

Transforming growth factor β_2 induced pleurodesis is not inhibited by corticosteroids

Y C G Lee, C J Devin, L R Teixeira, J T Rogers, P J Thompson, K B Lane, R W Light

Abstract

Background—Talc and tetracyclines induce pleurodesis by directly injuring the pleura. The injury results in intense inflammation which subsequently leads to fibrosis. Corticosteroids can inhibit talc pleurodesis by reducing the inflammatory process. We hypothesised that transforming growth factor β_2 (TGF β_2), a fibrogenic cytokine with immunomodulatory functions, could induce effective pleurodesis without generating significant pleural inflammation and therefore remain effective despite co-administration of corticosteroids.

Methods—Thirty rabbits were divided into two groups. Rabbits in the steroid group received weekly intramuscular injections of triamcinolone diacetate (0.8 mg/kg). Ten rabbits in each group were given 5.0 μ g TGF β_2 intrapleurally via a chest tube while the remaining five received 1.7 μ g TGF β_2 . Pleurodesis was graded macroscopically after 14 days from 1 (none) to 8 (>50% symphysis).

Results—TGF β_2 produced excellent pleurodesis at both 5.0 μ g and 1.7 μ g doses. The pleural effusions produced after the injection were low in all inflammatory markers. No significant differences were seen between the steroid group and controls in macroscopic pleurodesis scores (7.2 (1.3) v 7.1 (1.2)), levels of inflammatory markers in the pleural fluids (leucocyte 1107 (387)/mm³ v 1376 (581)/mm³; protein 3.1 (0.3) mg/dl v 2.9 (0.3) mg/dl, and LDH 478 (232) IU/l v 502 (123) IU/l), and the degree of microscopic pleural fibrosis and pleural inflammation.

Conclusions—TGF β_2 can induce effective pleurodesis and remains effective in the presence of high dose parenteral corticosteroids.

(Thorax 2001;56:643-648)

Keywords: transforming growth factor β ; pleurodesis; corticosteroids; pleural effusion

Pleurodesis remains an important treatment option in the management of malignant pleural effusions and recurrent pneumothorax. Commonly used pleurodesing agents such as talc and tetracycline derivatives produce pleurodesis by inducing acute pleural injury and hence inflammation and fibrosis.¹ This intense inflammatory process is essential to pleurodesis as we recently showed that co-administration of high dose parenteral corticosteroids reduced the inflammation and inhibited talc pleurodesis.² It is also likely that chest pain and fever,

common side effects after chemical pleurodesis, are the results of intense pleural inflammation.

The ideal pleurodesing agent would be one that can produce fibrosis without inducing pleural inflammation. Transforming growth factor beta (TGF β), with its unique capabilities of being a strong fibrogenic activator as well as a potent anti-inflammatory cytokine, provides an attractive option as a novel treatment for producing pleurodesis.

TGF β regulates fibrosis by increasing extracellular matrix formation and plays a vital role in fibrotic processes, including pulmonary fibrosis and wound healing.³ It is also a potent anti-inflammatory cytokine and can downregulate tumour necrosis factor α (TNF α) and inhibit lymphocyte function.⁴ We have recently reported that a single intrapleural injection of TGF β_2 produces excellent pleurodesis in rabbits^{5,6} and in sheep.⁷ The effusion produced was significantly less inflammatory than those resulting from talc or doxycycline.

In the present study we have investigated the effect of high dose systemic corticosteroids on TGF β_2 induced pleurodesis. We hypothesised that TGF β_2 produces pleurodesis without inducing significant pleural inflammation and hence systemic corticosteroids would have little or no inhibitory effect on TGF β_2 induced pleurodesis.

Methods

TRANSFORMING GROWTH FACTOR β_2

A recombinant human TGF β_2 (Genzyme Corporation, Framingham, MA, USA) produced in Chinese hamster ovary cells was used. TGF β_2 was formulated in a vehicle consisting of 20 mM sodium phosphate, 130 mM sodium chloride, 15% (w/w) propylene glycol, and 20% (w/w) polyethylene glycol 400. The pH of the solution was 7.2. The vehicle was prepared using USP/NF grade reagents in water for injection and sterile filtered through a 0.2 μ m filter. The TGF β_2 concentration was determined by the manufacturer with a sandwich enzyme linked immunosorbent assay using two monoclonal antibodies that cross react with both TGF β_2 and TGF β_3 , and the activity of TGF β_2 was determined using a mink lung cell (Mv1Lu) antiproliferation assay modified from the method described by Ogawa.⁸

ANIMAL EXPERIMENT

The method used was similar to that described in our previous studies.^{5,6,9} The study protocol was approved by the Vanderbilt University Institutional Animal Care and Use Committee. New Zealand white rabbits weighing 1.5–2.0 kg were anaesthetised with an intramuscular injection of

Departments of
Pulmonary Medicine,
St Thomas Hospital
and Vanderbilt
University, Nashville,
Tennessee, USA

Y C G Lee
C J Devin
J T Rogers
K B Lane
R W Light

Department of
Medicine, University
of Western Australia,
Perth, Australia

Y C G Lee
P J Thompson

Division of
Respiratory Diseases,
Heart Institute
(InCor), University of
São Paulo Medical
School, São Paulo,
Brazil

L R Teixeira

Correspondence to:
Dr Y C G Lee, Department
of Pulmonary Medicine, St
Thomas Hospital, 4220
Harding Road, Nashville,
TN 37202, USA
ycgarylee@hotmail.com

Received 6 September 2000
Returned to authors
18 December 2000
Revised version received
24 January 2001
Accepted for publication
9 May 2001

35 mg/kg ketamine hydrochloride (Fort Dodge Animal Health, Iowa, USA) and 5 mg/kg xylazine hydrochloride (Fermenta, Kansas City, MO, USA). The chest was shaven and the skin sterilised with 10% povidone iodine (Baxter, Deerfield, IL, USA). The rabbit was placed in the lateral decubitus position and a small (<3 cm) skin incision was made midway between the tip of the scapula and the sternum approximately 2 cm above the costal margin. Chest tubes were made from intravenous solution set tubes (Baxter, Deerfield, IL, USA) with three extra openings near the distal end of the tube to enhance drainage. The chest tube was inserted by blunt dissection into the right pleural cavity while the left was used for control. The chest tube was secured at the muscle layers with purse string sutures. The proximal end of the chest tube was then tunnelled underneath the skin and drawn out through the skin posteriorly and superiorly between the two scapulae. The exterior end of the chest tube was sealed with a one way valve with cap (Medexinc, Hilliard, OH, USA) via an adapter and sutured to the skin. A three way stopcock was attached to the end of the chest tube through which any aspirated air was evacuated from the pleural space.

Thirty rabbits were divided into two groups of 15. Rabbits in the steroid group received an intramuscular injection of triamcinolone diacetate (Fujisawa Inc, Deerfield, IL, USA) 0.8 mg/kg at the time of chest tube insertion and then weekly afterwards. Rabbits in the control group received no steroid injections. All rabbits received an intrapleural injection of TGF β_2 (Genzyme Corporation, Framingham, MA, USA) on the day following the chest tube insertion. Ten rabbits in each group received a TGF β_2 dose of 5.0 μ g in 2.5 ml while the remaining five in each group received a lower dose of 1.7 μ g in 2.5 ml via the chest tube. This was followed by the injection of 1.0 ml of 0.9% sodium chloride (Baxter, Deerfield, IL, USA) to clear the dead space of the chest tube. The results of the five rabbits that received 1.7 μ g TGF β_2 without steroid have previously been included in another study.¹⁰ These doses of TGF β_2 were chosen as they were shown to be the lowest effective doses in our previous studies.^{5,6}

After the intrapleural injection the chest tube was aspirated at 24 hour intervals for any pleural fluid. The chest tube was withdrawn under light sedation if the pleural fluid drainage was <5 ml over the preceding 24 hours. Rabbits that developed signs of inflammation or infection at the skin wound were given daily intramuscular injections of 12.5 mg gentamicin (Fermenta, Kansas City, MO, USA) for two consecutive days. Rabbits that showed evidence of reduced oral intake or dehydration after the anaesthesia were given subcutaneous injections of 50 ml 5% dextrose and 50 ml 0.9% sodium chloride (Baxter, Deerfield, IL, USA) every 24 hours.

The volume of the pleural fluid collected was recorded. The total leucocyte count was measured using an automated counter (Coulter Electronics, Luton, UK) which was

Grading of pleurodesis score

The degree of pleurodesis was graded on a scale of 1 to 8. Adhesions were defined as fibrous connections between the visceral and parietal pleurae. Symphysis was present if the visceral and parietal pleurae were difficult to separate as a result of adhesions.

- 1 = no adhesions between the visceral and parietal pleurae
- 2 = rare adhesions between the visceral and parietal pleurae with no symphysis
- 3 = scattered adhesions between the visceral and parietal pleurae with no symphysis
- 4 = many adhesions between the visceral and parietal pleurae with no symphysis
- 5 = many adhesions between the visceral and parietal pleurae with symphysis involving <5% of the hemithorax
- 6 = many adhesions between the visceral and parietal pleurae with symphysis involving 5–25% of the hemithorax
- 7 = many adhesions between the visceral and parietal pleurae with symphysis involving 25–50% of the hemithorax
- 8 = many adhesions between the visceral and parietal pleurae with symphysis involving >50% of the hemithorax

Grading of haemothorax

- 0 = no evidence of haemothorax
- 1 = haemothorax involving <15% of the hemithorax
- 2 = haemothorax involving 15–33% of the hemithorax
- 3 = haemothorax involving 33–75% of the hemithorax
- 4 = haemothorax involving >75% of the hemithorax

Box 1 Grading of pleurodesis scores and haemothorax.

calibrated daily. The first reading was ignored and the mean of the next three readings was recorded. The protein, glucose, and lactate dehydrogenase (LDH) levels were determined with an automated analyser (Johnson & Johnson, Rochester, NY, USA). The upper limit of normal serum LDH concentrations in our laboratory is 618 IU/l.

At day 14 the rabbits were sedated and euthanised with carbon dioxide. The thorax was removed en bloc. The lungs were expanded by the injection of 50 ml 10% neutral buffered formalin into the exposed trachea via a plastic catheter. The trachea was then ligated with a silk suture and the entire thorax submerged into 10% neutral buffered formalin solution for at least 48 hours.

PLEURODESIS SCORING SCHEME

Necropsy was performed by two blinded investigators (KBL and RWL). A consensus grading was reached by the investigators on the degree of pleurodesis and on the extent of any haemothorax using the semi-quantitative scheme shown in box 1. Any evidence of infection or empyema was also assessed and recorded if present.

Table 1 Macroscopic pleurodesis scores in different subgroups

	TGFβ ₂ (5.0 µg) + steroid	TGFβ ₂ (5.0 µg) only	p value	TGFβ ₂ (1.7 µg) + steroid	TGFβ ₂ (1.7 µg) only	p value
n	10	10		5	5	
Pleurodesis score	7.3 (1.3)	7.3 (1.3)	NS	6.8 (1.3)	7.0 (1.2)	NS

Data are presented as mean (SD).

NS = not significant by Student's *t* test.

Table 2 Pleural fluid characteristics

	TGFβ ₂ + steroid (n=15)	TGFβ ₂ only (n=15)	p value
Effusion volume in 24 hours (ml)	28.0 (10.5)	26.7 (8.8)	NS
Total effusion volume (ml)	55.0 (26.0)	47.3 (23.0)	NS
Total leucocyte count (/mm ³)	1107 (387)	1376 (581)	NS
Protein (mg/dl)	3.1 (0.3)	2.9 (0.3)	0.02
Glucose (mg/dl)	176 (38)	182 (50)	NS
LDH (IU/l)	478 (232)	502 (123)	NS

Data are presented as mean (SD).

LDH = lactate dehydrogenase; NS = not significant by Student's *t* test.

MICROSCOPIC EXAMINATION OF THE PLEURA

At the time of necropsy, samples of the visceral pleura and lung from each hemithorax were obtained and placed in 10% neutral buffered formalin. The tissue samples were stained with haematoxylin-eosin (H&E) and picrosirius stains. The degree of microscopic inflammation and fibrosis was graded from the H&E slide by an experienced examiner (LRT) blinded to the treatment agent. The pleural inflammation and fibrosis were graded as none (0), equivocal (1), mild (2), moderate (3), or severe (4), as in our previous studies.^{2,11} The thickness of the pleura was measured using the Leica Q500IW Imaging Workstation, Image Processing and Analysis System (Leica Ltd, Cambridge, UK). With this system, the image obtained was transformed from pixels to µm. Measurements were obtained at 10 different points on three different high power fields on each sample and the mean result was reported.

Collagen fibres were subdivided into immature (thin) and mature (thick) fibres using picrosirius staining as reported by Andrade *et al.*¹² With this method the tissue blocks were sectioned at 5 µm and stained for 1 hour in a 0.2% solution of Sirius Red, Direct Red 80 (Aldrich, Milwaukee, WI, USA) dissolved in aqueous saturated picric acid. The enhancement of collagen birefringence elicited by picrosirius staining is specific for collagen and discloses its distinct patterns of physical aggregation. Immature (thin) fibres, as those present in early granulation tissue, are shown as weakly birefringent green structures while mature (thick) fibres, characteristic of mature fibrotic lesions, are identified by their strong birefringence and their yellow or red colour. The areas covered by mature and immature collagen fibres were also measured by the Leica Q500IW Imaging System (Leica Ltd, Cambridge, UK). All pixels in the image that are equivalent to, or nearly equivalent to, the colour levels of the select area were detected. The percentages of the total area of pleura covered by mature and immature fibres in the same field were measured. For each sample, readings were taken from six representative fields and the mean result used for analysis.

STATISTICAL ANALYSIS

Normality of the data was determined using the Kolmogorov-Smirnov test. The Student's *t* test (for parametric data) and Mann-Whitney rank sum test (for non-parametric data) were used to compare the values between subgroups. The pleural fluid parameters were normally distributed, while the data on pleural fibrosis and inflammation, alveolar fibrosis and inflammation, pleural thickness, and collagen fibre deposition were not normally distributed. For the pleurodesis scores, those of the 5.0 µg TGFβ₂ groups (both steroid and control groups) were normally distributed, but those of the 1.7 µg groups were not normally distributed. A *p* value of <0.05 was considered significant. Data were presented as mean (SD) unless otherwise stated. All data were analysed with a Sigma Stat V2.03 statistic software program (Jandel Scientific; San Rafael, CA, USA).

Results

Intrapleural injection of TGFβ₂ in doses of both 5.0 µg and 1.7 µg produced effective pleurodesis in rabbits with mean pleurodesis scores of 7.3 (1.3) and 6.9 (1.2), respectively. There was no difference in the pleurodesis scores between rabbits that received TGFβ₂ and systemic corticosteroids and rabbits that received TGFβ₂ alone (7.2 (1.3) and 7.1 (1.2), respectively, *p*=NS, Mann-Whitney rank sum test). The results of the rabbits in each subgroup are presented in table 1.

The injection of TGFβ₂ stimulated the production of a large quantity of pleural fluid in all rabbits. The effusion produced was relatively non-inflammatory as evidenced by the low protein and LDH levels and total leucocyte count (table 2). The production of the fluid was transient and did not hinder the development of effective pleurodesis.

Rabbits that received TGFβ₂ and corticosteroids showed no significant difference from those given only TGFβ₂ in the amount of effusion produced and the biochemical analysis of the pleural fluid by the Student's *t* test (table 2). The only exception was the effusion protein level which was higher in the steroid group (mean 3.1 mg/dl *v* 2.9 mg/dl, *p*=0.02), although the difference is unlikely to be of clinical significance. There was no difference in the other markers of inflammation (LDH level and total leucocyte count) between the two groups. The inflammatory indices of the pleural fluid from both groups of rabbits were significantly lower than previously published results of pleural fluids induced by intrapleural injections of talc, doxycycline, and mitoxantrone (table 3).

There was no difference in the effusion protein, LDH, and total leucocyte counts between rabbits given 5.0 µg TGFβ₂ and those given 1.7 µg. Interestingly, although the mean volumes of effusion at 24 hours were not different (26.6 ml in the 5.0 µg group and 28.9 ml in the 1.7 µg group), rabbits that received the higher dose of TGFβ₂ had a significantly lower amount of total effusion drained (44.0 ml *v* 65.6 ml; *p*=0.02, Student's *t* test).

Table 3 Pleural fluid characteristics in the present study compared with published data of pleural fluids induced by other pleurodesing agents

Agent	Time after injection (hours)	No of rabbits	Total leucocyte count (/mm)	Protein (mg/dl)	LDH (IU/l)
<i>Previous studies</i>					
Talc ²	6	10	9844 (1099)	3.3 (0.2)	6873 (633)
Talc ³⁵	24	12	5500 (8314)	3.9 (0.4)	6364 (4164)
Plain talc ³⁶	24	10	2900 (632)	4.3 (0.3)	N/A
Ionised talc ³⁶	24	10	3400 (3795)	4.2 (0.3)	N/A
Talc ⁶	24	15	N/A	3.0 (0.5)	12733 (5249)
Talc ⁹	24	9	26298 (14088)	4.0 (0.6)*	29593 (26172)
Doxycycline ⁹	24	8	8989 (7538)	3.7 (0.6)*	26436 (21038)
Mitoxantrone ³⁷	24	7	4580 (2370)	5.1 (0.5)	4718 (2097)
<i>Present study</i>					
TGFβ ₂ + steroid	24	15	1107 (387)	3.1 (0.3)	478 (232)
TGFβ ₂ alone	24	15	1376 (581)	2.9 (0.3)	502 (123)

Data are presented as mean (SD).

N/A = not available.

*Data not published in the paper but obtained from the authors.

There were no significant differences in the microscopic grading scores for pleural fibrosis, pleural inflammation (fig 1), or in pleural thickness (fig 2) and collagen deposition (fig 3) between pleural tissues of the rabbits that received TGFβ₂ and steroid and the tissues of rabbits given only TGFβ₂ (Mann-Whitney rank sum test). The degree of alveolar inflammation and alveolar fibrosis was minimal in both groups (fig 1).

Four rabbits died and were replaced. Three of them died from causes related to surgery such as pneumothorax or pulmonary contusion during or within 48 hours of chest tube insertion. The fourth rabbit had a blocked chest tube and died from a tension hydrothorax 3 days after the injection of TGFβ₂ (5.0 μg). Six rabbits in both groups had evidence of a haemothorax at the time of necropsy, although most of them were mild. The median (25–75% range) score of haemothorax was 0 (0.0–2.7) in rabbits given TGFβ₂ + corticosteroids and 0 (0.0–2.0) in rabbits given TGFβ₂ only. None of the rabbits had evidence of empyema at necropsic examination. The weight gain was comparable in rabbits that received corticosteroids and those that did not (0.18 (0.16) kg *v* 0.22 (0.19) kg; *p*=NS, Student's *t* test). Rabbits that received the lower dose gained more weight than rabbits that received the higher dose of

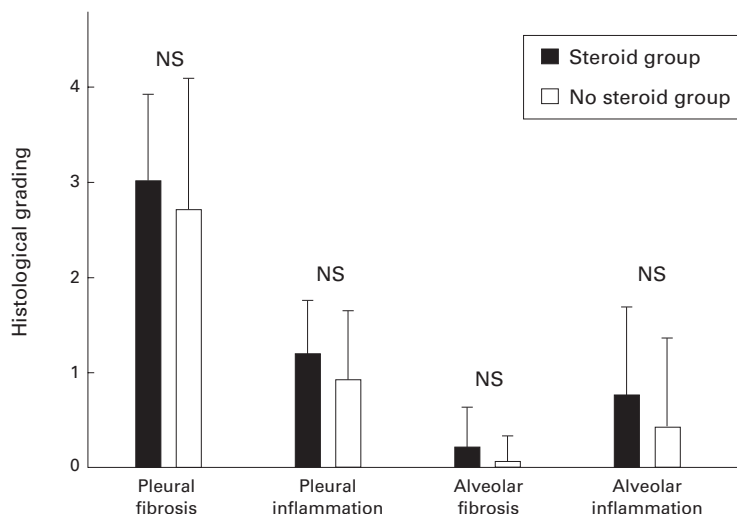


Figure 1 Comparison of histological changes in rabbits given TGFβ₂ + corticosteroids *v* rabbits given TGFβ₂ only.

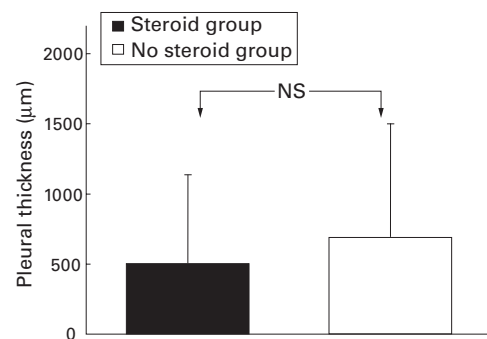


Figure 2 Pleural thickness of rabbits given TGFβ₂ + corticosteroids *v* rabbits given TGFβ₂ only.

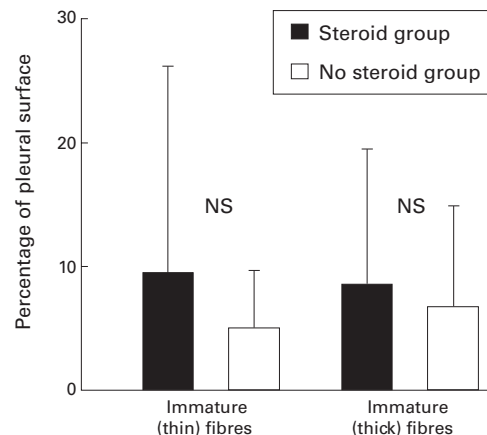


Figure 3 Collagen deposition in rabbits given TGFβ₂ + corticosteroids *v* rabbits given TGFβ₂ only.

TGFβ₂ (0.14 (0.16) kg *v* 0.34 (0.12) kg; *p*=0.001, Student's *t* test). Three rabbits (all in 5.0 μg group; two had corticosteroids) required gentamicin for skin wound infections. One rabbit (in the no steroid group) required treatment with supplementary subcutaneous fluid.

Discussion

This study shows that TGFβ₂ is effective in producing pleurodesis in a validated rabbit model. Systemic corticosteroids, at a dose that inhibits talc pleurodesis, did not affect the efficacy of TGFβ₂ induced pleurodesis nor the inflammatory indices in the effusion fluid. The study provides further evidence that TGFβ₂ induces pleurodesis via a pathway that involves minimal inflammation of the pleura and thus remains effective in the presence of high dose parenteral corticosteroids.

Conventional pleurodesing agents such as talc and tetracycline derivatives generate pleurodesis by creating acute pleural injury with subsequent inflammation and fibrosis.¹ The intense inflammation is likely to be the cause of pain and fever which commonly complicate pleurodesis. The inflammatory process is, however, essential to the pleurodesis they produce. We have previously shown that the concurrent use of corticosteroids markedly decreases the inflammatory reaction and inhibits the development of pleurodesis after the intrapleural injection of talc.²

TGFβ is a ubiquitous cytokine notable for its potent profibrotic and anti-inflammatory properties, both of which make TGFβ an attractive

option as a novel class of pleurodesing agent. Of the known growth factors, TGF β is the most potent in stimulating extracellular matrix production and also readily inhibits its degradation.⁴ Increased production and accumulation of TGF β is an important feature in fibrotic diseases including pulmonary fibrosis and glomerulosclerosis.³ At the same time, TGF β is a potent immunomodulatory cytokine. It can inhibit the production of TNF α , interleukin(IL)-1,³ and IL-8¹³ and is more potent than cyclosporin as an inhibitor of lymphocyte functions.⁴

In the present study we have confirmed that direct administration of TGF β_2 to the pleura creates excellent pleurodesis. The doses of TGF β_2 we used were the lowest effective doses that stimulated pleurodesis in our previous studies.^{5,6} We found that the pleurodesis induced by 5.0 μ g TGF β_2 was not inhibited by systemic corticosteroids at a dose that blocks talc pleurodesis. We also showed that corticosteroids did not inhibit the TGF β_2 induced pleurodesis even when we applied a lower dose of TGF β_2 (1.7 μ g) which produced lower pleurodesis scores. The results of this study provide further evidence that TGF β_2 induces pleurodesis without necessitating pleural injury and inflammation. This is also confirmed by the low inflammatory indices (protein, LDH, and leucocyte counts) in the pleural fluids induced after TGF β_2 injection compared with pleural fluids induced after the intrapleural administration of conventional pleurodesing agents such as talc, doxycycline, or mitoxantrone (table 3).

Interestingly, the intrapleural injection of TGF β_2 induced the production of large volumes of effusion. TGF β can alter the morphology of mesothelial cells in vitro and increase the permeability of the mesothelial cell monolayer.¹⁴ It is also a potent stimulator for the release of vascular endothelial growth factor (VEGF) which, in turn, increases vascular permeability.¹⁵ We found that concurrent administration of corticosteroids made no difference to the amount of fluid produced, indicating that the production of fluid after TGF β injection is unlikely to be due to inflammation. The total amount of effusion drained was significantly higher in rabbits receiving lower dose of TGF β_2 than in those receiving the higher dose, although there was no difference in the amount drained at 24 hours. We speculate that higher doses of TGF β_2 may induce more fibrosis and localizations in the first few days which impair pleural fluid drainage.

In our study rabbits that received 5 μ g TGF β_2 gained less weight than those given 1.7 μ g. Terrell *et al*¹⁶ observed that rabbits treated with a single intravenous bolus of mid or high dose human recombinant TGF β , had a transient reduction in food consumption and weight. There was an associated slight decrease in albumin and total protein on day 3 which was reversed by 2 weeks. Similar findings were observed in rats given repeated injections of high doses of intravenous TGF β ,¹⁶ or intraperitoneal TGF β_2 .¹⁷ Increased serum levels of TGF β and reduced levels of insulin-like

growth factor 1 have recently been shown in mice with poor food intake and weight loss.¹⁸ The relationship between high doses of TGF β and weight loss requires further investigation.

TGF β is multifunctional and its actions on target cells are critically dependent on the cell type, its state of differentiation, and the particular set of growth factors and hormones acting on the cell.¹⁹ Its interactions with corticosteroids are complex. While TGF β can reduce cortisol²⁰ and mineralocorticoid²¹ synthesis, glucocorticoids in turn are potent regulators of the expression of TGF β isoforms.²² These regulations are again cell and stimulus specific.^{22,23} For example, glucocorticoids enhance accumulation of TGF β mRNA in some cell lines including fibroblasts,²⁴ osteoblasts,²⁵ and T lymphocytes^{26,27} but inhibit the production of TGF β in others.^{28,29}

Our study is the first to examine the effects of corticosteroids on the action of TGF β in pleural tissues. Previous studies of the in vivo effects of corticosteroids and TGF β are limited and yielded conflicting results. In one study dexamethasone blocked the profibrotic effect of TGF β in the abdominal wounds of rats.³⁰ In another study, using a rat model of bleomycin induced pulmonary fibrosis, corticosteroids reduced the influx of inflammatory cells but the alveolar macrophage production of TGF β was not inhibited by the corticosteroids.³¹ Conversely, other authors have also demonstrated that both local^{32,33} and systemic³⁴ administrations of TGF β could reverse the steroid induced impairment in wound healing. Little, however, is known of the in vivo interactions between corticosteroids and TGF β in other organ systems.

In summary, TGF β_2 induces pleurodesis through a novel pathway whereby it induces pleural fibrosis without provoking excessive inflammation. We confirmed this by demonstrating that corticosteroids, at the same dose which blocks talc pleurodesis, did not impair the effectiveness of TGF β_2 in inducing pleurodesis. If these data can be extrapolated to humans, then TGF β_2 (as a non-inflammatory pleurodesing agent) may be less likely to induce pain and fever than talc and will remain effective in patients receiving corticosteroids.

This study was supported by the Saint Thomas Foundation, Nashville, TN, USA. We thank the Genzyme Corporation (Framingham, MA, USA) for providing the TGF β_2 used in the experiments. Dr Lee is a recipient of a United States-New Zealand Fulbright Graduate Award.

- Kennedy L, Harley RA, Sahn SA, *et al*. Talc slurry pleurodesis. Pleural fluid and histologic analysis. *Chest* 1995;107:1707-12.
- Xie C, Teixeira LR, McGovern JP, *et al*. Systemic corticosteroids decrease the effectiveness of talc pleurodesis. *Am J Respir Crit Care Med* 1998;157:1441-4.
- Border W, Noble NA. Transforming growth factor-beta in tissue fibrosis. *N Engl J Med* 1994;331:1286-92.
- Kelley J. Transforming growth factor-beta. In: Kelley J, ed. *Cytokines of the lung*. New York: Marcel Dekker, 1993:101-137.
- Light RW, Cheng DS, Lee YC, *et al*. A single intrapleural injection of transforming growth factor beta-2 produces an excellent pleurodesis in rabbits. *Am J Respir Crit Care Med* 2000;162:98-104.
- Lee YCG, Teixeira LR, Devin CJ, *et al*. Transforming growth factor- β_2 induces pleurodesis significantly faster than talc. *Am J Respir Crit Care Med* 2001;163:640-4.
- Lee YCG, Lane KB, Parker RE, *et al*. Transforming growth factor beta-2 (TGF β_2) produces effective pleurodesis in sheep with no systemic complications. *Thorax* 2000;55:1058-62.

- 8 Ogawa Y, Seyedin SM. Purification of transforming growth factor beta 1 and beta 2 from bovine and cell culture assays. *Methods Enzymol* 1991;198:317-27.
- 9 Cheng DS, Rogers JT, Wheeler A, et al. The effects of intrapleural polyclonal anti-tumor necrosis factor alpha (TNF alpha) Fab fragments on pleurodesis in rabbits. *Lung* 2000;178:19-29.
- 10 Malkerneker D, Lee YC, Devin CJ, et al. Transforming growth factor beta-2 but not fibronectin induces effective pleurodesis. *Chest* 2000;118:151S.
- 11 Vargas FS, Teixeira LR, Silva LM, et al. Comparison of silver nitrate and tetracycline as pleural sclerosing agents in rabbits. *Chest* 1995;108:1080-3.
- 12 Andrade GB, Riet-Correa F, Montes GS, et al. Dating of fibrotic lesions by the Picrosirius-polarization method. An application using the lesions of Lechiguana (bovine focal proliferative fibrogranulomatous panniculitis). *Eur J Histochem* 1997;41:203-9.
- 13 Smith WB, Noack L, Khew-Goddard Y, et al. Transforming growth factor-beta 1 inhibits the production of IL-8 and the transmigration of neutrophils through activated endothelium. *J Immunol* 1996;157:360-8.
- 14 Ikubo A, Morisaki T, Katano M, et al. A possible role of TGF-beta in the formation of malignant effusions. *Clin Immunol Immunopathol* 1995;77:27-32.
- 15 Berse B, Hunt JA, Diegel RJ, et al. Hypoxia augments cytokine (transforming growth factor-beta (TGF-beta) and IL-1)-induced vascular endothelial growth factor secretion by human synovial fibroblasts. *Clin Exp Immunol* 1999;115:176-82.
- 16 Terrell TG, Working PK, Chow CP, et al. Pathology of recombinant human transforming growth factor-beta 1 in rats and rabbits. *Int Rev Exp Pathol* 1993;34(Pt B):43-67.
- 17 Kelly FJ, Anderson S, Thompson MM, et al. Acute and chronic renal effects of recombinant human TGF-beta2 in the rat. *J Am Soc Nephrol* 1999;10:1264-73.
- 18 Tsao T, Fawcett J, Fervenza FC, et al. Expression of insulin-like growth factor-1 and transforming growth factor-beta in hypokalemic nephropathy in the rat. *Kidney Int* 2001;59:96-105.
- 19 Sporn MB, Roberts AB. Peptide growth factors are multifunctional. *Nature* 1988;332:217-9.
- 20 Feige J-J, Cochet C, Savona C, et al. Transforming growth factor beta 1: an autocrine regulator of adrenocortical steroidogenesis. *Endocrine Res* 1991;17:267-79.
- 21 Gupta P, Franco-Saenz R, Mulrow PJ. Transforming growth factor-beta 1 inhibits aldosterone biosynthesis in cultured bovine zona glomerulosa cells. *Endocrinology* 1993;132:1184-8.
- 22 Koli K, Keski-Oja J. Transforming growth factor-beta system and its regulation by members of the steroid-thyroid hormone family. *Cancer Res* 1996;70:63-94.
- 23 Almawi WY, Beyhum HN, Rahme AA, et al. Regulation of cytokine and cytokine receptor expression by glucocorticoids. *J Leukoc Biol* 1996;60:563-72.
- 24 Wang J, Kuliszewski M, Yee W, et al. Cloning and expression of glucocorticoid-induced genes in fetal rat lung fibroblasts. Transforming growth factor-beta 3. *J Biol Chem* 1995;270:2722-8.
- 25 Oursler MJ, Riggs BL, Spelsberg TC. Glucocorticoid-induced activation of latent transforming growth factor-beta by normal human osteoblast-like cells. *Endocrinology* 1993;133:2187-96.
- 26 Ayanlar-Batuman O, Ferrero AP, Diaz A, et al. Regulation of transforming growth factor-beta 1 gene expression by glucocorticoids in normal human T lymphocytes. *J Clin Invest* 1991;88:1574-80.
- 27 Correale J, Arias M, Gilmore W. Steroid hormone regulation of cytokine secretion by proteolipid protein-specific CD4+ T cell clones isolated from multiple sclerosis patients and normal control subjects. *J Immunol* 1998;161:3365-74.
- 28 Danielpour D, Kim KY, Winokur TS, et al. Differential regulation of the expression of transforming growth factor-beta 1 and 2 by retinoic acid, epidermal growth factor, and dexamethasone in NRD-49F and A549 cells. *J Cell Physiol* 1991;148:235-44.
- 29 Baumgartner RA, Deramo VA, Beaven MA. Constitutive and inducible mechanisms of synthesis and release of cytokines in immune cell lines. *J Immunol* 1996;157:4087-93.
- 30 Meisler N, Keefer KA, Ehrlich HP, et al. Dexamethasone abrogates the fibrogenic effect of transforming growth factor-beta in rat granuloma and granulation tissue fibroblasts. *J Invest Dermatol* 1997;108:285-9.
- 31 Khalil N, Whitman C, Zuo L, et al. Regulation of alveolar macrophage transforming growth factor-beta secretion by corticosteroids in bleomycin-induced pulmonary inflammation in the rat. *J Clin Invest* 1993;92:1812-8.
- 32 Slavin J, Nash JR, Kingsworth AN. Effect of transforming growth factor beta and basic fibroblast growth factor on steroid-impaired healing of intestinal wounds. *Br J Surg* 1992;79:69-72.
- 33 Pierce GF, Mustoe TA, Lingelbach J, et al. Transforming growth factor beta reverses the glucocorticoid-induced wound-healing deficit in rats: Possible regulation in macrophages by platelet-derived growth factor. *Proc Natl Acad Sci USA* 1989;86:2229-33.
- 34 Beck LS, DeGuzman L, Lee WP, et al. One systemic administration of transforming growth factor-beta 1 reverses age- or glucocorticoid-impaired wound healing. *J Clin Invest* 1993;92:2841-9.
- 35 Xie C, Teixeira LR, Wang N, et al. Serial observations after high dose talc slurry in the rabbit model for pleurodesis. *Lung* 1998;176:299-307.
- 36 Xie C, McGovern JP, Wu W, et al. Comparisons of pleurodesis induced by talc with or without thymol iodide in rabbits. *Chest* 1998;113:795-9.
- 37 Vargas FS, Teixeira LR, Antonangelo L, et al. Acute and chronic pleural changes after the intrapleural instillation of mitoxantrone in rabbits. *Lung* 1998;176:227-36.