Plasma phospholipase A$_2$ activity in patients with asthma: association with body mass index and cholesterol concentration

N L A Misso,[1] N Petrovic,[1] C Grove,[1] A Celenza,[2] J Brooks-Wildhaber,[1] P J Thompson[1]

[1] Lung Institute of Western Australia (Inc.), Centre for Asthma, Allergy and Respiratory Research, The University of Western Australia, Perth, Australia; [2] Discipline of Emergency Medicine, The University of Western Australia, Perth, Australia

Correspondence to:
Dr N L Misso, Lung Institute of Western Australia, Ground Floor, E Block, Sir Charles Gairdner Hospital, Nedlands, WA 6009, Australia; nmisso@liwa.uwa.edu.au

ABSTRACT

Background: Secretory phospholipases A$_2$ (sPLA$_2$) have functions relevant to asthmatic inflammation, including eicosanoid synthesis and effects on dendritic cells and T cells. The aim of this study was to measure sPLA$_2$ activity in patients with stable and acute asthma and to assess potential associations with body mass index (BMI), and plasma cholesterol and vitamin C concentrations.

Methods: Plasma sPLA$_2$ activity and concentrations of cholesterol and vitamin C were measured in 23 control subjects and 61 subjects with stable asthma (42 mild to moderate, 19 severe). In addition, sPLA$_2$ activity was measured in 36 patients experiencing acute asthma and in 22 of these patients after recovery from the acute attack.

Results: sPLA$_2$ activity was not significantly greater in severe (499.9 U; 95% confidence interval (CI) 439.4 to 560.4) compared with mild to moderate asthmatic subjects (464.8; 95% CI 425.3 to 504.3) or control subjects (445.7; 95% CI 392.1 to 499.4), although it was higher in patients with acute asthma (581.6; 95% CI 541.2 to 622.0; p<0.001). Male gender, high plasma cholesterol, increased BMI and atopy were associated with increased sPLA$_2$ activity, while plasma vitamin C was inversely correlated with sPLA$_2$ activity in patients with stable asthma and in control subjects. There were significant interactions between gender and plasma cholesterol and between gender and vitamin C in relation to sPLA$_2$ activity.

Conclusions: Plasma sPLA$_2$ may provide a biological link between asthma, inflammation, increased BMI, lipid metabolism and antioxidants. Interactions among these factors may be pertinent to the pathophysiology and increasing prevalence of both asthma and obesity.

Phospholipases A$_2$ (PLA$_2$) are a large group of enzymes that hydrolyse fatty acids, including arachidonic acid, from the sn-2 position of glycerophospholipids, providing the substrate for synthesis of leucotrienes, prostaglandins, platelet activating factor and lysophospholipids that are important in lung inflammation and asthma.[1] The PLA$_2$ superfamily also plays crucial regulatory roles in phospholipid metabolism, host defence and signal transduction.[2] The secretory phospholipases A$_2$ (sPLA$_2$) are low molecular weight enzymes found in extracellular fluids, including plasma. In humans, there may be as many as six or seven of these calcium dependent enzymes that are resistant to proteolysis and denaturation.[3 4] A role for these sPLA$_2$ enzymes in inflammatory processes has been suggested by studies showing increased serum sPLA$_2$ activity in acute pancreatitis and rheumatoid arthritis.[5 6]

Asthma is a chronic inflammatory disease, involving T lymphocytes, eosinophils, basophils, neutrophils and mast cells,[7] and many of these cells express and release sPLA$_2$.[4 8] Furthermore, recent studies indicate that sPLA$_2$ is likely to have numerous biological functions that are relevant to the pathogenesis of asthma, including eicosanoid synthesis,[1 4] maturation and migration of dendritic cells,[9 10] T cell proliferation[11] as well as cytokine and chemokine production by monocytes, macrophages, neutrophils and eosinophils.[12–15] However, patient studies directly implicating sPLA$_2$ in the pathophysiology of asthma are limited. Increased serum and leucocyte PLA$_2$ activity has been reported in patients with asthma and rhinitis.[16 17] An increase in sPLA$_2$ activity was also observed in nasal lavage following challenge of allergic subjects,[18] and in bronchoalveolar lavage fluid following antigen challenge of patients with asthma.[19]

Recent epidemiological studies have identified associations between obesity and increases in the incidence and prevalence of asthma.[20 21] Therefore, lipid parameters such as plasma triglyceride and cholesterol concentrations that are associated with increased body mass index (BMI)[22] may also be associated with the activities of lipid modifying enzymes, such as sPLA$_2$ in patients with asthma. Furthermore, the antioxidant vitamins C and E appear to inhibit the oxidative modification of lipoproteins that are substrates for sPLA$_2$.[23 24]

We recently developed an assay for a low molecular weight sPLA$_2$ in serum, and showed that this enzyme was associated with high density lipoproteins and was strongly correlated with total cholesterol, low density lipoprotein cholesterol and triglyceride concentrations in healthy subjects.[25] The aim of the present study was to extend these findings by comparing sPLA$_2$ activity in patients with stable, mild to moderate or severe asthma, or acute exacerbations of asthma, with that in healthy control subjects. In addition, we assessed the potential associations between sPLA$_2$ activity and BMI, plasma total cholesterol and plasma vitamin C concentrations in patients with stable asthma and in control subjects.

METHODS

Subjects

Patients with stable asthma were recruited from the databases of the Lung Institute of Western Australia and Sir Charles Gairdner Hospital, Perth.

These institutions treat patients with mild to moderate to severe asthma from the entire Perth metropolitan area. All patients had been diagnosed as asthmatic by a respiratory physician and were categorised as having severe asthma if they met at least four of the following criteria, which were modifications of the National Asthma Education and Prevention Program, Expert Panel Report II guidelines[26]: (1) β2 agonist use ≥3 times per day, on most days in the previous 3 months, (2) regular inhaled corticosteroid use (≥2000 μg beclomethasone equivalent/day) in the previous 3 months, (3) use of oral corticosteroids within the past 12 months, (4) hospital admission for asthma in the previous 12 months, (5) ≥3 unplanned visits to a general practitioner in the previous 12 months because of asthma exacerbations, (6) daily asthma symptoms including cough, wheeze, chest tightness and breathlessness when asthma was unstable in the previous 3 months and (7) nocturnal awakening due to asthma symptoms more than twice a week in the past 3 months. All patients were studied when their asthma was stable, and the severity categorisation was designed to reflect their chronic long term asthma severity. On this basis, 19 patients were categorised as having severe asthma, while the 42 patients who did not meet the criteria for severe asthma were categorised as having mild to moderate asthma.

In order to recruit non-asthmatic control subjects from a comparable population, letters inviting subjects to participate were mailed to addresses randomly selected from the Perth metropolitan telephone directory. The 23 non-asthmatic subjects who were recruited completed a brief questionnaire relating to their general health status. All patients and control subjects were recruited within a 6 month period. They completed a lung function test, and their atopic status was assessed by skin prick testing. A weal diameter ≥3 mm was considered to be a positive reaction. A blood sample was taken and placed on ice for transfer to the laboratory within 2 h. The characteristics of the patient and control groups are presented in table 1.

For the study of plasma sPLA₂ activity in acute asthma, patients were recruited on presentation to the emergency department at Sir Charles Gairdner Hospital with an acute exacerbation of their asthma. A blood sample was taken while the patient was being assessed and before oral corticosteroid treatment was commenced. These patients were followed-up 6–8 weeks later when their lung function had returned to normal or to their previous best, and at this time a second blood sample was taken. The characteristics of the patients recruited for the acute asthma study are presented in table 1. All of the research undertaken was approved by the Sir Charles Gairdner Hospital Human Research Ethics Committee and all subjects gave informed consent.

Assay of plasma sPLA₂ activity

Plasma samples were purified on heparin–sepharose affinity columns (HiTrap Heparin HP; Amersham Biosciences, Sydney, Australia) that were prewashed with 50 mM Tris-HCl, pH 7.5, before application of the plasma sample (0.5 ml). Columns were washed with 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, and sPLA₂ activity was eluted with 0.5 ml of 50 mM Tris-HCl, 1 M KCl. sPLA₂ activity in purified plasma samples was assayed using 4-nitro-3-octanoyloxy-benzoic acid as substrate, as described previously.[25] Briefly, substrate solution (180 μl, 2 mM final concentration) and purified plasma (20 μl) were incubated in microplate wells at room temperature for 2 h. Absorbances at 425 and 600 nm (to correct for turbidity in the sample) were then measured on a spectrophotometer (Molecular Devices, Sunnyvale, California, USA). A unit (U) of sPLA₂ activity was defined as nmol of product formed by 1 ml of plasma in a 1 h incubation and was calculated as

$$[(OD_{425nm} - OD_{600nm}) \times 78.62 \times 25]$$

where 78.62 is the nmol of product producing an OD_{425} of 1.0 in 0.2 ml and 25 is the correction factor for 20 μl of plasma to 1 ml and a 2 h incubation to 1 h.

Cholesterol and vitamin C concentrations

These analyses were performed in the Department of Clinical Biochemistry, PathWest, Perth, Western Australia. Total plasma cholesterol was determined by a standard automated spectrophotometric method. For vitamin C analyses, plasma was deproteinised with dithiothreitol-EDTA-perchloric acid, and ascorbic acid in the supernatant was separated and quantified by reverse phase high pressure liquid chromatography using external standards (0–227 μM). Ascorbic acid measurements

Table 1 Characteristics of patients with asthma and control subjects

	Controls	Mild to moderate asthma	Severe asthma	Acute asthma
Subjects (n)	23	42	19	36
Gender (M/F)	13/10	17/25	6/13	10/26
Age (y) (mean (SD))	44.3 (9.8)	48.7 (13.8)	47.5 (12.5)	37.5 (16.9)
Atopic (n (%))	14 (60.9)	36 (85.7)	13 (68.4)	17 (89.5)*
Smokers (n (%))	5 (21.7)	1 (2.4)	3 (15.8)	ND
FEV₁ (% predicted) (mean (SD))	99.4 (14.4)	87.9 (16.5)	69.9 (18.6)	79.6 (25.6)*
BMI (kg/m²) (mean (SD))	23.8 (3.5)	25.2 (5.7)	28.8 (7.0)	ND
Asthma medications				
Inhaled corticosteroids (n (%))		28 (67.7)	18 (94.7)	28 (80)
Daily ICS dose (μg) (mean (SEM))†		1013 (157)	2317 (254)	1566 (197)
Oral corticosteroids (n (%))‡		0	7 (36.8)	7 (19.4)
β adrenoceptor agonists (n (%))		29 (69)	18 (94.7)§	31 (88.6)¶

*Skin prick data were available for 19 acute patients, and FEV₁ when stable was measured in 20 acute patients.
†Beclomethasone equivalent dose.
‡Use of oral corticosteroids at the time of blood sampling.
§One patient refused to use β₂ agonists and used theophylline and sodium cromoglycate instead.
¶Five acute patients were not using β₂ agonists regularly prior to the acute attack.
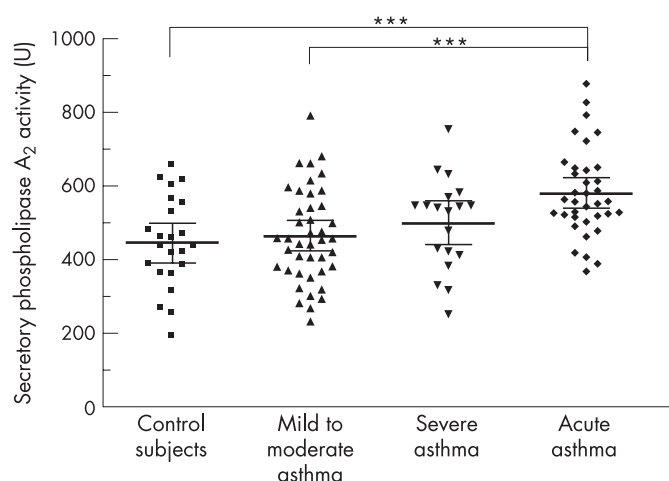BMI, body mass index; FEV₁, forced expiratory volume in 1 s; ICS, inhaled corticosteroid.

Figure 1 Plasma secretory phospholipase A_2 (sPLA$_2$) activity in control subjects (n = 23) and in patients with mild to moderate (n = 42), severe (n = 19) or acute asthma (n = 36). Horizontal bars indicate mean values and 95% CI. sPLA$_2$ activity was significantly greater in patients with an acute attack compared with controls and compared with patients with mild to moderate asthma. ***p<0.001.

Figure 2 Plasma secretory phospholipase A_2 (sPLA$_2$) activity in 22 patients with an acute exacerbation of asthma, and in the same patients after they had recovered from the acute exacerbation. Mean sPLA$_2$ activity during the acute episode (536.3 U; 95% CI 494.9 to 577.7) was not significantly different to that after recovery (540.2 U; 95% CI 506.0 to 574.4).

were not corrected for recovery, which was ~90%. The inter-analysis precision coefficient of variation was 5.4%.

Data analysis and statistics

Data are expressed as means with 95% confidence interval (CI), SD or SEM. Differences between group mean values were assessed for statistical significance by one way ANOVA followed by Bonferroni's multiple comparison test or by the unpaired t test as appropriate. In univariate analyses, Pearson's correlation was used to assess the statistical significance of associations between parameters of interest. In order to identify significant independent predictors of sPLA$_2$ activity after adjusting for confounding factors, and to assess interactions, a multiple linear regression analysis was performed. The independent variables entered into the model were gender, age, BMI, severe asthma, atopy, use of inhaled corticosteroids, smoking status, and plasma cholesterol and vitamin C concentrations. In addition, interactions between gender and cholesterol concentration, gender and vitamin C concentration and BMI and cholesterol concentration were assessed. All statistical analyses were performed using SPSS for Windows V.11.5 (SPSS Inc, Chicago, Illinois, USA) and a p value <0.05 was considered significant.

RESULTS

The characteristics of the patients with asthma and the control subjects are presented in table 1. Plasma sPLA$_2$ activity in patients with stable, mild to moderate (464.8 U; 95% CI 425.3 to 504.3) or severe asthma (499.9 U; 95% CI 439.4 to 560.4) did not differ significantly from that in control subjects (445.7 U; 95% CI 392.1 to 499.4) (fig 1). However, among these control subjects and patients with stable asthma, sPLA$_2$ activity was significantly greater in atopic (483.5 U; 95% CI 455.3 to 511.6) than in non-atopic subjects (419.7 U; 95% CI 349.6 to 489.7; p<0.05) and in males (501.3 U; 95% CI 455.4 to 547.3) compared with females (442.1 U; 95% CI 409.4 to 474.8; p<0.05). sPLA$_2$ activity in the 15 patients with asthma who were not using inhaled corticosteroids (516.5 U; 95% CI 452.0 to 581.1) was not significantly different compared with that in the 46 patients who were (462.4 U; 95% CI 424.5 to 500.4), and
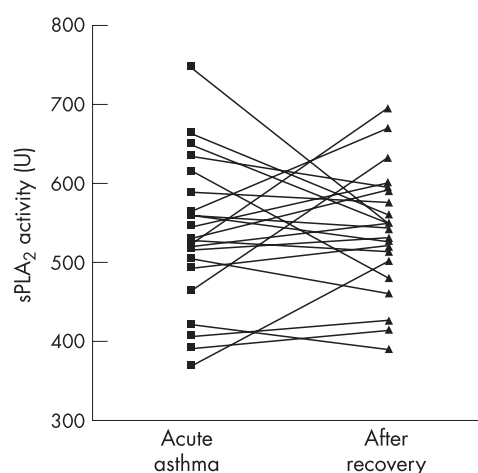
there was no correlation between inhaled corticosteroid dose and sPLA$_2$ activity.

sPLA$_2$ activity in patients presenting with an acute exacerbation of asthma (581.6 U; 95% CI 541.2 to 622.0) was significantly higher than that in control subjects or in patients with mild to moderate asthma (p<0.001) (fig 1). However, mean plasma sPLA$_2$ activity measured in 22 of the 36 patients after recovery from the acute attack (540.2 U; 95% CI 506.0 to 574.4) was not different to that measured during the acute asthma episode (536.3 U; 95% CI 494.9 to 577.7) (fig 2). While sPLA$_2$ activity decreased substantially after recovery in some patients, it increased or remained unchanged in others. There was no correlation between inhaled corticosteroid dose and sPLA$_2$ activity during or after recovery from the acute attack.

Mean BMI in patients with stable, severe asthma (28.8; 95% CI 25.4 to 32.1) was significantly higher than that of control subjects
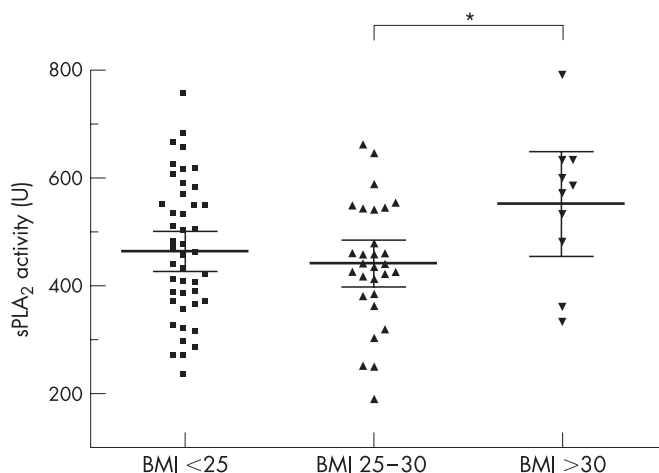

Figure 3 Plasma secretory phospholipase A_2 (sPLA$_2$) activity in control subjects and in patients with mild to moderate and severe asthma with a body mass index (BMI) <25 (n = 45), BMI 25–30 (n = 29) and BMI >30 (n = 10). Horizontal bars indicate mean values and 95% CI. sPLA$_2$ activity was greater in subjects with BMI >30 compared with those with BMI 25–30. *p<0.05.
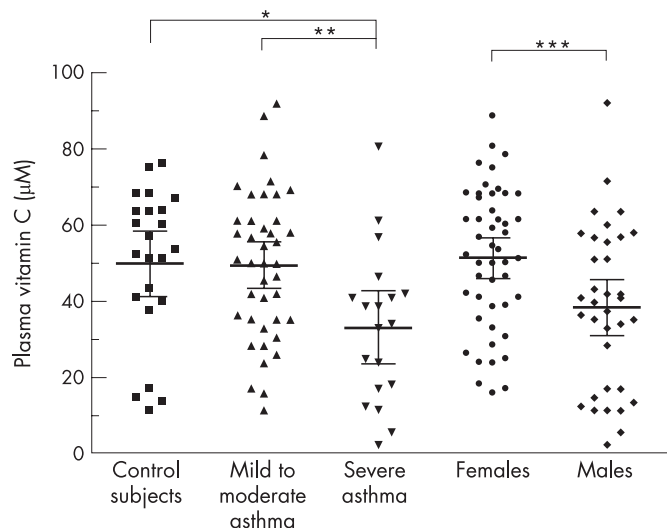
Figure 4 Plasma vitamin C concentration in control subjects (n = 23) and in patients with mild to moderate (n = 41) and severe asthma (n = 19), and in female (n = 48) and male (n = 35) control, mild to moderate and severe asthmatic subjects. Horizontal bars indicate mean values and 95% CI. Vitamin C concentration was significantly lower in patients with severe asthma compared with controls (*p = 0.02) and compared with patients with mild to moderate asthma (**p = 0.01), and in male compared with female subjects (***p<0.005).

(23.8; 95% CI 22.3 to 25.4; p<0.02). sPLA$_2$ activity was significantly greater in subjects who were clinically obese (BMI >30: 552.4 U; 95% CI 455.6 to 649.1) compared with those who were overweight (BMI 25–30: 442.1; 95% CI 399.2 to 485.0; p<0.05) (fig 3). Plasma total cholesterol concentration was significantly higher in subjects with BMI >25 (5.78 mM; 95% CI 5.42 to 6.13) compared with those with BMI ⩽25 (5.24 mM; 95% CI 4.98 to 5.50; p<0.02). Plasma cholesterol also tended to be higher in patients with severe asthma (5.96 mM; 95% CI 5.46 to 6.46) compared with controls (5.26 mM; 95% CI 4.89 to 5.63) or patients with mild to moderate asthma (5.41 mM; 95% CI 5.09 to 5.73), although the differences were not statistically significant.

Plasma vitamin C concentration was significantly lower in patients with severe asthma (33.1 μM; 95% CI 23.4 to 42.8) compared with those with mild to moderate asthma (49.6; 95% CI 43.5 to 55.6; p = 0.01) and control subjects (49.9; 95% CI 41.4 to 58.5; p = 0.02) (fig 4). Vitamin C concentration was also lower in males (38.4 μM; 95% CI 31.1 to 45.7) compared with females (51.4 μM; 95% CI 46.0 to 56.7; p<0.005) (fig 4).

Univariate analysis of the data showed weak but statistically significant positive correlations between sPLA$_2$ activity and BMI (r = 0.23; p<0.05) and also between sPLA$_2$ activity and cholesterol (r = 0.26; p<0.02) (fig 5A) in control subjects and in those with stable asthma. The correlation between sPLA$_2$ activity and cholesterol was, however, stronger in patients with severe asthma (r = 0.62; p = 0.005). In contrast with these positive associations, sPLA$_2$ activity was inversely correlated with plasma vitamin C concentration (r = −0.33; p = 0.002) (fig 5B).

A multiple linear regression analysis to identify factors independently associated with sPLA$_2$ activity in the control subjects and in patients with stable asthma showed that male gender and higher plasma cholesterol were independently associated with greater sPLA$_2$ activity (table 2). In addition, there were indications that atopy and increased BMI, although not reaching statistical significance in this model, were associated with increased sPLA$_2$ activity. Although there was
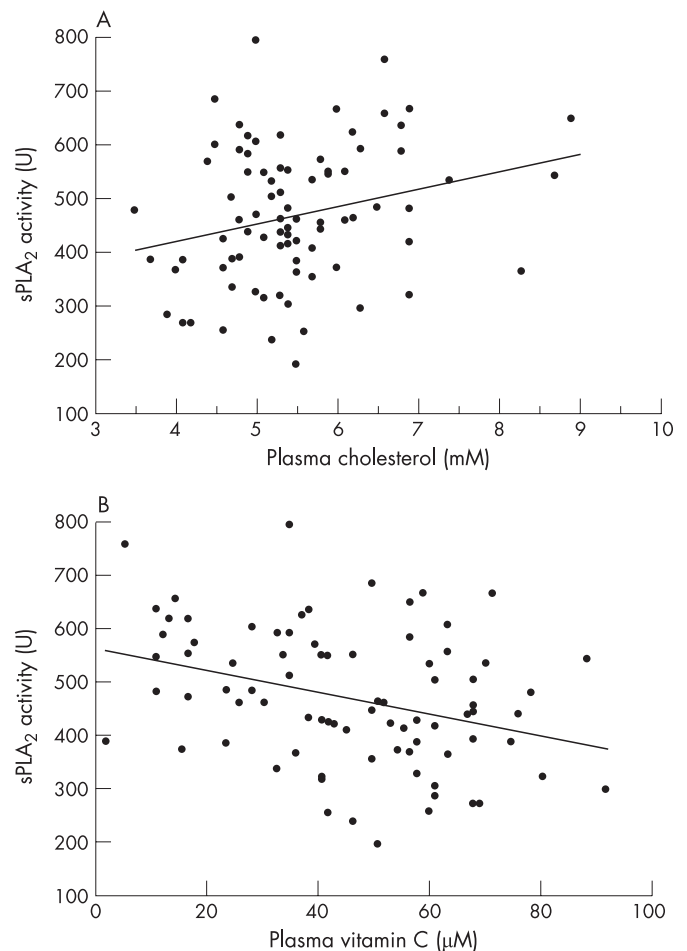

Figure 5 (A) Correlation between secretory phospholipase A$_2$ (sPLA$_2$) activity and plasma total cholesterol concentration in control subjects and in patients with stable asthma (n = 83, r = 0.26; p<0.02). (B) Inverse correlation between sPLA$_2$ activity and plasma vitamin C concentration in control subjects and in patients with stable asthma (n = 83, r = −0.33; p = 0.002).

no independent association between sPLA$_2$ activity and plasma vitamin C, there were significant interactions between gender and plasma vitamin C and also between gender and plasma cholesterol in relation to sPLA$_2$ activity.

DISCUSSION

Cytosolic PLA$_2$ plays a major role in the production of arachidonic acid derived lipid mediators, such as prostaglandins and leucotrienes, that contribute to airway inflammation in asthma.[1] However, recent evidence suggests that secretory PLA$_2$ isozymes may also have an important role in eicosanoid synthesis and amplification of the inflammatory process, both directly and by functional coupling with cytosolic PLA$_2$ and cyclooxygenase-2.[27–29] In the present study, the activity of potentially proinflammatory plasma sPLA$_2$ enzymes with heparin binding domains was measured in patients with stable asthma and patients presenting with acute asthma, in whom upregulation of inflammatory pathways might be expected. We previously measured "total" sPLA$_2$ activity in unpurified serum samples[25] whereas in the present study samples were purified on heparin affinity columns and the sPLA$_2$ activity measured was therefore much less. However, it was more likely to represent the activities of groups IIA, IID and V sPLA$_2$ that interact with

Table 2 Multiple linear regression analysis for independent predictors of secretory phospholipase A₂ activity

Variable	Coefficient (B) (95% CI)	Standardised coefficient	p Value
Gender (1 = M, 0 = F)	496.7 (158.8 to 834.5)	1.953	0.005
Cholesterol (mM)	156.1 (11.9 to 300.2)	1.243	0.034
Atopy (1 = yes, 0 = no)	56.0 (−5.1 to 117.1)	0.194	0.072
BMI (kg/m²)	26.6 (−3.6 to 56.7)	1.223	0.083
Age (y)	−1.92 (−4.24 to 0.41)	−0.188	0.105
Smoking (1 = yes, 0 = no)	−64.5 (−150.4 to 21.5)	−0.16	0.139
Use of ICS (1 = yes, 0 = no)	−22.2 (−82.7 to 38.4)	−0.088	0.468
Severe asthma (1 = yes, 0 = no)	20.3 (−55.1 to 95.7)	0.068	0.593
Vitamin C (μM)	0.09 (−1.88 to 2.05)	0.014	0.93
Interaction terms			
Gender×cholesterol	−59.9 (−113.4 to −6.4)	−1.335	0.029
Gender×vitamin C	−2.9 (−5.52 to −0.28)	−0.538	0.031
BMI×cholesterol	−4.3 (−9.79 to 1.22)	−1.492	0.125

BMI, body mass index; ICS, inhaled corticosteroids.

heparan sulphate proteoglycans in lipid rafts, bringing them into contact with enzymes such as cyclooxygenase and 5-lipoxygenase that are important for the generation of eicosanoid mediators in asthma.[4]

The observation that plasma sPLA₂ activity was not significantly increased in patients with stable, severe asthma compared with those with mild to moderate asthma and control subjects may suggest that this sPLA₂ activity is not proinflammatory or that inflammatory pathways were not upregulated in these patients with stable, severe asthma. However, there was a trend towards increased sPLA₂ activity in patients with severe asthma, and a power calculation indicated that 80 subjects would have been required in each group for the measured difference between controls and patients with severe asthma to be considered statistically significant. Therefore, another explanation for the lack of a significant difference in sPLA₂ activity between these two groups is that the study was insufficiently powered. There is one previous report that serum PLA₂ activity was significantly increased in patients with stable asthma.[16] However, in that study, it is likely that total sPLA₂ activity was measured, rather than the activity of heparin binding isoforms, and it is possible that other isoforms of sPLA₂ excluded by heparin affinity purification may be differentially expressed in patients with asthma. Furthermore, the patients in that study had not received any corticosteroid treatment, which may reduce sPLA₂ activity in patients with stable asthma, either directly by inhibiting sPLA₂ expression[30] or indirectly by moderating inflammation.[31]

The possibility that there was a real increase in sPLA₂ activity in patients with severe asthma, but that the present study was inadequately powered to detect this increase, is supported by the finding of a significant increase in plasma sPLA₂ activity in patients experiencing an acute exacerbation of asthma. This increase in plasma sPLA₂ activity in patients with acute asthma is also consistent with previous findings in patients with acute appendicitis, rheumatoid arthritis and coronary artery disease.[6 32 33] The link between sPLA₂ and inflammation is further supported by the finding in the present study of a probable association between atopy and increased sPLA₂ activity. Atopy may be associated with an increase in airway and/or systemic inflammation, as evidenced by the increase in exhaled nitric oxide in atopic subjects.[34] The mechanisms that might lead to an increase in plasma sPLA₂ activity and the source of this enzyme are uncertain, but one possibility is that sPLA₂ is an acute phase protein, released as a consequence of systemic inflammation, together with other molecules, such as

interleukin 6 (IL6) and C reactive protein.[27 35] Increased sPLA₂ activity may also result from localised production and release of sPLA₂ in the respiratory tract, as indicated by the finding of increased sPLA₂ activity in bronchoalveolar lavage fluid after allergen challenge of patients with asthma.[19] In addition, eosinophils contain significant amounts of sPLA2,[8] which may be secreted in the lungs and peripheral blood of patients with asthma. However, a recent trial showed that a sPLA₂ inhibitor had no effect on early or late forced expiratory volume in 1 s following allergen challenge of atopic asthmatics.[36]

It is also possible that sPLA₂ activity is genetically determined, and is high in patients who are prone to acute asthma episodes. As such, sPLA₂ may be a potential biomarker of patients who are more likely to experience acute exacerbations. Furthermore, sPLA₂ activity was not reduced in many of the 22 patients in whom it was measured after recovery from the acute episode of asthma, suggesting that treatment, including corticosteroids, may not suppress inflammatory processes such as sPLA₂ activity in all patients with asthma.

The previously observed correlation between sPLA₂ activity and total cholesterol[25] was confirmed in the present study, but the correlation was much stronger in patients with severe asthma. In addition, mean BMI was significantly higher in patients with severe asthma, while plasma cholesterol was increased in overweight subjects and sPLA₂ activity was greater in obese subjects and positively associated with increased BMI. These results are strongly suggestive of interactions between increased BMI, dysregulation of lipid metabolism and more severe asthma, lending support to the large number of recent epidemiological studies that have reported associations between the prevalence and incidence of asthma, and increased BMI in both adults and children (reviewed by Ford[21]).

There are a number of plausible mechanisms that may explain the association between increased BMI and asthma.[20] In obese subjects, adipose tissue may contribute significantly to inflammation through production of IL6, tumour necrosis factor α (TNFα), transforming growth factor β1 and eotaxin, as well as the adipokine, leptin, which has been shown to regulate T cell proliferation and activation.[20 37] The results from the present study suggest that upregulation of sPLA₂ may be a proinflammatory consequence of increases in BMI and adipose tissue metabolism, similar to the increases in other inflammation associated molecules, such as TNFα, IL6, leptin and C reactive protein, that have been observed in obese as well as asthmatic subjects.[37–40] However, it is interesting that in this study,

sPLA$_2$ activity was higher in atopic subjects whereas increased levels of C reactive protein were associated with non-allergic asthma.[40] Upregulation of sPLA$_2$ activity may have important functional consequences for inflammatory processes in both obesity and asthma. Specifically, lipoprotein associated sPLA$_2$ may increase the catabolism of phospholipids to biologically active lipids, including prostaglandins, leucotrienes, platelet activating factor and lysophospholipids, which are likely to play a role in chronic inflammation in asthmatic and obese subjects. In addition, sPLA2, like leptin, has been implicated in T cell proliferation and cytokine and chemokine production.[11–15]

sPLA$_2$ activity was inversely correlated with plasma vitamin C concentration, although the latter was not an independent predictor of lower sPLA$_2$ activity. Importantly, however, there was a marked gender interaction, with sPLA$_2$ activity being higher and vitamin C concentration lower in males compared with females. Lower plasma vitamin C concentrations may contribute to increased oxidative modification of lipoproteins[23] and a resultant increase in the activity of enzymes such as sPLA$_2$ that catalyse the hydrolysis of lipoproteins.[24] In contrast, the multiple regression analysis performed in the present study indicated that the positive association between sPLA$_2$ activity and plasma cholesterol was stronger in females than in males. This is in keeping with observations from a number of studies suggesting a stronger association between asthma and obesity in females than in males.[20 21] Factors such as the influence of female sex hormones,[20] low dietary intake of vitamin C by male asthmatics and lower plasma concentrations of vitamin C in severe asthma,[41] are likely to be confounding factors in the associations among asthma, increased BMI, sPLA$_2$ activity and cholesterol and vitamin C concentrations.

In conclusion, this study has demonstrated a significant upregulation of plasma sPLA$_2$ activity in patients with acute asthma. In addition, the study has identified potentially important associations between sPLA$_2$ activity, increased plasma cholesterol, increased BMI and atopy, as well as gender interactions in the positive association of cholesterol and the inverse association of vitamin C with sPLA$_2$ activity. sPLA$_2$ may therefore be one of the biological links between asthma, inflammation, increased BMI, lipid metabolism and antioxidants. Further studies, possibly with larger numbers of subjects, are required to unravel these complex interactions and the mechanisms by which they may contribute to the pathophysiology and increasing prevalence of both asthma and obesity.

REFERENCES

1. **Dennis EA.** Phospholipase A2 in eicosanoid generation. *Am J Respir Crit Care Med* 2000;**161**:S32–5.
2. **Dennis EA.** The growing phospholipase A2 superfamily of signal transduction enzymes. *Trends Biochem Sci* 1997;**22**:1–2.
3. **Balsinde J,** Balboa MA, Insel PA, *et al.* Regulation and inhibition of phospholipase A2. *Annu Rev Pharmacol Toxicol* 1999;**39**:175–89.
4. **Triggiani M,** Granata F, Giannattasio G, *et al.* Secretory phospholipases A2 in inflammatory and allergic diseases: not just enzymes. *J Allergy Clin Immunol* 2005;**116**:1000–6.
5. **Nevalainen TJ.** Serum phospholipases A2 in inflammatory diseases. *Clin Chem* 1993;**39**:2453–9.
6. **Lin MK,** Farewell V, Vadas P, *et al.* Secretory phospholipase A2 as an index of disease activity in rheumatoid arthritis. Prospective double blind study of 212 patients. *J Rheumatol* 1996;**23**:1162–6.
7. **Djukanovic R.** Asthma: a disease of inflammation and repair. *J Allergy Clin Immunol* 2000;**105**:S522–6.
8. **Blom M,** Tool AT, Wever PC, *et al.* Human eosinophils express, relative to other circulating leukocytes, large amounts of secretory 14-kD phospholipase A2. *Blood* 1998;**91**:3037–43.
9. **Perrin-Cocon L,** Agaugue S, Coutant F, *et al.* Secretory phospholipase A2 induces dendritic cell maturation. *Eur J Immunol* 2004;**34**:2293–302.
10. **Ramoner R,** Putz T, Gander H, *et al.* Dendritic-cell activation by secretory phospholipase A2. *Blood* 2005;**105**:3583–7.
11. **Tessier C,** Hichami A, Khan NA. Implication of three isoforms of PLA(2) in human T-cell proliferation. *FEBS Lett* 2002;**520**:111–16.
12. **Triggiani M,** Granata F, Oriente A, *et al.* Secretory phospholipases A2 induce cytokine release from blood and synovial fluid monocytes. *Eur J Immunol* 2002;**32**:67–76.
13. **Triggiani M,** Granata F, Balestrieri B, *et al.* Secretory phospholipases A2 activate selective functions in human eosinophils. *J Immunol* 2003;**170**:3279–88.
14. **Jo EJ,** Lee HY, Lee YN, *et al.* Group IB secretory phospholipase A2 stimulates CXC chemokine ligand 8 production via ERK and NF-kappa B in human neutrophils. *J Immunol* 2004;**173**:6433–9.
15. **Granata F,** Petraroli A, Boilard E, *et al.* Activation of cytokine production by secreted phospholipase A2 in human lung macrophages expressing the M-type receptor. *J Immunol* 2005;**174**:464–74.
16. **Kashima N,** Nakajima H, Katsura T, *et al.* Study of serum phospholipase A2 activity in bronchial asthmatic patients. *Arerugi* 1993;**42**:723–7.
17. **Mehta D,** Gupta S, Gaur SN, *et al.* Increased leukocyte phospholipase A2 activity and plasma lysophosphatidylcholine levels in asthma and rhinitis and their relationship to airway sensitivity to histamine. *Am Rev Respir Dis* 1990;**142**:157–61.
18. **Stadel JM,** Hoyle K, Naclerio RM, *et al.* Characterization of phospholipase A2 from human nasal lavage. *Am J Respir Cell Mol Biol* 1994;**11**:108–13.
19. **Bowton DL,** Seeds MC, Fasano MB, *et al.* Phospholipase A2 and arachidonate increase in bronchoalveolar lavage fluid after inhaled antigen challenge in asthmatics. *Am J Respir Crit Care Med* 1997;**155**:421–5.
20. **Beuther DA,** Weiss ST, Sutherland ER. Obesity and asthma. *Am J Respir Crit Care Med* 2006;**174**:112–19.
21. **Ford ES.** The epidemiology of obesity and asthma. *J Allergy Clin Immunol* 2005;**115**:897–909.
22. **Brown CD,** Higgins M, Donato KA, *et al.* Body mass index and the prevalence of hypertension and dyslipidemia. *Obes Res* 2000;**8**:605–19.
23. **Carr AC,** Zhu BZ, Frei B. Potential antiatherogenic mechanisms of ascorbate (vitamin C) and alpha-tocopherol (vitamin E). *Circ Res* 2000;**87**:349–54.
24. **Pruzanski W,** Stefanski E, de Beer FC, *et al.* Lipoproteins are substrates for human secretory group IIA phospholipase A2: preferential hydrolysis of acute phase HDL. *J Lipid Res* 1998;**39**:2150–60.
25. **Petrovic N,** Grove C, Langton PE, *et al.* A simple assay for a human serum phospholipase A2 that is associated with high-density lipoproteins. *J Lipid Res* 2001;**42**:1706–13.
26. **National Institutes of Health**. *National Asthma Education and Prevention Program Expert Panel Report II. Guidelines for the diagnosis and management of asthma.* Bethesda: National Institutes of Health, 1997 NIH Publication No. 97-4051.
27. **Touqui L,** Alaoui-El-Azher M. Mammalian secreted phospholipases A2 and their pathophysiological significance in inflammatory diseases. *Curr Mol Med* 2001;**1**:739–54.
28. **Kim YJ,** Kim KP, Han SK, *et al.* Group V phospholipase A2 induces leukotriene biosynthesis in human neutrophils through the activation of group IVA phospholipase A2. *J Biol Chem* 2002;**277**:36479–88.
29. **Balsinde J,** Shinohara H, Lefkowitz LJ, *et al.* Group V phospholipase A(2)-dependent induction of cyclooxygenase-2 in macrophages. *J Biol Chem* 1999;**274**:25967–70.
30. **Nakano T,** Ohara O, Teraoka H, *et al.* Glucocorticoids suppress group II phospholipase A2 production by blocking mRNA synthesis and post-transcriptional expression. *J Biol Chem* 1990;**265**:12745–8.
31. **Barnes PJ.** Inhaled glucocorticoids for asthma. *N Engl J Med* 1995;**332**:868–75.
32. **Gronroos JM,** Forsstrom JJ, Irjala K, *et al.* Phospholipase A2, C-reactive protein, and white blood cell count in the diagnosis of acute appendicitis. *Clin Chem* 1994;**40**:1757–60.
33. **Kugiyama K,** Ota Y, Takazoe K, *et al.* Circulating levels of secretory type II phospholipase A(2) predict coronary events in patients with coronary artery disease. *Circulation* 1999;**100**:1280–4.
34. **Franklin PJ,** Stick SM, Le Souef PN, *et al.* Measuring exhaled nitric oxide levels in adults: the importance of atopy and airway responsiveness. *Chest* 2004;**126**:1540–5.
35. **Wouters EFM.** The systemic face of airway diseases: the role of C-reactive protein. *Eur Respir J* 2006;**27**:877–9.
36. **Bowton DL,** Dmitrienko AA, Israel E, *et al.* Impact of a soluble phospholipase A2 inhibitor on inhaled allergen challenge in subjects with asthma. *J Asthma* 2005;**42**:65–71.
37. **Fantuzzi G.** Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol* 2005;**115**:911–19.
38. **Sood A,** Ford ES, Camargo CA Jr. Association between leptin and asthma in adults. *Thorax* 2006;**61**:300–5.
39. **Ford ES.** Asthma, body mass index, and C-reactive protein among US adults. *J Asthma* 2003;**40**:733–9.
40. **Olafsdottir IS,** Gislason T, Thjodleifsson B, *et al.* C reactive protein levels are increased in non-allergic but not allergic asthma: a multicentre epidemiological study. *Thorax* 2005;**60**:451–4.
41. **Misso NL,** Brooks-Wildhaber J, Ray S, *et al.* Plasma concentrations of dietary and nondietary antioxidants are low in severe asthma. *Eur Respir J* 2005;**26**:257–64.