

Ca²⁺ homeostasis and structural and functional remodelling of airway smooth muscle in asthma

Katharina Mahn, Oluwaseun O Ojo, Gianna Chadwick, Philip I Aaronson, Jeremy P T Ward, Tak H Lee

King's College London, MRC and Asthma UK Centre in Allergic Mechanisms of Asthma, London, UK

Correspondence to

Professor Tak H Lee, King's College London, MRC and Asthma UK Centre in Allergic Mechanisms of Asthma, 5th Floor, Tower Wing, Guy's Hospital, London SE1 9RT, UK; tak.lee@kcl.ac.uk

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ABSTRACT

Asthma is characterised by airway hyper-responsiveness and remodelling, and there is mounting evidence that alterations in the phenotype of airway smooth muscle (ASM) play a central role in these processes. Although the concept that dysregulation of ASM Ca²⁺ homeostasis may underlie at least part of these alterations has been around for many years, it is only relatively recently that the availability of ASM biopsies from subjects with mild and moderate asthma has allowed it to be properly investigated. In this article, critical components of the pathobiology of asthmatic ASM, including contractile function, proliferation, cell migration and secretion of proinflammatory cytokines and chemokines, are reviewed and related to associated changes in ASM Ca²⁺ homeostasis. Based on this evidence, it is proposed that a unifying mechanism for the abnormal asthmatic phenotype is dysregulation of Ca²⁺ homeostasis caused at least in part by a downregulation in expression and function of sarcoendoplasmic Ca²⁺ ATPases (SERCAs).

Asthma is a chronic inflammatory disease which is characterised by widespread structural changes in the airways (airway remodelling), encompassing epithelial dysfunction, mucous gland hypertrophy, deposition of extracellular matrix (ECM) and changes in airway smooth muscle (ASM) phenotype.¹ ASM from patients with asthma exhibits enhanced contractile activity,^{2–4} as well as enhanced proliferation, migration and secretion of proinflammatory cytokines.^{5–8} This article reviews critical components of asthmatic ASM pathobiology and suggests that a unifying mechanism for the abnormal asthmatic phenotype is dysregulation of Ca²⁺ homeostasis caused at least in part by a downregulation in expression of sarcoendoplasmic Ca²⁺ ATPases (SERCAs).

CONTRACTILITY OF ASM IN ASTHMA

It is widely agreed that alterations in the contractility and dynamics of ASM contribute to airway narrowing and hyper-responsiveness in asthma. The contractile apparatus of ASM is composed of myosin and actin filaments (F-actin), and regulatory protein kinases and phosphatases that control their interaction. Myosin heavy chain (MHC) has ATPase activity, and binds to two light chains including the 20 kDa regulatory light chain (MLC₂₀). F-actin associates with regulatory proteins including tropomyosin, caldesmon and calponin, and promotes myosin assembly and ATPase activity. Contraction is normally

initiated by elevation of intracellular [Ca²⁺]_i ([Ca²⁺]_i), Ca²⁺-calmodulin-dependent activation of myosin light chain kinase (MLCK), and consequently phosphorylation of MLC₂₀ and myosin-actin cross-bridge cycling. MLC₂₀ is dephosphorylated by myosin light chain phosphatase (MLCP), which is regulated by Rho kinase, protein kinase C and telokin, so the net phosphorylation state of MLC₂₀ and therefore force production is primarily dependent on the relative activities of MLCK and MLCP⁹ (figure 1). However, actin-based mechanisms also play an important role through modulation of myosin ATPase activity and cytoskeletal dynamics.¹⁰ Pathways that initiate or modulate ASM contraction therefore include those that regulate [Ca²⁺]_i, Ca²⁺ sensitivity (via MLCP) and the actin cytoskeleton. Alterations to several such pathways have been implicated in asthma.

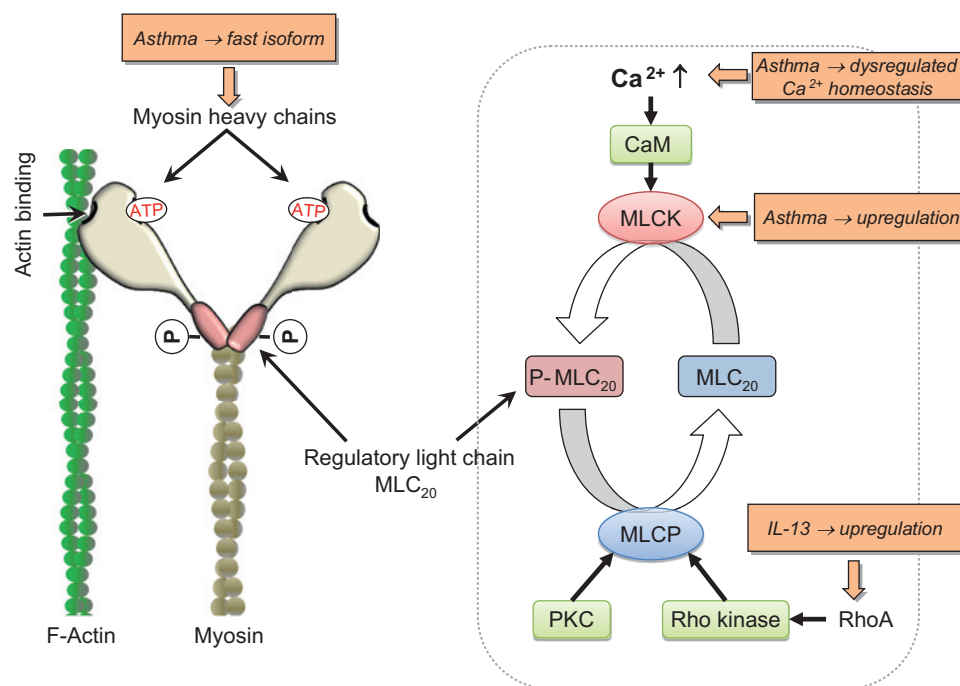
Increased expression of both a fast MHC isoform and MLCK has been reported in ASM from patients with asthma, which could increase the velocity of ASM shortening.^{2,4,11} Interestingly, Fairbank *et al*¹² have shown that repeated increases in ASM tone lead to increased MLCK expression, MLC₂₀ phosphorylation and changes in cytoskeletal organisation, suggesting that in asthma these could be secondary phenomena. Rho kinase has also been implicated in the enhanced response of ASM from subjects with asthma,³ though it is unclear whether its expression is altered. Notably, interleukin 13 (IL-13) is reported to increase expression of RhoA, a key activator of Rho kinase, in murine ASM.¹³

Cytosolic [Ca²⁺]_i is dependent on Ca²⁺ entry and removal across the cell membrane, and sequestration into and release from intracellular stores including the sarcoplasmic reticulum (SR) and mitochondria.¹⁴ The highly non-linear relationship between [Ca²⁺]_i and force in smooth muscle (due to 4:1 stoichiometry of Ca²⁺-calmodulin interaction) implies that even a small subthreshold increase in [Ca²⁺]_i could greatly increase subsequent responses to agonists. It was therefore suggested decades ago that the exaggerated contractile response of ASM typical of asthma could result from abnormal Ca²⁺ handling.¹⁵ This also has implications for mechanisms other than contraction.

PROLIFERATION AND MIGRATION OF ASM IN ASTHMA

ASM hyperplasia and proliferation may account for the increased ASM mass characteristic of asthma,⁵ and have been associated with increased ASM cell

Figure 1 Basic molecular mechanisms underlying force development in airway smooth muscle (ASM), and their regulation. Force is generated by ATP-dependent interaction between myosin and F-actin, regulated by phosphorylation of myosin 20 kDa regulatory light chain (MLC₂₀) by myosin light chain kinase (MLCK). A rise in $[Ca^{2+}]_i$ activates MLCK via Ca^{2+} –calmodulin (CaM). MLC₂₀ is dephosphorylated by myosin light chain phosphatase (MLCP), under the regulation of kinases including Rho kinase and protein kinase C (PKC). Highlighted boxes mark changes reported in asthma or following stimulation with interleukin 13 (IL-13). See text for details.



division, an altered ECM and reduced apoptosis.¹⁶ Cytokines, growth factors and ECM proteins induce ASM hyperplasia, and platelet-derived growth factor (PDGF), epidermal growth factor (EGF), T helper 2 (Th2) cytokines (IL-13 and IL-4), leukotriene D₄ (LTD₄) and transforming growth factor β (TGF β) all increase proliferation of ASM *in vitro*.¹⁶ All these mitogens appear to activate the GTPase p21Ras, which in turn activates phosphatidylinositol 3-kinase (PI3 kinase) or extracellular signal-regulated kinase (ERK); both of these regulate ASM proliferation through the cyclin D1 promoter.^{17–18} Glucocorticoids, a mainstay of asthma treatment, suppress proliferation in part via decreasing cyclin D1 expression,¹⁹ mediated by a complex of the glucocorticoid receptor and CCAAT/enhancer-binding protein α (C/EBP α). However, ASM from subjects with asthma exhibits a tissue-specific lack of C/EBP α , and this has been postulated to confer an ASM phenotype that is more susceptible to mitogenic and contractile stimuli.²⁰

Migration of proliferating ASM and precursor cells may contribute to increased ASM mass in asthma, and involves reorganisation of the actin cytoskeleton, assembly of focal contacts and myosin–actin-mediated force generation. Many factors integrate to promote actin disassembly, polymerisation and attachment to focal adhesions, and regulation of myosin activity; several are also involved in ASM growth (eg, PI3 kinase and ERK) and contraction (eg, MLCK, MLCP, RhoA and Rho kinase) (see review by Gerthoffer²¹). While the role of cell migration in airway remodelling remains unclear, ASM cells derived from subjects with asthma exhibit enhanced migration.^{6,8} Migration is strongly Ca^{2+} dependent, as myosin activity is regulated by Ca^{2+} –calmodulin-dependent activation of MLCK, and reorganisation of the actin cytoskeleton involves several Ca^{2+} -dependent mechanisms, notably the actin-severing protein gelsolin.²¹

SECRETION OF CYTOKINES AND CHEMOKINES IN ASTHMA

ASM is a rich source of chemokines and cytokines, including eotaxin, RANTES (regulated upon activation, normal T cell expressed and secreted), TARC (thymus and activation-regulated

chemokine), SCF (stem cell factor) and GM-CSF (granulocyte–macrophage colony-stimulating factor).²² For example, eotaxin release following stimulation of ASM with IL-13 or IL-4 mediates eosinophil recruitment,²³ and ASM-derived RANTES induces chemotaxis of inflammatory cells, thereby contributing to the inflammatory process.²⁴ Ca^{2+} signalling has an important role in release of cytokines and chemokines (eg, Denecker *et al*^{25–26}), and the finding that IL-13-induced eotaxin-1 secretion from healthy ASM is enhanced by knockdown of SERCA2 also implicates a key role for $[Ca^{2+}]_i$.⁸

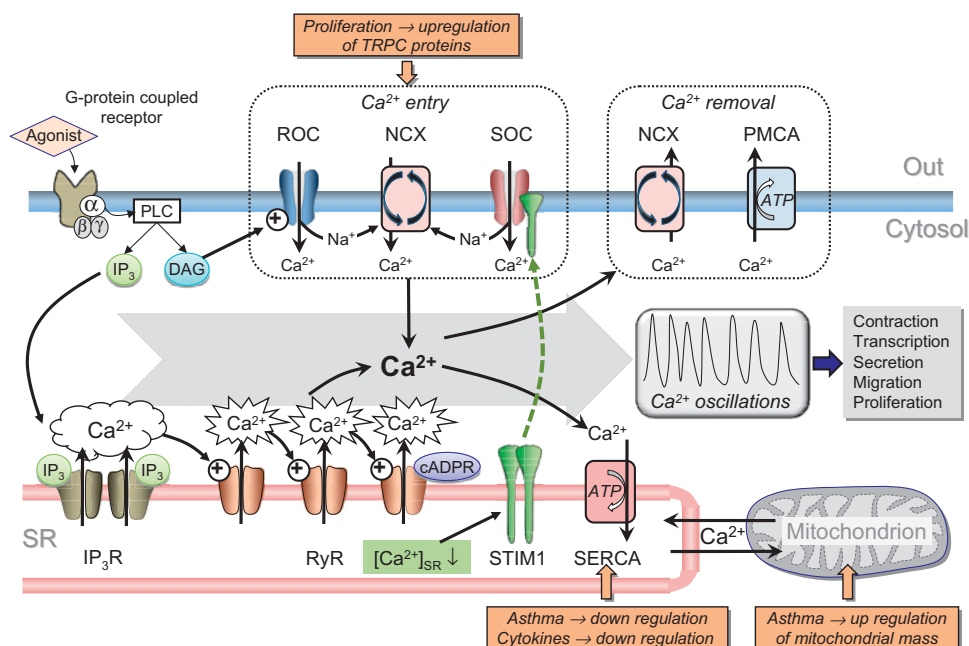
Ca^{2+} HOMEOSTASIS IN ASM

Intracellular Ca^{2+} has a central role in transduction of signals leading to diverse cellular responses, including contraction, metabolism, cell motility and protein secretion, as well as cell differentiation and proliferation.^{14–21–27} As such, it has long been suggested that alterations in ASM Ca^{2+} homeostasis may be central to the development of the asthmatic ASM phenotype and hyper-responsiveness.^{15–28–29} Elevation of ASM $[Ca^{2+}]_i$ can be elicited by activation of Ca^{2+} influx pathways and/or release from the SR. Ca^{2+} then binds to and stimulates high affinity binding proteins such as calmodulin, which activates MLCK,¹⁴ and the phosphatase calcineurin, which activates the transcription factor NFAT (nuclear factor of activated T cells) and indirectly NF- κ B (nuclear factor- κ B), both of which are implicated in inflammation and airway remodelling in asthma.^{30–31} $[Ca^{2+}]_i$ is pumped back into the SR or out of the cell in order to terminate its effects, the former mediated by the SERCA, and the latter by the plasma membrane Ca^{2+} ATPase (PMCA) and Na^+/Ca^{2+} exchanger (NCX)¹⁴ (figure 2).

Agonists binding to G_q/G₁₁ protein- or tyrosine kinase-coupled receptors can initiate elevation of $[Ca^{2+}]_i$ via phospholipase C (PLC)-mediated generation of diacylglycerol (DAG) and inositol trisphosphate (IP₃). DAG activates non-selective cation channels (permeable to both Na^+ and Ca^{2+}), often referred to as receptor-operated channels (ROCs)^{29–32}; these may be formed of TRPC6 (transient receptor potential channel 6) proteins which are expressed in human ASM.³³ IP₃ binds to clusters of receptors

Figure 2 Ca^{2+} mobilisation pathways in airway smooth muscle (ASM).

Binding of an agonist to a G-protein-coupled receptor activates phospholipase C (PLC) to generate diacylglycerol (DAG) and inositol trisphosphate (IP_3). DAG activates receptor-operated channels (ROCs), eliciting Na^+ and Ca^{2+} entry, while IP_3 activates clusters of IP_3 receptors (IP_3Rs) on the sarcoplasmic reticulum (SR), causing Ca^{2+} release which may activate adjacent ryanodine receptors (RyRs) to increase the frequency of sparks. RyRs may also be activated or potentiated by cADP ribose (cADPR). Store depletion is sensed by stromal-interacting molecule 1 (STIM1) within the SR, which translocates to and activates store-operated channels (SOCs). The elevation in subplasmalemmal $[\text{Na}^+]$ resulting from activation of non-selective ROCs or SOCs may be sufficient to drive reverse-mode operation of $\text{Na}^+/\text{Ca}^{2+}$ exchanger (NCX), leading to Ca^{2+} entry. Much of the Ca^{2+} entering the cell and released from stores may be sequestered by the superficial SR through sarcoendoplasmic Ca^{2+} ATPases (SERCAs) and mitochondria. Cycling between SR Ca^{2+} uptake and release mechanisms leads to Ca^{2+} oscillations. Ca^{2+} is removed from the cell by the plasma membrane Ca^{2+} ATPase (PMCA) and normal operation of the NCX. TRPC, transient receptor potential channel.



(IP_3Rs) in the SR to initiate Ca^{2+} release and highly localised elevations in $[\text{Ca}^{2+}]_i$ ('puffs'), which consequently activate ryanodine receptors (RyRs) in the SR (Ca^{2+} -induced Ca^{2+} release, CICR), thereby amplifying the elevation in cytosolic $[\text{Ca}^{2+}]_i$ and leading to Ca^{2+} 'sparks', and potentially regenerative Ca^{2+} waves and oscillations.¹⁴ RyRs are also activated by cADP ribose (cADPR), which is synthesised by the multifunctional enzyme CD38 (an ADP-ribosyl cyclase) and may contribute to generation of Ca^{2+} sparks and waves; it has been proposed that this pathway plays an important role in ASM.³⁴ It has been reported that some agonists such as LTD_4 do not elicit IP_3 -mediated Ca^{2+} release in human bronchioles, but do activate Ca^{2+} entry via ROCs.³²

Depletion of SR Ca^{2+} content by the above processes activates store-operated Ca^{2+} entry (SOCE) via the SR Ca^{2+} -sensing protein stromal-interacting molecule 1 (STIM1), which associates with the plasmalemmal protein Orai causing it to form channels which mediate SOCE.^{35–36} In smooth muscle many studies suggest that the channels underlying SOCE (store-operated channels; SOCs) are non-selective (ie, Na^+ and Ca^{2+} permeable), unlike classical STIM1–Orai which is Ca^{2+} selective. Possibly explaining this, there is evidence that the STIM1–Orai complex also incorporates TRPC proteins in smooth muscle, although this remains controversial.³⁷ As the degree of SR Ca^{2+} loading regulates SOCE, SERCA activity directly influences Ca^{2+} influx.

Removal of Ca^{2+} from the smooth muscle cell is handled by PMCA and NCX. PMCA is an ATP-dependent transporter activated by protein kinase G and A,¹⁴ providing one mechanism by which agents such as β_2 -adrenoceptor agonists cause ASM relaxation. NCX uses the Na^+ electrochemical gradient to pump out 1 Ca^{2+} in exchange for 3 Na^+ , and as such is dependent on membrane potential and cytosolic $[\text{Na}^+]$. Elevation of subsarcolemmal $[\text{Na}^+]$ can therefore cause reverse-mode operation of NCX and Ca^{2+} entry, and there is evidence that in ASM

agonist-induced Na^+ entry through non-selective cation channels (see above) does just this, with NCX then contributing to the elevation of $[\text{Ca}^{2+}]_i$ and store refilling.^{38–39}

The intracellular structure of ASM is highly organised, with peripheral SR, mitochondria and plasmalemma closely approximated in localised signalling microdomains. This provides a 'superficial buffer barrier', which buffers Ca^{2+} entry and limits its pervasion into the deeper cytosol, and through its highly regulated Ca^{2+} release and uptake mechanisms provides a level of precise control over the spatial and temporal characteristics of changes in $[\text{Ca}^{2+}]_i$.^{28–40} As such, it facilitates a high level of cross-talk between these organelles and consequently the mechanisms discussed above, and thus the ability to engender regenerating Ca^{2+} waves and oscillations. In ASM, agonists have indeed been shown to evoke Ca^{2+} waves or sustained oscillations of $[\text{Ca}^{2+}]_i$, caused by sequential Ca^{2+} release from and re-uptake into the SR, involving IP_3Rs , RyRs and SERCAs^{29–41–42} (figure 2). These waves seem to be responsible for acetylcholine-mediated contraction in human ASM, since their suppression by blockade of SERCAs or RyRs causes relaxation.⁴¹ Ca^{2+} entry is nevertheless important, as wave activity is terminated and contraction strongly suppressed by blockade of non-selective cation channels (ie, SOCs and ROCs) and reverse-mode NCX, although inhibition of L-type Ca^{2+} channels has only a minor effect.^{29–41} This implies that Ca^{2+} entry is primarily voltage independent and funnelled into the SR rather than serving to elevate $[\text{Ca}^{2+}]_i$ directly. Importantly, Ca^{2+} waves and oscillations also activate other Ca^{2+} -dependent processes, including gene transcription.^{29–43}

SERCAs clearly play a critical role in the above processes. There are three tissue-specific members of the mammalian SERCA family, SERCA1, SERCA2 and SERCA3, of which SERCA2 is expressed in muscle cells. Alternative splicing of exon 20 of the *ATP2A2* gene gives rise to SERCA2A, SERCA2B and SERCA2C mRNA transcripts, which differ in the final exon

usage.⁴⁴ We have recently demonstrated that the predominant isoform in ASM is SERCA2B, with the other splice variants expressed at very low levels.⁸ SERCAs are regulated by phospholamban, which under basal conditions acts to inhibit its activity. When phospholamban is phosphorylated, for example by protein kinase A, this inhibition is relieved and SR Ca^{2+} uptake dramatically increased.⁴⁵ This provides another important mechanism by which cAMP-raising agents induce bronchial relaxation.

ALTERED ASM Ca^{2+} HOMEOSTASIS IN ASTHMA: ROLE OF SERCAs

We have recently reported that SERCA2 protein expression in both native and cultured ASM from endobronchial biopsies of patients with mild and moderate/severe asthma is diminished compared with that of healthy subjects; the extent of this effect correlated with disease severity.⁸ Agonist-induced Ca^{2+} release from the SR was similarly depressed, consistent with a reduced store content, and the rate at which bradykinin-induced elevation of $[\text{Ca}^{2+}]_i$ recovered to baseline levels was significantly slowed (figure 3). In addition, ASM cells from subjects with asthma were significantly more active in a Ca^{2+} -dependent spreading assay that reflects migration. Importantly, suppression of SERCA2 by small interfering RNA (siRNA) in ASM cells derived from donors without asthma recapitulated the asthmatic phenotype, with similarly slowed recovery of $[\text{Ca}^{2+}]_i$ to baseline and increased rates of proliferation and cell spreading. Moreover, IL-13-induced eotaxin release, which is greatly enhanced in cultured ASM cells from subjects with moderate/severe asthma, was also increased by siRNA-induced SERCA2 knockdown.⁸

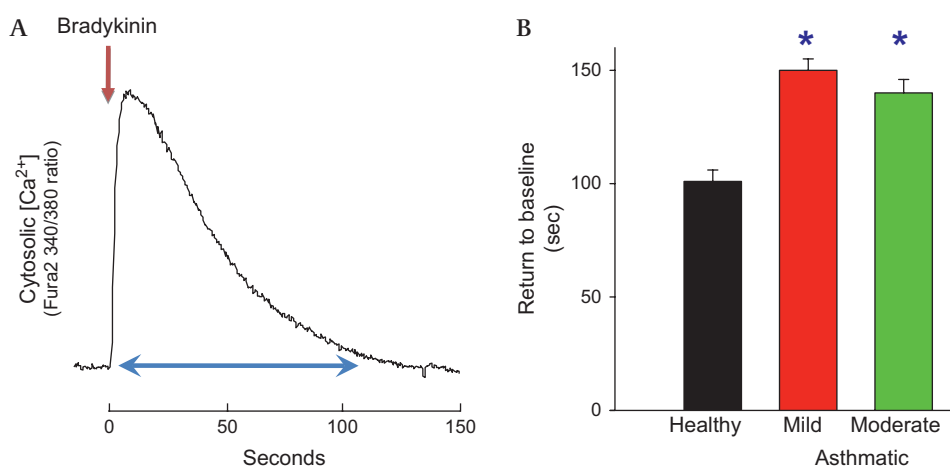
The above strongly suggests that downregulation of SERCA2 plays an important, if not critical role in the dysregulation of ASM Ca^{2+} homeostasis in asthma and the consequent alterations in phenotype. Importantly, Sathish *et al*⁴⁶ have now reported that treatment of healthy ASM with the proinflammatory cytokines IL-13 and tumour necrosis factor α (TNF α) causes suppression of SERCA2 expression and function, thereby providing a putative link between airway inflammation and reduced ASM SERCA2 expression in asthma. Interestingly, IL-13- and TNF α -induced hyper-responsiveness has also been associated with altered SR Ca^{2+} handling via the CD38/cADPR pathway.^{47 48}

It is noteworthy that a fall in expression of SERCA2A/2B occurs in Darier disease, an autosomal dominant skin condition

in which only one copy of the SERCA2 gene (*ATP2A2*) is functional. This disease is characterised by lesions demonstrating abnormal keratinisation and loss of epidermal cellular adhesion,⁴⁹ though no association with asthma has been reported. Similarly *ATP2A2* heterozygous knockout mice eventually develop hyperkeratinised squamous cell tumours.⁵⁰ These mice also show a deficient cardiac relaxation, and it is known that SERCA2 expression is reduced in human cardiac failure and hypertrophy⁵¹; notably, SERCA2 gene transfer into the myocardium is currently undergoing clinical trials for treatment of this condition.

It is not yet known whether the decrease in ASM SERCA2 expression in asthma could have a genetic basis, in which case it might be expected that downregulation of SERCA2 would be observed in other types of cell. Although asthma has not been associated with cardiac dysfunction, which might be predicted if SERCA2 expression is also reduced in cardiac myocytes, patients with Darier disease also do not exhibit any abnormalities of cardiac function.⁵² Thus even this substantial genetically determined deficit in SERCA2 can cause abnormalities which manifest only in certain types of cell. It may therefore be that mutations in the *ATP2A2* gene, or in genes which affect its expression, exist in those with asthma, but for some reason exert restricted effects. This would be analogous to the situation in familial pulmonary arterial hypertension, which is associated with the decreased expression of the BMPR-II receptor caused by autosomal dominant mutations in the *BMPR2* gene.⁵³ However, this mutation has a penetrance of only ~20% and, even though this receptor is functionally important in other organs, causes a lung-specific pathology. This implies that the disease only manifests itself when other factors are present ('second hit' hypothesis). It is, for example, feasible that a localised functional suppression of SERCA2 activity may also occur in the powerfully oxidising and cytokine-rich milieu of the inflamed asthmatic lung⁵⁴ which could constitute a focal 'second hit' sufficient to cause a significant effect on cellular Ca^{2+} handling restricted mainly to these cells. Interestingly, it has recently been reported that SERCA function is inhibited by ORMDL3, which is implicated in the inflammation-promoting unfolded protein response and has been identified as a genetic risk factor for childhood asthma.⁵⁵ This is of clear relevance to whether the observed reduction in ASM SERCA2 expression is of genetic origin or secondary to an increase in proinflammatory cytokines.⁴⁶ It is noteworthy that the disease severity-related fall in SERCA2B expression persisted in ASM cells through multiple

Figure 3 Recovery of Ca^{2+} transient is delayed in airway smooth muscle (ASM) cells from subjects with asthma. (A) An example trace of a Ca^{2+} transient elicited by 1 μM bradykinin in ASM cells cultured from a healthy subject, measured using Fura2. The time taken for the increase in Ca^{2+} to return to baseline is indicated. (B) Mean data obtained from ASM cells derived from healthy subjects and those with mild and moderate asthma (data from Mahn *et al*⁸). * $p < 0.05$.



passages in culture,⁸ suggesting either a genetic or an epigenetic origin.

Downregulation of SERCA2 function has previously been reported to promote proliferation in a number of tissues including other smooth muscles, possibly mediated via calcineurin.²⁷ It is noteworthy in this regard that ciclosporin, a calcineurin antagonist, has been used as a treatment for steroid-resistant asthma.⁵⁶ Trian *et al*⁵⁷ have proposed that the enhanced proliferation in ASM from subjects with asthma is due to an increase in mitochondrial mass, driven by upregulation of mitochondrial transcription factors as a consequence of altered ASM Ca^{2+} homeostasis. While it is unclear how increased mitochondria might promote proliferation, this mechanism could be an important downstream effector pathway for SERCA dysfunction, although Trian *et al*⁵⁷ advocated an enhanced Ca^{2+} entry pathway. Notably, a variety of Ca^{2+} influx pathways are also upregulated when SERCA expression or function is suppressed, for example NCX and TRPC proteins in cardiomyocytes, again associated with an increase in transcript levels of a calcineurin-dependent transcription factor complex and activity of NFAT.^{58–59} It is therefore interesting that ASM proliferation has been associated with a marked upregulation of TRPCs.⁶⁰

As SERCA2 plays a key role in the operation of the superficial buffer barrier and generation of Ca^{2+} oscillations,^{14 28 40} it can be predicted that reduced expression and/or function of SERCA2 would result in not only increased basal levels of $[\text{Ca}^{2+}]_i$, but also potentiation of Ca^{2+} influx-induced elevations of $[\text{Ca}^{2+}]_i$ and altered dynamics of Ca^{2+} oscillations. Consistent with this, an increased resting $[\text{Ca}^{2+}]_i$ and slowing of Ca^{2+} oscillations has been observed in ASM under conditions where SERCA activity is reduced.⁴⁶ This could underpin, at least in part, both ASM hyper-responsiveness to agonists and, through alterations to the frequency of Ca^{2+} oscillations, differential activation of transcription factors such as NFAT and NF- κB .⁴³

CONCLUSION

It is clear that Ca^{2+} signalling plays a central and critical role in most if not all of the cellular processes that are known to be altered in ASM from subjects with asthma, including contractility, responsiveness to agonists, proliferation, migration and secretion of inflammatory mediators. We therefore propose that a unifying mechanism for the abnormal asthmatic phenotype is dysregulation of Ca^{2+} homeostasis, mediated at least in part by downregulation of SERCA2 expression and function driven by proinflammatory mediators. Many questions remain unanswered, including whether the reduction in SERCA2 expression is induced or of genetic origin, and whether observed changes in other Ca^{2+} homeostatic mechanisms occur independently or are secondary to reduced SERCA2 function. However, the fact that experimental suppression of SERCA2 expression in healthy ASM appears to recapitulate key aspects of the asthmatic phenotype⁸ strongly suggests that SERCA2 downregulation plays a central role in the altered function of asthmatic ASM.

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REFERENCES

- Hirst SJ. Airway smooth muscle as a target in asthma. *Clin Exp Allergy* 2000;**30** (Suppl 1):54–9.
- Ma X, Cheng Z, Kong H, *et al*. Changes in biophysical and biochemical properties of single bronchial smooth muscle cells from asthmatic subjects. *Am J Physiol Lung Cell Mol Physiol* 2002;**283**:L1181–9.
- Matsumoto H, Moir LM, Oliver BG, *et al*. Comparison of gel contraction mediated by airway smooth muscle cells from patients with and without asthma. *Thorax* 2007;**62**:848–54.
- Leguillet R, Lavolette M, Bergeron C, *et al*. Myosin, transgelin, and myosin light chain kinase: expression and function in asthma. *Am J Respir Crit Care Med* 2009;**179**:194–204.
- Johnson PR, Roth M, Tamm M, *et al*. Airway smooth muscle cell proliferation is increased in asthma. *Am J Respir Crit Care Med* 2001;**164**:474–7.
- Kaur D, Saunders R, Berger P, *et al*. Airway smooth muscle and mast cell-derived CC chemokine ligand 19 mediate airway smooth muscle migration in asthma. *Am J Respir Crit Care Med* 2006;**174**:1179–88.
- Chan V, Burgess JK, Ratoff JC, *et al*. Extracellular matrix regulates enhanced eotaxin expression in asthmatic airway smooth muscle cells. *Am J Respir Crit Care Med* 2006;**174**:379–85.
- Mahn K, Hirst SJ, Ying S, *et al*. Diminished sarco/endoplasmic reticulum Ca^{2+} -ATPase (SERCA) expression contributes to airway remodelling in bronchial asthma. *Proc Natl Acad Sci USA* 2009;**106**:10775–80.
- Somlyo AP, Somlyo AV. Ca^{2+} sensitivity of smooth muscle and nonmuscle myosin II: modulated by G proteins, kinases, and myosin phosphatase. *Physiol Rev* 2003;**83**:1325–58.
- Gunst SJ, Zhang W. Actin cytoskeletal dynamics in smooth muscle: a new paradigm for the regulation of smooth muscle contraction. *Am J Physiol Cell Physiol* 2008;**295**:C576–87.
- Benayoun L, Druilhe A, Dombret MC, *et al*. Airway structural alterations selectively associated with severe asthma. *Am J Respir Crit Care Med* 2003;**167**:1360–8.
- Fairbank NJ, Connolly SC, Mackinnon JD, *et al*. Airway smooth muscle cell tone amplifies contractile function in the presence of chronic cyclic strain. *Am J Physiol Lung Cell Mol Physiol* 2008;**295**:L479–88.
- Chiba Y, Nakazawa S, Todoroki M, *et al*. Interleukin-13 augments bronchial smooth muscle contractility with an up-regulation of RhoA protein. *Am J Respir Cell Mol Biol* 2009;**40**:159–67.
- Berridge MJ. Smooth muscle cell calcium activation mechanisms. *J Physiol* 2008;**586**:5047–61.
- Triggle DJ. Calcium, the control of smooth muscle function and bronchial hyperreactivity. *Allergy* 1983;**38**:1–9.
- Hirst SJ, Martin JG, Bonacci JV, *et al*. Proliferative aspects of airway smooth muscle. *J Allergy Clin Immunol* 2004;**114**(Suppl 2):S2–17.
- Ramakrishnan M, Musa NL, Li J, *et al*. Catalytic activation of extracellular signal-regulated kinases induces cyclin D1 expression in primary tracheal myocytes. *Am J Respir Cell Mol Biol* 1998;**18**:736–40.
- Page K, Li J, Wang Y, *et al*. Regulation of cyclin D(1) expression and DNA synthesis by phosphatidylinositol 3-kinase in airway smooth muscle cells. *Am J Respir Cell Mol Biol* 2000;**23**:436–43.
- Fernandes D, Guida E, Koutsoubos V, *et al*. Glucocorticoids inhibit proliferation, cyclin D1 expression, and retinoblastoma protein phosphorylation, but not activity of the extracellular-regulated kinases in human cultured airway smooth muscle. *Am J Respir Cell Mol Biol* 1999;**21**:77–88.
- Borger P, Tamm M, Black JL, *et al*. Asthma: is it due to an abnormal airway smooth muscle cell? *Am J Respir Crit Care Med* 2006;**174**:367–72.
- Gerthoffer WT. Migration of airway smooth muscle cells. *Proc Am Thorac Soc* 2008;**5**:97–105.
- Tiliba O, Panettieri RA Jr. Noncontractile functions of airway smooth muscle cells in asthma. *Annu Rev Physiol* 2009;**71**:509–35.
- Zuyderduyn S, Hiemstra PS, Rabe KF. TGF-beta differentially regulates TH2 cytokine-induced eotaxin and eotaxin-3 release by human airway smooth muscle cells. *J Allergy Clin Immunol* 2004;**114**:791–8.
- Hershenson MB, Brown M, Camoretti-Mercado B, *et al*. Airway smooth muscle in asthma. *Annu Rev Pathol* 2008;**3**:523–55.
- Denecker NE, Kullberg BJ, Drenth JP, *et al*. Regulation of the production of pro-inflammatory cytokines and antagonists during chemotherapy-induced neutropenia in patients with haematological malignancies. *Cytokine* 1997;**9**:702–10.
- Huang CD, Ammit AJ, Tiliba O, *et al*. G-protein-coupled receptor agonists differentially regulate basal or tumor necrosis factor-alpha-stimulated activation of interleukin-6 and RANTES in human airway smooth muscle cells. *J Biomed Sci* 2005;**12**:763–76.
- Lipskaia L, Hulot JS, Lompre AM. Role of sarco/endoplasmic reticulum calcium content and calcium ATPase activity in the control of cell growth and proliferation. *Pflügers Arch* 2009;**457**:673–85.
- Parameswaran K, Janssen LJ, O'Byrne PM. Airway hyperresponsiveness and calcium handling by smooth muscle: a "deeper look". *Chest* 2002;**121**:621–4.
- Perez-Zoghbi JF, Karner C, Ito S, *et al*. Ion channel regulation of intracellular calcium and airway smooth muscle function. *Pulm Pharmacol Ther* 2009;**22**:388–97.
- Crabtree GR, Olson EN. NFAT signaling: choreographing the social lives of cells. *Cell* 2002(Suppl 109):S67–79.

31. **Poynter ME**, Cloots R, van Woerkom T, *et al*. NF-kappa B activation in airways modulates allergic inflammation but not hyperresponsiveness. *J Immunol* 2004;**173**:7003–9.
32. **Snetkov VA**, Hapgood KJ, McVicker CG, *et al*. Mechanisms of leukotriene D₄-induced constriction in human small bronchioles. *Br J Pharmacol* 2001;**133**:243–52.
33. **Corteling RL**, Li S, Giddings J, *et al*. Expression of transient receptor potential C6 and related transient receptor potential family members in human airway smooth muscle and lung tissue. *Am J Respir Cell Mol Biol* 2004;**30**:145–54.
34. **Deshpande DA**, White TA, Dogan S, *et al*. CD38/cyclic ADP-ribose signaling: role in the regulation of calcium homeostasis in airway smooth muscle. *Am J Physiol Lung Cell Mol Physiol* 2005;**288**:L773–88.
35. **Hewavitharana T**, Deng X, Soboloff J, *et al*. Role of STIM and Orai proteins in the store-operated calcium signaling pathway. *Cell Calcium* 2007;**42**:173–82.
36. **Peel SE**, Liu B, Hall IP. Orai and store-operated calcium influx in human airway smooth muscle cells. *Am J Respir Cell Mol Biol* 2008;**38**:744–9.
37. **Ong HL**, Cheng KT, Liu X, *et al*. Dynamic assembly of TRPC1–STIM1–Orai1 ternary complex is involved in store-operated calcium influx. Evidence for similarities in store-operated and calcium release-activated calcium channel components. *J Biol Chem* 2007;**282**:9105–16.
38. **Algara-Suarez P**, Romero-Mendez C, Chrones T, *et al*. Functional coupling between the Na⁺/Ca²⁺ exchanger and nonselective cation channels during histamine stimulation in guinea pig tracheal smooth muscle. *Am J Physiol Lung Cell Mol Physiol* 2007;**293**:L191–8.
39. **Hirota S**, Pertens E, Janssen LJ. The reverse mode of the Na⁺/Ca²⁺ exchanger provides a source of Ca²⁺ for store refilling following agonist-induced Ca²⁺ mobilisation. *Am J Physiol Lung Cell Mol Physiol* 2007;**292**:L438–47.
40. **van Breemen C**, Chen Q, Laher I. Superficial buffer barrier function of smooth muscle sarcoplasmic reticulum. *Trends Pharmacol Sci* 1995;**16**:98–105.
41. **Dai JM**, Kuo KH, Leo JM, *et al*. Acetylcholine-induced asynchronous calcium waves in intact human bronchial muscle bundle. *Am J Respir Cell Mol Biol* 2007;**36**:600–8.
42. **Pabelick CM**, Sieck GC, Prakash YS. Invited review: significance of spatial and temporal heterogeneity of calcium transients in smooth muscle. *J Appl Physiol* 2001;**91**:488–96.
43. **Dolmetsch RE**, Xu K, Lewis RS. Calcium oscillations increase the efficiency and specificity of gene expression. *Nature* 1998;**392**:933–6.
44. **Hauser K**, Pavlovic N, Kissmehl R, *et al*. Molecular characterization of a sarco(endo)plasmic reticulum Ca²⁺-ATPase gene from *Paramecium tetraurelia* and localization of its gene product to sub-plasmalemmal calcium stores. *Biochem J* 1998;**334**:31–8.
45. **East JM**. Sarco(endo)plasmic reticulum calcium pumps: recent advances in our understanding of structure/function and biology (review). *Mol Membr Biol* 2000;**17**:189–200.
46. **Sathish V**, Thompson MA, Bailey JP, *et al*. Effect of proinflammatory cytokines on regulation of sarcoplasmic reticulum Ca²⁺ reuptake in human airway smooth muscle. *Am J Physiol Lung Cell Mol Physiol* 2009;**297**:L26–34.
47. **Deshpande DA**, Dogan S, Walseth TF, *et al*. Modulation of calcium signaling by interleukin-13 in human airway smooth muscle: role of CD38/cyclic adenosine diphosphate ribose pathway. *Am J Respir Cell Mol Biol* 2004;**31**:36–42.
48. **Guedes AG**, Jude JA, Paulin J, *et al*. Role of CD38 in TNF-alpha-induced airway hyperresponsiveness. *Am J Physiol Lung Cell Mol Physiol* 2008;**294**:L290–9.
49. **Sakuntabhai A**, Ruiz-Perez V, Carter S, *et al*. Mutations in ATP2A2, encoding a Ca²⁺ pump, cause Darier disease. *Nat Genet* 1999;**21**:271–7.
50. **Liu LH**, Boivin GP, Prasad V, *et al*. Squamous cell tumors in mice heterozygous for a null allele of Atp2a2, encoding the sarco(endo)plasmic reticulum Ca²⁺-ATPase isoform 2 Ca²⁺ pump. *J Biol Chem* 2001;**276**:26737–40.
51. **Meyer M**, Schillinger W, Pieske B, *et al*. Alterations of sarcoplasmic reticulum proteins in failing human dilated cardiomyopathy. *Circulation* 1995;**92**:778–84.
52. **Mayosi BM**, Kardos A, Davies CH, *et al*. Heterozygous disruption of SERCA2a is not associated with impairment of cardiac performance in humans: implications for SERCA2a as a therapeutic target in heart failure. *Heart* 2006;**92**:105–9.
53. **Upton PD**, Morrell NW. TGF-beta and BMPR-II pharmacology—implications for pulmonary vascular diseases. *Curr Opin Pharmacol* 2009;**9**:274–80.
54. **Gutierrez-Martin Y**, Martin-Romero FJ, Inesta-Vaquera FA, *et al*. Modulation of sarcoplasmic reticulum Ca²⁺-ATPase by chronic and acute exposure to peroxynitrite. *Eur J Biochem* 2004;**271**:2647–57.
55. **Cantero-Recasens G**, Fandos C, Rubio-Moscardo F, *et al*. The asthma-associated ORMDL3 gene product regulates endoplasmic reticulum-mediated calcium signaling and cellular stress. *Hum Mol Genet* 2010;**19**:111–21.
56. **Frew AJ**, Plummeridge MJ. Alternative agents in asthma. *J Allergy Clin Immunol* 2001;**108**:3–10.
57. **Triana T**, Benard G, Begueret H, *et al*. Bronchial smooth muscle remodeling involves calcium-dependent enhanced mitochondrial biogenesis in asthma. *J Exp Med* 2007;**204**:3173–81.
58. **Brittsan AG**, Ginsburg KS, Chu G, *et al*. Chronic SR Ca²⁺-ATPase inhibition causes adaptive changes in cellular Ca²⁺ transport. *Circ Res* 2003;**92**:769–76.
59. **Seth M**, Sumbilla C, Mullen SP, *et al*. Sarco(endo)plasmic reticulum Ca²⁺-ATPase (SERCA) gene silencing and remodeling of the Ca²⁺ signaling mechanism in cardiac myocytes. *Proc Natl Acad Sci USA* 2004;**101**:16683–8.
60. **Sweeney M**, Yu Y, Platoshyn O, *et al*. Inhibition of endogenous TRP1 decreases capacitative Ca²⁺ entry and attenuates pulmonary artery smooth muscle cell proliferation. *Am J Physiol Lung Cell Mol Physiol* 2002;**283**:L144–55.