation. 128 Immediate oxidative responses to particulate stimuli were enhanced after azithromycin exposure, but there were reductions in IL8 and IL6 concentrations and delayed and prolonged reduction in the oxidative burst which persisted for up to 28 days after administration of azithromycin. The authors hypothesised that acute neutrophilic degranulation and oxidative burst may contribute to an antimicrobial effect and delayed inflammatory responses may contribute to an antiinflammatory effect of macrolides. Inhibition of neutrophil elastase can also be induced by the 14 membered macrolides erythromycin and flurithromycin. 129

Antiproteases

In order to protect the lung from damage mediated by neutrophil elastase, the use of AAT has been investigated. The neutrophil elastase burden in adults has been shown to be suppressed with aerosolised plasma purified AAT. 130

A double blinded, randomised, placebo controlled, parallel group trial in 39 patients with CF was carried out using nebulised transgenic AAT. 131 The safety and tolerability of AAT was demonstrated. Although MPO levels were generally lower on AAT, sputum free neutrophil elastase activity remained unchanged. However, in a study of 52 patients with CF by Griese et al, elastase activity and neutrophil numbers were reduced following inhalation of AAT. 132 Unfortunately, these studies did not show an improvement in lung function following treatment. As the study drug was only administered for 4 weeks, it is possible that more prolonged periods of AAT usage may be required before a clinical effect is seen. However, 4 weeks of inhaled AAT by subjects with CF improved the bacterial killing capacity of airway neutrophils and as a consequence the number of colony forming units of *P aeruginosa* in CF sputum decreased.89

Other treatments

Several other treatments have been recently investigated as potential anti-inflammatory therapies. Deoxyribonuclease is an established treatment modality in CF which is thought to have its major action as a mucolytic agent. It has also been demonstrated that it can stabilise bronchoalveolar lavage neutrophil numbers, elastase activity and IL8 concentration over time. It can also decrease bronchoalveolar lavage MMP levels. A recent study has investigated the use of high dose oral N-acetylcysteine, a glutathione prodrug, in 18 stable patients with CF. It revealed a reduction in airway neutrophil burden and sputum elastase activity. Furthermore, as cysteinyl

leukotrienes have been found in the sputum of patients with CF, a recent study investigated the effects of montelukast. ¹³⁶ In this 20 week, randomised, double blind, placebo controlled, crossover trial in 26 patients with CF, the authors discovered that montelukast treatment increased FEV₁ and decreased sputum levels of IL8 and MPO. Additional multicentre studies

are needed to evaluate the potential anti-inflammatory roles of

these therapies in patients with CF. Research into currently available drugs have investigated their anti-inflammatory properties and therefore may provide future therapies in CF. Pioglitazone, a peroxisome proliferator activated receptor gamma ligand, can through its action on nuclear factor κB attenuate lung ischaemia–reperfusion injury in rats. ^137 These protective effects involve inhibition of the production of proinflammatory cytokines and reduce lung neutrophil accumulation. The administration of simvastatin, a lipid lowering drug, to a murine inflammatory model of acute lung injury demonstrated reduced bronchoalveolar lavage myeloperoxidase activity and total neutrophil counts.

Anticytokines may also play an important role. Anti-TNF is an anti-inflammatory treatment for rheumatoid arthritis but TNF also plays an important role in host defense and tumour growth control. A recent metanalysis of patients with rheumatoid arthritis treated with anti-TNF therapy revealed an increased risk of serious infections and a dose dependent increased risk of malignancies, 159 suggesting that the anti-inflammatory effects can potentially have unexpected and deleterious effects on patients with chronic inflammatory conditions.

CONCLUSIONS

Review

In this review, we have followed the neutrophil in its path to the airway lumen and described its functions and modulation. Host and bacterial derived chemoattractants play a key role in the migration of neutrophils into the lung but this is a complex area and not fully understood. Acknowledging the limitations of non-CF neutrophil experimental data and its interpretation in this complex inflammatory milieu, it is recognised that there is an increased burden of neutrophils in the CF airways and that there are functional differences in these neutrophils. Importantly, neutrophils are not only important effectors of bacterial phagocytosis but are also at the centre of the inflammatory process in CF. This duality is important to understand and to investigate as insights into CF specific neutrophil function may lead to novel therapies.

Competing interests: None.

REFERENCES

- Chow CW, Landau LI, Taussig LM. Bronchial mucous glands in the newborn with cystic fibrosis. Eur J Paediatr 1982;139:240–3.
- Armstrong DS, Hook SM, Jamsen KM, et al. Lower airway inflammation in infants with cystic fibrosis detected by newborn screening. Pediatr Pulmonol 2005;40:500–10.
- Konstan MW, Hilliard KA, Norvell TM, et al. Bronchoalveolar lavage findings in cystic fibrosis patients with stable, clinically mild lung disease suggest ongoing infection and inflammation. Am J Respir Crit Care Med 1994;150:448–54.
- Khan TZ, Wagener JS, Bost T, et al. Early pulmonary inflammation in infants with cystic fibrosis. Am J Respir Crit Care Med 1995;151:1075–82.
- Lawrence MB, Springer TA. Leukocytes roll on a selectin at physiologic flow rates: distinction from and prerequisite for adhesion through integrins. Cell 1991;65:859–73.
- De Rose V, Oliva A, Messore B, et al. Circulating adhesion molecules in cystic fibrosis. Am J Respir Crit Care Med 1998;157:1234–9.
- Hubeau C, Lorenzato M, Couetil JP, et al. Quantitative analysis of inflammatory cells infiltrating the cystic fibrosis airway mucosa. Clin Exp Immunol 2001;124:69–76.
- Picker LJ, Warnock RA, Burns AR, et al. The neutrophil selectin LECAM-1 presents carbohydrate ligands to the vascular selectins ELAM-1 and GMP-140. Cell 1991;66:921–33.

- Li X, Steeber DA, Tang ML, et al. Regulation of L-selectin-mediated rolling through receptor dimerization. J Exp Med 1998;188:1385–90.
- Jutila MA, Rott L, Berg EL, et al. Function and regulation of the neutrophil MEL-14 antigen in vivo: comparison with LFA-1 and MAC-1. J Immunol 1989;143:3318–24.
- Kishimoto TK, Jutila MA, Berg EL, et al. Neutrophil Mac-1 and MEL-14 adhesion proteins inversely regulated by chemotactic factors. Science 1989;245:1238–41.
- Russell KJ, McRedmond J, Mukherji N, et al. Neutrophil adhesion molecule surface expression and responsiveness in cystic fibrosis. Am J Respir Crit Care Med 1998:157:756–61.
- Hafezi-Moghadam A, Ley K. Relevance of L-selectin shedding for leukocyte rolling in vivo. J Exp Med 1999;189:939–48.
- Jung U, Norman KE, Scharffetter-Kochanek K, et al. Transit time of leukocytes rolling through venules controls cytokine-induced inflammatory cell recruitment in vivo. J Clin Invest 1998;102:1526–33.
- van Eeden SF, Miyagashima R, Haley L, et al. A possible role for L-selectin in the release of polymorphonuclear leukocytes from bone marrow. Am J Physiol 1997:272:H1717–24.
- Lichtman MA, Weed RI. Alteration of the cell periphery during granulocyte maturation: relationship to cell function. Blood 1972;39:301–16.
- Giordano GF, Lichtman MA. Marrow cell egress. The central interaction of barrier pore size and cell maturation. J Clin Invest 1973;52:1154

 –64.
- Boggs DR. The kinetics of neutrophilic leukocytes in health and in disease. Semin Hematol 1967;4:359–86.
- Lawrence E, Van Eeden S, English D, et al. Polymorphonuclear leukocyte (PMN) migration in streptococcal pneumonia: comparison of older PMN with those recently released from the marrow. Am J Respir Cell Mol Biol 1996;14:217–24.
- van Eeden SF, Kitagawa Y, Klut ME, et al. Polymorphonuclear leukocytes released from the bone marrow preferentially sequester in lung microvessels. Microcirculation 1997;4:369–80.
- Downey DG, Brockbank S, Ennis M, et al. The effect of treatment of cystic fibrosis pulmonary exacerbations on airways and systemic inflammation. Pediatric Pulmonol 2007:42:729–35
- Rot A. Endothelial cell binding of NAP-1/IL-8: role in neutrophil emigration. *Immunol Today* 1992;13:291–4.
- Gimbrone MA Jr, Obin MS, Brock AF, et al. Endothelial interleukin-8: a novel inhibitor of leukocyte-endothelial interactions. Science 1989;246:1601—3.
- Kuijpers TW, Hakkert BC, Hoogerwerf M, et al. Role of endothelial leukocyte adhesion molecule-1 and platelet-activating factor in neutrophil adherence to IL-1prestimulated endothelial cells. Endothelial leukocyte adhesion molecule-1-mediated CD18 activation. J Immunol 1991;147:1369–76.
- Bevilacqua MP. Endothelial-leukocyte adhesion molecules. Annu Rev Immunol 1993:11:767–804
- Martin-Padura I, Lostaglio S, Schneemann M, et al. Junctional adhesion molecule, a novel member of the immunoglobulin superfamily that distributes at intercellular junctions and modulates monocyte transmigration. J Cell Biol 1998;142:117–27.
- Muller WA, Weigl SA, Deng X, et al. PECAM-1 is required for transendothelial migration of leukocytes. J Exp Med 1993;178:449–60.
- Vaporciyan AA, DeLisser HM, Yan HC, et al. Involvement of platelet-endothelial cell adhesion molecule-1 in neutrophil recruitment in vivo. Science 1993;262:1580–2.
- Yang L, Froio RM, Sciuto TE, et al. ICAM-1 regulates neutrophil adhesion and trancellular migration of TNF-activated vascular entothelium under flow. Blood 2005;106:584–92.
- Doerschuk CM, Tasaka S, Wang Q. CD11/CD18-dependent and -independent neutrophil emigration in the lungs. Am J Respir Cell Mol Biol 2000;23:133–6.
- Doerschuk CM, Winn RK, Coxson HO, et al. CD18-dependent and -independent mechanisms of neutrophil emigration in the pulmonary and systemic microcirculation of rabbits. J Immunol 1990;144:2327–33.
- Ramamoorthy C, Sasaki SS, Su DL, et al. CD18 adhesion blockade decreases bacterial clearance and neutrophil recruitment after intrapulmonary E. coli, but not after S. aureus. J Leukoc Biol 1997;61:167–72.
- Kumasaka T, Doyle NA, Quinlan WM, et al. Role of CD11/CD18 in neutrophil emigration during acute and recurrent Pseudomonas aeruginosa-induced pneumonia in rabbits. Am J Pathol 1996;148:1297–305.
- Mackarel AJ, Plant BJ, FitzGerald MX, et al. Cystic fibrosis sputum stimulates CD18-independent neutrophil migration across endothelial cells. Exp Lung Res 2005;31:377–90.
- Burns A, Simon S, Kukielka G, et al. Chemotactic factors stimulate CD18dependent canine neutrophil adherence and motility on lung fibroblasts. J Immunol 1996;156:3389–401.
- Kubes P, Niu X, Smith C, et al. A novel beta 1-dependent adhesion pathway on neutrophils: a mechanism invoked by dihydrocytochalasin B of endothelial transmigration. FASEB 1995;9:1103

 –11.
- Werr J, Eriksson EE, Herqvist P, et al. Engagement of beta2 integrins induces surface expression of beta1 integrin receptors in human neutrophils. J Leukoc Biol 2000:68:553–60
- Plow EF, Haas TA, Zhang L, et al. Ligand binding to integrins. J Biol Chem 2000;275:21785–8.
- Wei J, Hong L, You-Wen H. The extracellular matrix protein mindin serves as an integrin ligand and is critical for inflammatory cell recruitment. *Blood* 2005:106:3854–9.
- Burns AR, Walker DC, Smith CW. Relationship between tight junctions and leukocyte transmigration. In: Cereijido M, Anderson J, eds. *Tight junctions*. Boca Raton, FI: CRC, 2000:629–52.

- Damiano VV, Cohen A, Tsang AL, et al. A morphologic study of the influx of neutrophils into dog lung alveoli after lavage with sterile saline. Am J Pathol 1980;100:349–64.
- Burns AR, Smith CW, Walker DC. Unique structural features that influence neutrophil emigration into the lung. *Physio Rev* 2003;83:309–36.
- Colgan SP, Parkos CA, McGuirk D, et al. Receptors involved in carbohydrate binding modulate intestinal epithelial-neutrophils interactions. J Biol Chem 1995:270:10531–9
- 44. **Burns AR,** Takei F, Doerschuk CM. Quantitation of ICAM-1 expression in mouse lung during pneumonia. *J Immunol* 1994:**153**:3189–98.
- Taguchi M, Sampath D, Koga T, et al. Patterns for RANTES secretion and intercellular adhesion molecule 1 expression mediate transepithelial T cell traffic based on analyses in vitro and in vivo. J Exp Med 1998;187:1927–40.
- Wegner CD, Gundel RH, Reilly P, et al. Intercelllular adhesion molecule-1 (ICAM-1) in the pathogenesis of asthma. Science 1990:247:456–9.
- Humlicek AL, Pang L, Look DC. Modulation of airway inflammation and bacterial clearance by epithelial cell ICAM-1. Am J Physiol Cell Mol Physiol 2004;287:L598–607.
- Tabary O, Corvol H, Boncoeur E, et al. Adherence of airway neutrophils and inflammatory response are increased in CF airway epithelial cell-neutrophil interactions. Am J Physiol Lung Cell Mol Physiol 2006;290:L588–96.
- Parkos CA, Colgan SP, Liang TW, et al. CD47 mediates post-adhesive events required for neutrophil migration across polarized intestinal epithelia. J Cell Biol 1996:132:437–50
- Rosseau S, Selhorst J, Wiechmann K, et al. Monocyte migration through the alveolar epithelial barrier: adhesion molecule mechanisms and impact of chemokines. J Immunol 2000;164:427–35.
- Liu Y, Buhring HJ, Zen K, et al. Signal regulatory protein (SIRP), a cellular ligand for CD47, regulates neutrophil transmigration. J Biol Chem 2002;277:10028–36.
- Liu Y, O'Connor MB, Mandell KJ, et al. Peptide-mediated inhibition of neutrophil transmigration by blocking CD47 interactions with signal regulatory protein α. J Immunol 2004;172:2578–85.
- Adib-Conquy M, Pedron T, Petit-Betron AF, et al. Neutrophils in cystic fibrosis display a distinct gene expression pattern. Mol Med 2008;14:36–44.
- Pizurki L, Morris MA, Chanson M, et al. Cystic fibrosis transmembrane conductance regulator does not affect neutrophil migration across cystic fibrosis airway epithelial monolayers. Am J Pathol 2000;156:1407–16.
- Painter RG, Valentine, VG, Lanson NA Jr, et al. CFTR expression in human neutrophils and the phagolysosomal chlorination defect in cystic fibrosis. Biochemistry 2006;45:10260–9.
- Smedly LA, Tonnesen MG, Sandhaus RA, et al. Neutrophil-mediated injury to endothelial cells. Enhancement by endotoxin and essential role of neutrophil elastase. J Clin Invest 1986;77:1233–43.
- Taggart C, Coakley RJ, Greally P, et al. Increased elastase release by CF neutrophils is mediated by tumor necrosis factor-alpha and interleukin-8. Am J Physiol Lung Cell Mol Physiol 2000;278:133–41.
- Petit-Bertron AF, Tabary O, Corvol H, et al. Circulating and airway neutrophils in cystic fibrosis display different TLR expression and responsiveness to interleukin-10. Cytokine 2008:41:54–60.
- Witko-Sarsat V, Rieu P, Descamps-Latscha B, et al. Neutrophils: molecules, functions and pathophysiological aspects. Lab Invest 2000;80:617–53.
- Greenberg S, Grinstein S. Phagocytosis and innate immunity. Curr Opin Immunol 2002;14:136–45.
- Lew PD, Monod A, Waldvogel FA, et al. Quantitative analysis of the cytosolic free calcium dependency of exocytosis from three subcellular compartments in intact human neutrophils. J Cell Biol. 1986:102:2197–204.
- Jaconi MEE, Lew DP, Carpentier JL, et al. Cytosolic free calcium elevation mediates the phagosome-lysosome fusion during phagocytosis in human neutrophils. J Cell Biol 1990;110:1555–64.
- Coakley RJ, Taggart C, McElvaney NG, et al. Cytosolic pH and the inflammatory microenvironment modulate cell death in human neutrophils after phagocytosis. Blood 2002;100:3383–91.
- Singh PK, Schaefer AL, Parsek MR, et al. Quorum sensing signals indicate that cystic fibrosis lungs are infected. Nature 2000;407:762

 –4.
- Jesaitis AJ, Franklin MJ, Berglund D, et al. Compromised host defence on Pseudomonas aeruginosa biofilms: characterization of neutrophil and biofilm interactions. J Immunol 2003;171:4329–39.
- Bass DA, DeChatelet LR, Burk RF, et al. Polymorphonuclear leukocyte bactericidal activity and oxidative metabolism during glutathione peroxidase deficiency. *Infect Immun* 1977:18:78–84.
- Pietarinen-Runtti P, Lakari E, Raivio KO, et al. Expression of antioxidant enzymes in human inflammatory cells. Am J Physiol 2000;278:C118.
- 68. **Walker TS**, Tomlin KL, Worthen GS, *et al*. Enhanced Pseudomonas aeruginosa biofilm development mediated by human neutrophils. *Infect immune* 2005;**73**:3693–701.
- Morris MR, Doull IJ, Dewitt S, et al. Reduced iC3b-mediated phagocytotic capacity of pulmonary neutrophils in cystic fibrosis. Clin Exp Immunol 2005;142:68–75.
- Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. Science 2004;303:1532–5.
- Morel F, Doussiere J, Vignais PV. The superoxide-generating oxidase of phagocytic cells. Physiological, molecular and pathological aspects. *Eur J Biochem* 1991;201:523–46.
- Cantin AM, North SL, Fells GA, et al. Oxidant-mediated epithelial cell injury in idiopathic pulmonary fibrosis. J Clin Invest 1987;79:1665–73.

- Cantin A, Woods DE. Protection by antibiotics against myeloperoxidase-dependent cytotoxicity to lung epithelial cells in vitro. J Clin Invest 1993;9:38–45.
- Worlitzsch D, Herberth G, Ulrich M, et al. Catalase, myeloperoxidase and hydrogen peroxide in cystic fibrosis. Eur Respir J 1998;11:377–83.
- Witko-Sarsat V, Allen RC, Paulais M, et al. Disturbed myeloperoxidase-dependent activity of neutrophils in cystic fibrosis homozygotes and heterozygotes, and its correction by amiloride. J Immunol. 1996:157:2728–35.
- Van Der Vliet A, Nguyen MN, Shigenaga MK, et al. Myeloperoxidase and protein oxidation in cystic fibrosis. Am J Physiol Lung Cell Mol Physiol 2000;279:L537–46.
- Regelmann WE, Siefferman CM, Herron JM, et al. Sputum peroxidase activity correlates with the severity of lung disease in cystic fibrosis. Pediatr Pulmonol 1995:19:1–9
- Witko-Sarsat V, Delacourt C, Rabier D, et al. Neutrophil-derived long-lived oxidants in cystic fibrosis sputum. Am J Respir Crit Care Med 1995;152:1910–16.
- Reynolds WF, Sermet-Gaudelus I, Gausson V, et al. Myeloperoxidase promoter polymorphism –463G is associated with more severe clinical expression of cystic fibrosis pulmonary disease. Mediators Inflamm 2006;2:36735.
- Brockbank S, Downey D, Elborn JS, et al. Effect of cystic fibrosis exacerbations on neutrophil function. Int Immunopharmacol 2005;5:601–8.
- Janoff A, White R, Carp H, et al. Lung injury induced by leukocytic proteases. Am J Pathol 1979;97:111–36.
- Downey DG, Martin SL, Dempster M, et al. The relationship of clinical and inflammatory markers to outcome in stable patients with cystic fibrosis. Pediatr Pulmonol 2007;42:216–20.
- Fick RB Jr, Robbins RA, Squier SU, et al. Complement activation in cystic fibrosis respiratory fluids: in vivo and in vitro generation of C5a and chemotactic activity. Pediatr Res 1986;20:1258–68.
- Nakamura H, Yoshimura K, McElvaney NG, et al. Neutrophil elastase in respiratory epithelial lining fluid of individuals with cystic fibrosis induces interleukin-8 gene expression in a human bronchial epithelial cell line. J Clin Invest 1992;89:1478–84.
- Amitani R, Wilson R, Rutman A, et al. Effects of human neutrophil elastase and Pseudomonas aeruginosa proteinases on human respiratory epithelium. Am J Respir Cell Mol Biol 1991;4:26–32.
- Sommerhoff CP, Nadel JA, Basbaum CB, et al. Neutrophil elastase and cathepsin G stimulate secretion from cultured bovine airway gland serous cells. J Clin Invest 1990:85:682–9.
- Birrer P, McElvaney NG, Rudeberg A, et al. Protease-antiprotease imbalance in the lungs of children with cystic fibrosis. Am J Respir Crit Care Med 1994;150:207–13.
- Berger M, Sorensen RU, Tosi MF, et al. Complement receptor expression on neutrophils at an inflammatory site, the Pseudomonas-infected lung in cystic fibrosis. J Clin Invest 1989;84:1302–13.
- Hartl D, Latzin P, Hordijk P, et al. Clevage of CXCR1 on neutrophils disables bacterial killing in cystic fibrosis lung disease. Nat Med 2008;13:1423

 –30.
- Parks WC, Shapiro SD. Matrix metalloproteinases in lung biology. Respir Res 2001;2:10–19.
- Dunsmore SE, Saarialho-Kere UK, Roby JD, et al. Matrilysin expression and function in airway epithelium. J Clin Invest 1998:102:1321–31.
- Sires UI, Murphy G, Baragi VM, et al. Matrilysin is much more efficient than other matrix metalloproteinases in the proteolytic inactivation of alpha 1-antitrypsin. Biochem Biophys Res Commun 1994;204:613–20.
- Shapiro SD. Elastolytic metalloproteinases produced by human mononuclear phagocytes. Potential roles in destructive lung disease. Am J Respir Crit Care Med 1994;150:S160–4.
- Liu Z, Zhou X, Shapiro SD, et al. The serpin alpha1-proteinase inhibitor is a critical substrate for gelatinase B/MMP-9 in vivo. Cell 2000;102:647–55.
- Sagel SD, Kapsner RK, Osberg I. Induced sputum matrix metalloproteinase-9 correlates with lung function and airway inflammation in children with cystic fibrosis. Pediatr Pulmonol 2005;39:224–32.
- Vogelmeier C, Hubbard RC, Fells GA, et al. Anti-neutrophil elastase defense of the normal human respiratory epithelial surface provided by the secretory leukoprotease inhibitor. J Clin Invest 1991;87:482–8.
- Suter S, Schaad UB, Tegner H, et al. Levels of free granulocyte elastase in bronchial secretions from patients with cystic fibrosis: effect of antimicrobial treatment against Pseudomonas aeruginosa. J Infect Dis 1986;153:902–9.
- Khan TZ, Wagener JS, Bost T, et al. Early pulmonary inflammation in infants with cystic fibrosis. Am J Respir Crit Care Med 1995;151:1075–82.
- Cantin A, Bilodeau G, Begin R. Granulocyte elastase-mediated proteolysis of alpha 1-antitrypsin in cystic fibrosis bronchopulmonary secretions. *Pediatr Pulmonol* 1989;7:12–17.
- Cox G, Crossley J, Xing Z. Macrophage engulfment of apoptotic neutrophils contributes to the resolution of acute pulmonary inflammation in vivo. Am J Respir Cell Mol Biol. 1995:12:232

 –7.
- Cohen JJ. Programmed cell death in the immune system. Adv Immunol 1991:50:55–85.
- Savill JS, Wyllie AH, Henson JE, et al. Macrophage phagocytosis of aging neutrophils in inflammation. Programmed cell death in the neutrophil leads to its recognition by macrophages. J Clin Invest 1989;83:865–7.
- Meagher LC, Savill JS, Baker A, et al. Phagocytosis of apoptotic neutrophils does not induce macrophage release of thromboxane B2. J Leukoc Biol 1992;52:269–73.
- Liles WC, Klebanoff SJ. Regulation of apoptosis in neutrophils—Fas track to death? *J Immunol* 1995;155:3289–91.

- 105. Liles WC, Kiener PA, Ledbetter JA, et al. Differential expression of Fas (CD95) and Fas ligand on normal human phagocytes: implications for the regulation of apoptosis in neutrophils. J Exp Med 1996;184:429–40.
- Vignaux F, Golstein P. Fas-based lymphocyte-mediated cytotoxicity against syngeneic activated lymphocytes: a regulatory pathway? Eur J Immunol 1994:24:923–7
- Murray J, Barbara JA, Dunkley SA, et al. Regulation of neutrophil apoptosis by tumor necrosis factor-alpha: requirement for TNFR55 and TNFR75 for induction of apoptosis in vitro. Blood 1997;90:2772–83.
- Tartaglia LA, Ayres TM, Wong GH, et al. A novel domain within the 55 kd TNF receptor signals cell death. Cell 1993;74:845–53.
- Gon S, Gatanaga T, Sendo F. Involvement of two types of TNF receptor in TNFalpha induced neutrophil apoptosis. Microbiol Immunol 1996;40:463–5.
- Usher LR, Lawson RA, Geary I, et al. Induction of neutrophil apoptosis by the Pseudomonas aeruginosa exotoxin pyocyanin: a potential mechanism of persistent infection. J Immunol 2002;168:1861–8.
- Watt AP, Courtney J, Moore J, et al. Neutrophil cell death, activation and bacterial infection in cystic fibrosis. *Thorax* 2005:60:659–64.
- Vandivier RW, Fadok VA, Hoffmann PR, et al. Elastase-mediated phosphatidylserine receptor cleavage impairs apoptotic cell clearance in cystic fibrosis and bronchiectasis. J Clin Invest 2002;109:661–70.
- Konstan MW, Byard PJ, Hoppel CL, et al. Effect of high-dose ibuprofen in patients with cystic fibrosis. N Engl J Med 1995;332:848–54.
- Konstan MW, Krenicky JE, Finney MR, et al. Effect of ibuprofen on neutrophil migration in cystic fibrosis and healthy subjects. J Pharm Exp Ther 2004;306:1086–91.
- Fennell PB, Quante J, Wilson K, et al. Use of high-dose ibuprofen in a pediatric cystic fibrosis center. J Cyst Fibros 2007;6:153–8.
- Van Overveld FJ, Demkow UA, Gorecka D, et al. Inhibitory capacity of different steroids on neutrophil migration across a bilayer of endothelial and bronchial epithelial cells. Eur J Pharmacol 2003;477:261–7.
- Eigen H, Rosenstein BJ, FitzSimmons S, et al. A multicenter study of alternate-day prednisone therapy in patients with cystic fibrosis. Cystic Fibrosis Foundation Prednisone Trial Group. J Pediatr 1995;126:515–23.
- Nikolaizik WH, Schoni MH. Pilot study to assess the effect of inhaled corticosteroids on lung function in patients with cystic fibrosis. J Pediatr 1996:128:271–4.
- Balfour-Lynn IM, Lees B, Hall P, et al. Multicenter randomized controlled trial of withdrawal of inhaled corticosteroids in cystic fibrosis. Am J Respir Crit Care Med 2006;173:1356–62.
- Wolter J, Seeney S, Bell S, et al. Effect of long term treatment with azithromycin on disease parameters in cystic fibrosis: a randomised trial. Thorax 2002;57:212–16.
- Equi A, Balfour-Lynn IM, Bush A, et al. Long term azithromycin in children with cystic fibrosis: a randomised, placebo-controlled crossover trial. Lancet 2002;360:978–84.
- Saiman L, Marshall BC, Mayer-Hamblett N, et al. Azithromycin in patients with cystic fibrosis chronically infected with Pseudomonas aeruginosa: a randomized controlled trial. JAMA 2003;290:1749–56.

- Southern KW, Barber PM. Azithromycin for cystic fibrosis. Eur Respir J 2004:24:834–8.
- Jaffe A, Bush A. Macrolides in cystic fibrosis. In: Rubin BK, Tamaoki J, eds. *Antibiotics as anti-inflammatory and immunomodulatory agents*. Basel: Berghauser Verlage. 2005;167–91.
- Legssyer R, Huaux F, Lebacq J, et al. Azithromycin reduces spontaneous and induced inflammation in DeltaF508 cystic fibrosis mice. Respir Res 2006;7:134.
- 126. Anderson R. Erythromycin and roxithromycin potentiate human neutrophil locomotion in vitro by inhibition of leukoattractant-activated superoxide generation and autooxidation. J Infect Dis 1989;159:966–73.
- Aoshiba K, Nagai A, Konno K. Erythromycin shortens neutrophil survival by accelerating apoptosis. Antimicrob Agents Chemother 1995;39:872–77.
- Culic O, Erakovic V, Cepelak I, et al. Azithromycin modulates neutrophil function and circulating inflammatory mediators in healthy human subjects. Eur J Pharmacol 2002;450:277–89.
- Gorrini M, Lupi A, Viglio S, et al. Inhibition of human neutrophil elastase by erythromycin and flurythromycin, two macrolide antibiotics. Am J Respir Cell Mol Biol 2001;25:492–9.
- McElvaney NG, Hubbard RC, Birrer P, et al. Aerosol alpha 1-antitrypsin treatment for cystic fibrosis. Lancet 1999;337:392–4.
- Martin SL, Downey D, Bilton D, et al. Recombinant AAT CF Study Team. Safety and efficacy of recombinant alpha(1)-antitrypsin therapy in cystic fibrosis. Pediatr Pulmonol 2006;41:177–83.
- Griese M, Latzin P, Kappler M, et al. Alpha 1-antitrypsin inhalation reduces airway inflammation in cystic fibrosis patients. Eur Respir J 2007;29:240–50.
- 133. Paul K, Rietschel E, Ballmann M, et al. Bronchoalveolar Lavage for the Evaluation of Antiinflammatory Treatment Study Group. Effect of treatment with dornase alpha on airway inflammation in patients with cystic fibrosis. Am J Respir Crit Care Med 2004;169:719–25.
- 134. **Ratjen F**, Hartog CM, Paul K, *et al*. Matrix metalloproteases in BAL fluid of patients with cystic fibrosis and their modulation by treatment with dornase alpha. *Thorax* 2002:57:930—4
- Tirouvanziam R, Conrad CK, Bottiglieri T, et al. High-dose oral N-acetylcysteine, a glutathione prodrug, modulates inflammation in cystic fibrosis. Proc Natl Acad Sci U S A 2006;103:4628–33.
- Stelmach I, Korzeniewska A, Stelmach W, et al. Effects of montelukast treatment on clinical and inflammatory variables in patients with cystic fibrosis. Ann Allergy Asthma Immunol 2005;95:372–80.
- Ito K, Shimada J, Kato D, et al. Protective effects of preischemic treatment with pioglitazone, a peroxisome proliferator-activated receptor-gamma ligand, on lung ischemia-reperfusion injury in rats. Eur J Cardiothorac Surg 2004;25:530–6.
- Jacobson JR, Barnard JW, Grigoryev DN, et al. Simvastatin attenuates vascular leak and inflammation in murine inflammatory lung injury. Am J Physiol Lung Cell Mol Physiol 2005;288:I 1026–32.
- Bongartz T, Sutton AJ, Sweeting MJ, et al. Anti-TNF antibody therapy in rheumatoid arthritis and the risk of serious infections and malignancies: systematic review and meta-analysis of rare harmful effects in randomized controlled trials. JAMA 2006;295:2275–85.