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Sleep disordered breathing in an adult with cherubism

Cherubism is a rare disorder with bilateral enlargement of the mandible that regresses with time. Bone degrading osteoclasts and bone building osteoblasts function abnormally causing the changes. It is an autosomal dominant condition which manifests in early childhood by the age of 2–5 years, but later regresses with time. Airway obstruction occurs due to backward displacement of the tongue affecting respiration.

An 18 year old male was referred to us for snoring. On inquiry the mother reported swelling of the lower face since childhood, which had progressed to its present size. The patient had difficulty in speech, mastication, and swallowing with mental retardation. There was a history of excessive daytime somnolence, nocturia, and increased irritability during the previous few years. On examination he had characteristic features of cherubism. The lower eyelids were retracted and his oral examination revealed gingival hypertrophy with most of the upper and lower teeth embedded in the gums. The tongue was enlarged and the posterior pharyngeal wall could not be visualised. Fine needle aspiration biopsy of the mandible revealed the presence of giant cells with fibrous tissue. A diagnostic record of night time recording of respiratory variables was performed which showed the presence of snoring and an apnoea/hypopnoea index (AHI) of 12/hour associated with oxygen desaturation. A second study was performed with nasal continuous positive airway pressure (CPAP) of 7 cm H_2O, following which AHI reduced to 3/hour with no oxygen desaturation or snoring. Obstructive sleep apnoea has been reported only once earlier in a 5 year old boy who was treated with tracheostomy.[1] This is the second report associating cherubism with obstructive sleep apnoea and mental retardation, and the first report of an adult who responded to CPAP therapy.

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Sarcoidosis presenting as upper extremity venous thrombosis

Sarcoidosis is a multisystem granulomatous disorder of unknown aetiology which is characterised pathologically by the presence of non-caseating granulomas. Typical presenting symptoms include cough and dyspnoea in addition to skin and/or eye lesions. We report a case of sarcoidosis presenting as upper extremity venous thrombosis.

A 39 year old woman presented with sudden swelling of the right upper limb and right side of the neck and face. She reported no cough, dyspnoea, chest pain, or systemic symptoms of fever or weight loss. She had no previous medical history, smoked 20 cigarettes daily, and was on an oral contraceptive pill (levonorgestrel, Schering Health Care Ltd). On examination her body mass index was 31. She had extensive swelling of the right upper limb and right side of the neck and face. Her upper limb pulses were normal and she had no palpable lymphadenopathy. Full blood count, coagulation screen, electrolytes, renal function indices, and serum angiotensin converting enzyme levels were normal. Liver function tests and fasting cholesterol were mildly raised. Serum calcium and 24 hour urinary calcium levels were normal. Pulmonary function testing revealed normal spirometric parameters (FEV$_1$ 2.32 litres (85% predicted), FVC 2.97 litres (93% predicted), reduced carbon monoxide transfer factor at 15.7 mmol/min/mm Hg/l (62% predicted)). A plain chest radiograph showed slight widening of the superior mediastinum, and a computed tomographic (CT) scan showed thrombosis of the right brachiocephalic vein with enlargement of the axillary and mediastinal lymph nodes (fig 1). She underwent mediastinoscopy with biopsy of lymph nodes and histopathological examination revealed fibrosis and hyalinised granulomas consistent with sarcoidosis. She was treated with low molecular weight heparin and intravenous steroids which resulted in resolution of her symptoms and clinical signs over the subsequent week. Oral anticoagulants and corticosteroids were continued as an outpatient. A thrombophilia screen performed 16 weeks after the thrombosis revealed normal protein C and S levels, normal antithrombin III levels, and testing for the lupus anticoagulant was negative.

This is the first report to our knowledge of sarcoidosis presenting as upper extremity deep vein thrombosis, although a recent case reported in *Thorax* described a patient with tuberculous mediastinal lymphadenopathy associated with pulmonary artery occlusion.[1] In addition to venous compression by enlarged lymph nodes, possible contribu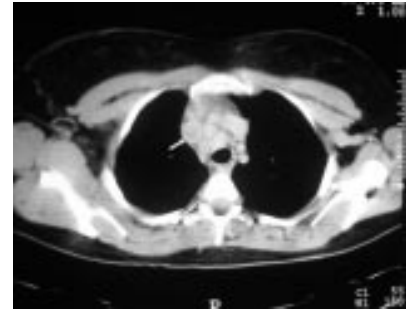ting factors in our patient include the use of the oral contraceptive pill, cigarette smoking and obesity, all of which have been associated with an increased risk of venous thromboembolic disease related to venous stasis and/or altered coagulation. Upper extremity thrombosis due to venous compression from enlarged mediastinal lymph nodes is usually associated with malignant lymphadenopathy. However, the present report shows that benign mediastinal lymphadenopathy, such as that associated with sarcoidosis, also has the potential to cause venous thrombosis.

Figure 1 CT scan of thorax showing thrombosed right brachiocephalic vein as indicated by arrow and mediastinal lymphadenopathy.

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Genotype-phenotype correlations in PCD patients carrying *DNAH5* mutations

Primary ciliary dyskinesia (PCD) is usually inherited as an autosomal recessive disorder. Affected individuals suffer from recurrent infections of the upper and lower respiratory tract due to reduced mucociliary clearance. Half of the affected offspring exhibit a complete situs inversus because of randomisation of left-right body asymmetry.[1 2] The PCD phenotype results from axonemal abnormalities in cilia and flagella. Total or partial absence of dynein arms are found in 70–80% of PCD cases.[3]

PCD represents a heterogeneous group of genetic disorders. Distinct PCD loci have been mapped to chromosome 9p13-p21 (*DNAI1*), 19q13.3-qter and 5p15-p14, respectively.[4–6] We identified *DNAH5* as the gene responsible for PCD located on 5p. *DNAH5* encodes a protein highly similar to the *Chlamydomonas* γ-dynein heavy chain.[6] Mutants of the *Chlamydomonas* orthologue show a slow swimming phenotype and are characterised by axonemal abnormalities consisting of outer dynein arm (ODA) defects.[7 8] This phenotype appears similar to that observed in a large Arab family
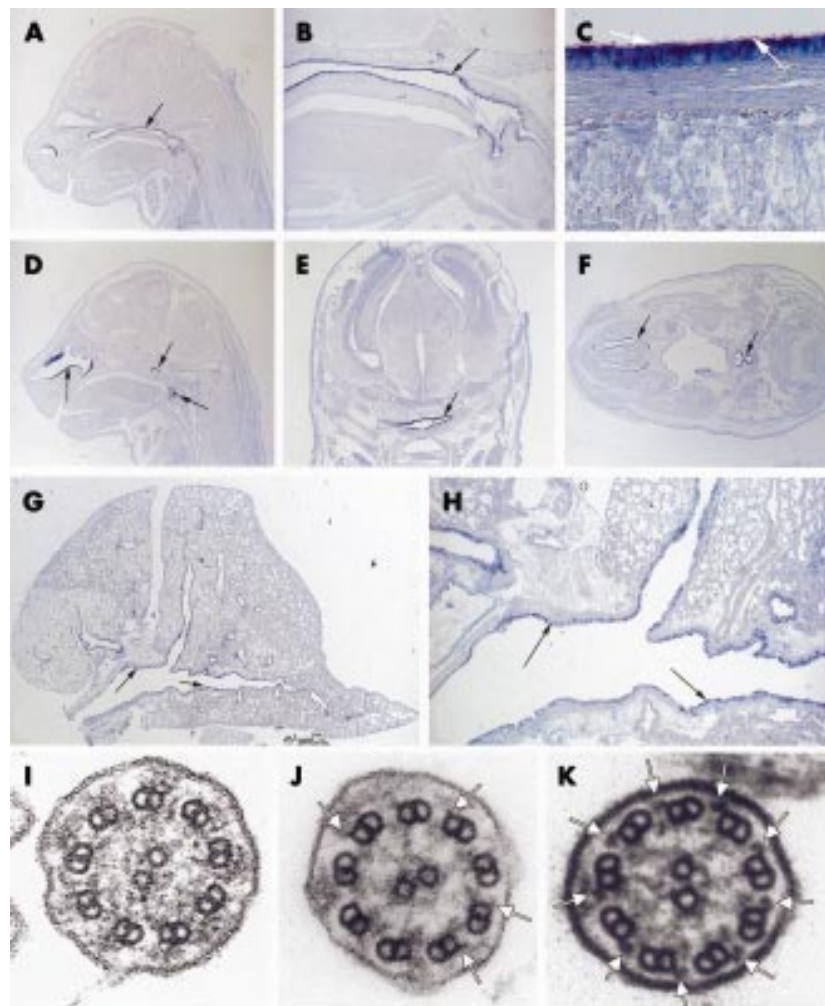
Figure 1 In situ hybridisation analysis of mouse *Dnahc5* expression in the developing and adult respiratory system and ultrastructural defects in patients with PCD carrying *DNAH5* mutations. (A)–(F) Section in situ hybridisation analysis of *Dnahc5* expression in mouse 16 days post coitum embryos. Photographs show details of expression in the nasopharynx, larynx and trachea, respectively. Note cilia on epithelial cells in C (white arrows). (G), (H) Section in situ hybridisation analysis of *Dnahc5* expression in mouse adult lung. Note epithelial expression in the whole bronchial system. (I) Electron micrograph of a cross section of a respiratory cilium from an affected individual from PCD family F658. Absence of outer dynein arms (ODA) is observed on all of the nine peripheral doublets. (J) Electron micrograph of a cross section of a cilium from the respiratory epithelium of an affected individual of PCD family UNC-7. The number of visible ODA on the peripheral doublets is reduced. The remaining ODA are shorter (arrows). (K) Electron micrograph of a cross section of a respiratory cilium from a healthy individual (control). Arrows indicate the location of ODA.

used to map the PCD locus.[6] Sequence analysis of the *DNAH5* gene in PCD patients with randomisation of left-right asymmetry identified mutations resulting in non-functional DNAH5 proteins.[9]

The murine orthologue of human *DNAH5*, called *Dnahc5*, is predominantly expressed in the lung as shown by Northern blot analysis.[9] During gastrulation expression is confined to the node, which explains the randomisation of left-right asymmetry in PCD. Mice with a targeted mutation in *Dnahc5* display a phenotype highly similar to that observed in patients with PCD.[10] *Dnahc5* deficient mice develop respiratory symptoms due to reduced mucociliary clearance, and half of the affected offspring have complete situs inversus.

In order to gain insight into the development of the disease phenotype and the function of *DNAH5*, we have studied the expression of *Dnahc5* in the murine respiratory tract in both embryonic and adult tissue of mice using section in situ hybridisation analysis as described previously.[9] We found that *Dnahc5* expression is confined to ciliated epithelial cells of the upper and lower airways (fig 1A–H). This expression pattern is consistent with the PCD phenotype of humans resulting from mutations in *DNAH5* and mice with a targeted disruption of *Dnahc5*, respectively.[9 10] Our expression data strongly suggest that ultrastructural abnormalities resulting from *DNAH5* mutations should be present in ciliated respiratory epithelia of the nasopharynx, the larynx, and the bronchi. Sampling of respiratory cilia at different sites of the airway should not therefore affect ultrastructural findings caused by *DNAH5* mutations.

Individuals of different PCD families exhibit various degrees of respiratory symptoms. We hypothesised that the severity of the disease phenotype might correlate with the molecular nature of the *DNAH5* mutation in a family. Ultrastructural analysis of respiratory cilia in three families carrying homozygous mutations of *DNAH5* indicated such a genotype-phenotype correlation. Electron microscopic photographs of respiratory cilia from families F373 and F658 have been reported previously and were compared with ultrastructural findings of the UNC-7 family.[7 10] Mutations causing premature translational termination of DNAH5 (1855NfsX5, 2814fsX1) result in a complete absence of all ODA in respiratory cilia (fig1I–K). In contrast, a splice site mutation predicting a loss of exon 75 (IVS74–1G>C) did not cause total absence of ODA. We semiquantitatively assessed ciliary axonemes from the affected siblings of UNC-7 for the presence of ODA in a blinded manner (n=36 cilia for one sibling, n=9 from the other).[11] Both siblings had shortened stubby ODA compared with normal. Computer aided quantitative measurement showed that 54% of the ODA were less than half the average length of ODA in normal subjects, which indicates partial ODA deficiency.

We provide evidence for the first genotype-phenotype correlation in PCD. *DNAH5* mutations should be considered in individuals with total and partial absence of ODA.

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